Environmental Chamber for In Situ Dynamic Control of Temperature and Relative Humidity During X-Ray Scattering

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Salas-de la Cruz, D., Denis, J. G., Griffith, M. D., King, D. R., Heiney, P. A., & Winey, K. I. (2012). Environmental Chamber for In Situ Dynamic Control of Temperature and Relative Humidity During X-Ray Scattering. Retrieved from [http://repository.upenn.edu/physics_papers/229](http://repository.upenn.edu/physics_papers/229)


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Abstract
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Disciplines
Physical Sciences and Mathematics | Physics

Comments

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Environmental chamber for in situ dynamic control of temperature and relative humidity during x-ray scattering

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(Received 30 September 2011; accepted 31 January 2012; published online 21 February 2012)

We have designed, constructed, and evaluated an environmental chamber that has in situ dynamic control of temperature (25 to 90 °C) and relative humidity (0% to 95%). The compact specimen chamber is designed for x-ray scattering in transmission with an escape angle of 2θ = ±30°. The specimen chamber is compatible with a completely evacuated system such as the Rigaku PSAXS system, in which the specimen chamber is placed inside a larger evacuated chamber (flight path). It is also compatible with x-ray systems consisting of evacuated flight tubes separated by small air gaps for sample placement. When attached to a linear motor (vertical displacement), the environmental chamber can access multiple sample positions. The temperature and relative humidity inside the specimen chamber are controlled by passing a mixture of dry and saturated gas through the chamber and by heating the chamber walls. Alternatively, the chamber can be used to control the gaseous environment without humidity. To illustrate the value of this apparatus, we have probed morphology transformations in Nafion® membranes and a polymerized ionic liquid as a function of relative humidity in nitrogen.

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I. INTRODUCTION

The environmental chamber described in this paper for transmission x-ray scattering can be used to characterize materials that have scientifically interesting structural transformations as a function of temperature, relative humidity, non-corrosive gases, or mixtures of gases. This device allows materials to be studied under conditions that mimic various applications and scenarios including: membranes for fuel cells,1–5 membranes for actuators,6,7 materials in corrosive environments (simulated aging),8,9 naturally occurring materials,10 biomimetic materials,11 and catalyst materials affectivity.12 For instance, Smith et al.13 demonstrated a correlation between chemical potential of water and morphology by performing x-ray scattering studies of oriented dimyristoylphosphatidylcholine-water lamellar films using an in situ triple axis x-ray spectrometer at a controlled temperature and relative humidity. Comparatively, Halim and Scherer14 were able to correlate and understand the electrochemical and morphological properties of Nafion® film as a function of relative humidity using small- and wide-angle x-ray scattering under different humidities. Finally, Bang et al.15 investigated the effect of humidity on the ordering of poly(ethylene oxide)-based copolymer thin films where it was shown that by annealing a poly(ethylene oxide-b-methylmethacrylate-b-styrene)triblock copolymer under high humidity the lamellar morphology in the bulk developed a hexagonal array of poly(ethylene oxide) microdomains.

Environmental chambers have been previously designed for a range of specific x-ray scattering or neutron scattering experiments. For example, Kim et al.16 designed an in situ vapor sorption apparatus, for small-angle neutron scattering, with a temperature range from 0 to 150 °C and a relative humidity range from 0% to 90%. The % RH is controlled by measuring the absolute pressure and the apparatus has the capability to inject a pore-masking liquid in the sample cell. In contrast, Smith et al.13,17 developed a humidity oven for neutron and synchrotron x-ray scattering which has a temperature range from 0 to 80 °C and a relative humidity range from 0% to 100% RH. Katsaras and Watson18 assembled a sample cell capable of attaining 100% RH for a synchrotron x-ray diffraction system. The sample cell has a temperature range from 5 to 80 °C and the humidity is controlled by a porous membrane located near the sample. Giahi et al.19 developed an alternative design using two humidity chambers where the humidity was controlled by an internal water/salt mixture. The chambers allow experiments in aqueous solutions and are fully compatible with x-ray scattering, x-ray non-specular (diffuse) scattering and grazing incidence scattering.

We report a new design for an environmental chamber (EC) that controls temperature and relative humidity during transmission scattering. Key features of this new design include a large escape angle, compatibility with an evacuated system such as the Rigaku PSAXS system, a sample mount for multiple specimens, dynamic control of temperature and relative humidity, and integrated computer control for both chamber conditions and data collection. The EC can also be used to control the gaseous environment during scattering experiments to probe chemical reactions. This in situ apparatus will enable advances in the fundamental understanding of the
TABLE I. Environmental chamber (EC) descriptions and capabilities.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type of radiation</td>
<td>X-ray</td>
</tr>
<tr>
<td>Geometry</td>
<td>Transmission</td>
</tr>
<tr>
<td>Escape angle, 2θ (horizontally)</td>
<td>±30°</td>
</tr>
<tr>
<td>Temperature range</td>
<td>25–90°C</td>
</tr>
<tr>
<td>Temperature precision</td>
<td>0.05–0.5°C</td>
</tr>
<tr>
<td>Temperature gradient across sample</td>
<td>&lt;1°C</td>
</tr>
<tr>
<td>Humidity range</td>
<td>0%–95%</td>
</tr>
<tr>
<td>Humidity precision</td>
<td>1.5% RH at 30°C</td>
</tr>
<tr>
<td></td>
<td>3% RH at 80°C</td>
</tr>
</tbody>
</table>

relationship between morphology, temperature, relative humidity, and gaseous environments. To illustrate the value of this apparatus, transient morphologies in Nafion® membranes (E. I. DuPont de Nemours and Company) and imidazolium-based polymerized ionic liquids have been investigated as a function of relative humidity.

II. INSTRUMENT DESIGN

A. Overall description

The EC includes a specimen chamber, a recirculating temperature-controlled water bath, a humidity generation system, and a proportional-integral-derivative (PID) control system. Table I shows the equipment specifications and brief descriptions. Temperature and relative humidity inside the specimen chamber are controlled primarily by mixing dry and saturated gas, wherein the saturated gas is generated by bubbling gas through temperature-controlled water reservoir. In addition, the temperature of the chamber walls is controlled by a double pass heat exchanger using water as the cooling and heating fluid.

The temperature and humidity inside the specimen chamber are recorded using an electric temperature/hydrometer from Rotronic® (Rotronic USA). The dry and saturated flows are controlled by two mass flow controllers from MKS Instruments. The temperature indicators, temperature-controllers, and flow meters are monitored and controlled via a custom Labview® 8.5 program (National Instruments). The software incorporates a PID sequence to control the temperature and relative humidity inside the specimen chamber.

B. Specimen chamber

To study transient morphologies as a function of relative humidity or non-corrosive gases, we designed the specimen chamber to have a large escape angle to maximize the range of accessible scattering angles. In the horizontal direction the maximum escape angle is \(2\theta = \pm 30^\circ\), while it is even larger in the vertical direction. The specimen chamber prevents condensation inside the chamber and the compact size minimizes x-ray absorption. A novel attribute of our specimen chamber is the capability to mount multiple samples for increased throughput and for environmental conformity for all samples. The specimen chamber (Figure 1) includes the following components: (1) RH chamber, (2) sample mount, (3) door, (4) sample clamp, (5) window frames, (6) y-direction motor mount for vertical displacement, (7) insulator, (8) strain relief, (9) insulator for strain relief, (10) cooling loops, and (11) beryllium (Be) window. Key features of this new design include large escape angle (x-ray scattering angle \(2\theta = \pm 30^\circ\), horizontal), compatibility with an evacuated x-ray chamber, capability to hold multiple samples, dynamic control of temperature and relative humidity, and computer control of temperature and relative humidity integrated with data collection.

FIG. 1. (Color online) Specimen chamber 3D illustration. The specimen chamber includes the following components: (1) RH chamber, (2) sample mount, (3) door, (4) sample clamp, (5) window frames, (6) y-direction motor mount for vertical displacement, (7) insulator, (8) strain relief, (9) insulator for strain relief, (10) cooling loops, and (11) beryllium (Be) window. The outer dimensions are 4.5 in. \(\times\) 1.5 in. \(\times\) 3.4 in. and the inner dimensions are 3.55 in. \(\times\) 0.75 in. \(\times\) 2.9 in. The chamber is made from stainless steel 309. The sample mount is removable to facilitate sample changes. Each sample mount holds up to five individual samples (~0.50 in. \(\times\) 0.25 in.) using sample clamps (Figure 1). In essence, the sample mount allows the transmission of x-rays and is connected to a removable door via four bolts.

To control the temperature and relative humidity of the sample environment, the specimen chamber has inlet and outlet connections to enable the flow of dry or moist
temperature-controlled gas. The design of the specimen chamber allows good mixing and ample vapor exchange inside the specimen chamber. An internal electric temperature/hydrometer sensor probes the atmosphere within the sample chamber. This sensor measures from −40 to 100 °C with an accuracy of ±0.1 °C and relative humidities from 0% to 100% with an accuracy of ±1.5% RH. The sensor position can be changed in the y-direction within the sample chamber using a robust leak-tight connection. The removable door has a thermocouple located in the outer wall to record the temperature of the door and sample mount that serves as a secondary measure of the sample temperature in contact with the sample mount.

The specimen chamber includes two beryllium (Be) windows to minimize x-ray absorption. The Be windows (3.5 in. × 1.5 in. × 0.025 in.) provide a maximum horizontal exit angle of ±30° to allow a wide collection angle for the x-ray scattering experiments and to access the multiple sample positions. The x-ray path length between the Be windows is only 0.75 in. The Be windows are sealed to the chamber using a paste adhesive and further secured by window frames. The inner side of the Be windows is coated with a corrosion inhibitor due to the moist environment in the specimen chamber. The Be windows have a wall thickness of 0.025 in. (635 μm). The thickness of the beryllium windows and the window frames provide a robust design to accommodate the pressure difference between the inside of the sample chamber (1 atm = 1013 mbar) and the evacuated flight path chamber (0.1 mbar).

The walls of the specimen chamber include a double pass heat exchanger copper tube and a hole for a temperature sensor to measure the temperature of the chamber wall. The double pass heat exchanger controls the temperature of the specimen chamber walls using an external temperature-controlled water recirculation bath. The wall temperature is set to 3 °C warmer than the chamber atmosphere to prevent condensation on the walls and Be windows. The specimen chamber is connected to a linear motor mount, which controls the position in the y-direction (Figure 1). To prevent thermal loss due to conduction of heat to the motor mount, the specimen chamber is thermally isolated using an insulating spacer.

C. Recirculating temperature-controlled water bath

The wall temperature of the specimen chamber is controlled via a recirculation water bath using predetermined calibration curves. The walls of the specimen chamber incorporate copper tubing through which the water passes before returning to the temperature-controlled water bath. The 7 l water recirculation bath temperature control range of −25 to 150 °C with a stability of ± 0.01 °C. The water recirculation bath temperature is regulated using a PID controller, which is capable of cooling at 500 W and heating at 800 W. The pressure pump has a maximum flow of 15 l/min at a maximum pressure of 5 psi. The inlet and outlet lines are connected to the specimen chamber through the wall of the evacuated flight path by two feed-throughs. The inlet and outlet lines inside the evacuated flight path are insulated to prevent heat losses to the environment.

D. Humidity generator

The temperature and relative humidity in the specimen chamber are controlled by flowing through the chamber warm, moist gas (typically nitrogen) from an external humidity generator. Saturated gas is generated when dry gas passes through a bottle filled with water at a designated temperature, the bottle temperature. The humidity generator includes the following: an immersion heater, two insulated Teflon bottles, a dry gas flow meter, a wet gas flow meter, a compressed gas line, a mixing line, and two heat traces.

The bottle incorporates two one-directional diffusers that prevent condensation and liquid water from entering the specimen chamber. The saturated gas flow is mixed with dry gas flow to achieve the desired relative humidity in the specimen chamber. The percent relative humidity (%RH) is defined as the partial pressure of water vapor in the gas divided by the vapor pressure of water at a given temperature. Assuming that both the dry and saturated gases behave as ideal gases, the relative humidity can be expressed as

\[
\text{RH}(\%) = \frac{p_{\text{H}_2\text{O}}^*}{p_{\text{H}_2\text{O}}^*} \times 100 = \frac{x_{\text{H}_2\text{O}}}{x_{\text{H}_2\text{O}}} \times 100
\]

where \( p_{\text{H}_2\text{O}}^* \) is the desired partial pressure of water vapor, \( x_{\text{H}_2\text{O}} \) is the saturated vapor pressure, \( x_{\text{H}_2\text{O}}^* \) is the mole fraction of water, \( F_1 \) is the flow rate of saturated gas escaping the bottle, and \( F_2 \) is the flow rate of dry gas. The bottle incorporates a secondary water reservoir to minimize the loss of water inside the bottle. Since \( F_{\text{H}_2\text{O}}^* \approx F_1 \), the relative humidity can be controlled by varying \( F_1 \) and \( F_2 \). The total flow to the specimen chamber is held constant at flow rate of 2000 cc/min.

The inlet line carrying the humid gas is wrapped with a heat trace outside and inside the evacuated flight path to prevent condensation. There is no heat trace on the outlet line outside the evacuated flight path. The temperature of the heat trace is maintained at 20 °C above the bottle temperature.

E. PID controller

The EC is controlled by a program written in LabVIEW 8.5 as outlined in the flow diagram, in Figure 2. The EC system has six independent variables that determine the temperature and humidity inside the specimen chamber. These variables are bottle temperature, outside heat trace temperature (HT1), inside heat trace temperature (HT2), \( F_1 \), \( F_2 \), and recirculating water bath temperature.

The program operates under two modes: manual and automatic. In manual mode, the user defines each of the six independent variables, and there is no PID function. In the automatic mode, the user sets the desired temperature and relative humidity; subsequently the six independent variables are calculated using predetermined calibration curves. Once each component has been set, the program will run a PID function to maintain %RH at ±1.5% relative to the set value. The
FIG. 2. Process control instrumentation flow diagram. Here, water recirculating bath temperature, bottle temperature, HT1 and HT2, and gas flow rates are the controlled variables. The system incorporates four continuous process controls and two manipulated variables. The control loop feedback mechanism regulates the saturation level of the gas by controlling the dry and saturated gas inlet flows using Eq. (1).

LabVIEW program also communicates directly with the PID controller from the humidity generator and the recirculating water bath system.

The control software is currently designed to collect x-ray scattering data at static conditions of T and %RH. Naturally, modifications to the software could readily allow x-ray scattering data to be collected while transitioning from one condition to another. To control the %RH in the specimen chamber, the program uses feedback from changes in %RH to alter saturated gas flow, $F_1$, and the dry gas flow, $F_2$, values. The PID function uses the ratio $F_2: F_1 (F_1 + F_2)$ to regulate the desired condition. The equation is as follows:

$$MV(t) = K_p \left( e(t) + \frac{1}{t_i} \int_{\max(0,t-500)}^{t} e(\tau) d\tau + t_d \frac{d}{dt} e(t) \right),$$

where $MV(t)$ is the correction factor, $K_p$ is the proportional constant, $e(t)$ is the difference between measured and set point %RH, $t_i$ is the integral time, and $t_d$ is the derivative time. This is the traditional PID feedback control equation using parameters that were calibrated for this device. It is important to note, however, that Eq. (2) integrates in a transient mode over the last 500 s of data. This calculation is made every 5 s, and the $F_2$ ratio is then corrected by $MV(t)$. $F_1$ and $F_2$ values are then set using the new $F_2$ ratio.

The first PID sequence runs until the bottle temperature in the humidity generator is 1 to 5 ºC below its set point. In addition, the humidity must be within 3% RH of its set point. When these conditions are met, the first PID loop ends. Next, it copies the last 500 s of %RH data to a second PID loop. The second PID sequence uses the same parameters as the first and runs for an amount of time that the user specifies. During this process, the program records the humidity and temperature values of the specimen chamber every 5 s. Simultaneously, the program initiates the collection of x-ray scattering data, for a designated period of time. Upon completion, the software goes to the next run.

Figure 3 shows the measured temperature and relative humidity as a function of time when the EC is set to 30 ºC with stepwise changes in relative humidity. The temperature as a function of time shows a temperature precision of 0.05 ºC. At relative humidities from 0% to 95% RH the EC exhibits excellent temperature control. The stepwise changes in the relative humidity show a response time of 1 to 2 min.

FIG. 3. Measured temperature (a) and relative humidity (b) for the EC when the temperature is set to 30 ºC and the relative humidity is changes stepwise. The measured temperature is controlled to a precision of ± 0.05 ºC. The standard deviation of the %RH is reported in part b and is 0.4–1.41% RH.
FIG. 4. Measured temperature (a) and relative humidity (b) for the EC when the temperature is set to 80°C and the relative humidity is changes stepwise. The measured temperature is controlled to a precision of ±0.38°C. The standard deviation of the %RH is reported in part b and is 1.25–3.18% RH. The relative humidity is maintained at ±1.5% RH of the set point. The fluctuation in the relative humidity with time is due to minor pressure fluctuations inside the specimen chamber as more saturated vapor enters and exits the system. These data demonstrate that our control system provides precise relative humidity and temperature control within the environmental chamber.

Figure 4 shows the measured temperature and relative humidity as a function of time when the set temperature is 80°C. The temperature shows a variation from the set point of 4°C. However, the measured temperature shows precisions of 0.5°C as a function of time. The response time for changes in the RH is about 1 to 3 min and the overshoot is 50% of the set point at 20% RH and 5% of the set point above 60% RH. The measured humidity shows a precision of up to 3% RH. The cell is less stable at higher temperature due to a reduced ability of the unit to efficiently removing the inlet gas stream with a higher mass rate of vapor. This causes pressure fluctuations inside the specimen chamber which result in changes in dew point. The stability of the specimen chamber could be increased by increasing the outlet orifice size which currently has a diameter of 1/4 in.

III. RESULTS AND DISCUSSION

X-ray scattering was performed with the multi-angle x-ray scattering system (MAXS) at the University of Pennsylvania. The MAXS system generates Cu-Kα x-rays, \( \lambda = 0.154 \text{ nm} \), from a Nonius FR 591 rotating anode operated at 40 kV and 85 mA. The bright, highly collimated beam was obtained via Osmic Max-Flux optics and pinhole collimation in an integral vacuum system. The scattering data were collected using a Bruker Hi-Star two-dimensional (2D) detector with sample-to-detector distances of 11 and 54 cm. As previously discussed the x-ray path length between the Be windows is 0.75 in., a distance that allows good vapor exchange near the samples while minimizing attenuation. The incident beam exhibits a 35% reduction in intensity upon passing through the evacuated specimen chamber due to the two coated Be windows. The incident beam intensity remains constant at 60°C as a function of relative humidity from 10% to 75% RH, Figure 5. This result indicates that attenuation due to water vapor is minimal. Moreover, no new features appear as a function of relative humidity, which allows for easy subtraction. Note that the coating on the inside of the Be windows to prevent corrosion can exhibit Bragg scattering. As Figure 5 demonstrates, the scattering from the coated Be does not interfere with the x-ray scattering measurements.

The capabilities and application of our in situ EC were tested on two polymers: Nafion® membranes and an imidazolium-based polymerized ionic liquid (PIL), Figure 6. The synthesis and characterization of these two polymer systems have been described elsewhere.20, 21

Figure 7 shows the 2D scattering patterns (collected at 86% RH, 25°C) for the Nafion® cast film and electrospun Nafion® nanofiber. The data indicate that ionic aggregates are isotropic in the cast film and anisotropic in the nanofiber, which indicates the presence of oriented ionic domains along the fiber axis direction (Figure 7(b)). Figures 7(c) and 7(d) show the x-ray scattering profiles as a function of relative humidity for Nafion® cast film and electrospun Nafion® nanofiber. The dry cast film and nanofiber have scattering maxima at \( q \approx 1.77 \) and 1.95 nm\(^{-1}\), respectively, indicating the presence of ionic aggregates. Upon exposure to relative humidity, the scattering maxima in both the cast film and nanofibers move to lower \( q \), indicating the system is swelling.
on the nanometer scale. Similar results have been reported elsewhere for cast Nafion® membranes using small-angle x-ray scattering or small-angle neutron scattering as a function of relative humidity. For example, Kim et al. demonstrated that the shift to lower $q$ of the inter-aggregate scattering peak of Nafion® polymer with increasing relative humidity is due to expansion of the distance between ionic domains. The Nafion® nanofiber has a shift in $q$ with increasing relative humidity due to an increase in distance between the aligned nano-domains.

Figure 8 shows the x-ray scattering as a function of relative humidity for poly(MEBIM-OH). The scattering peaks are designated as follows: $q_I = 4.25 \text{ nm}^{-1}$, $q_{II} = 6.14 \text{ nm}^{-1}$, and $q_{III} = 15.15 \text{ nm}^{-1}$ (amorphous halo). As the relative humidity increases, the scattering peaks $q_I$ and $q_{II}$ evolve into one scattering peak located at $4.95 \text{ nm}^{-1}$, while $q_{III}$ moves to a higher angle. We suspect that at 80% RH, the low-$q$ scattering peak results from the combination of two backbone-to-backbone correlation lengths arising from different anions (OH$^-$, HCO$_3^-$, and CO$_3^{2-}$) in the poly(MEBIM-OH) polymer. Fukuta et al. reported that CO$_2$ absorbs to alkaline membranes, that is, OH$^-$ degrades in CO$_2$ to produce HCO$_3^-$ and CO$_3^{2-}$. Garand et al. showed evidence to suggest that water molecules bind to the negatively charged CO$_2$ moieties of the HCO$_3^-$ playing a central role in the acid-based equilibrium formed when CO$_2$ is dissolved in water. Further analysis and discussion is provided elsewhere. Upon reducing the relative humidity to less than 1% in the EC, the new scattering maximum reverted back to two scattering peaks, indicating that the morphology is fully reversible. Overall, these two polymer systems demonstrate the importance of collecting x-ray scattering as a function of relative humidity. The new morphological information will advance the understanding of the fundamental relationship between morphology and other properties such as ionic conductivity and will enable the design of enhanced materials.

**IV. CONCLUSION**

We have reported the design and characterization of an *in situ* environmental chamber for transmission x-ray scattering experiments. Key features include large escape angles, the capability to mount multiple samples, dynamic control of temperature and relative humidity, and integrated computer control. The specimen chamber is compatible with a completely evacuated system such as the Rigaku PSAXS system, in which the specimen chamber is placed inside a larger evacuated chamber (flight path). It is also compatible with x-ray systems consisting of flight tubes interspersed with small air gaps near the sample. Minor modification would adapt the chamber for synchrotron x-ray sources and *in situ* neutron scattering. This environmental chamber can be utilized to study transient morphologies as a function of temperature, relative humidity, and exposure to non-corrosive gases.

We have demonstrated the efficacy and reproducibility of the system to control temperature and relative humidity.

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**FIG. 6.** Chemical structures for (a) Nafion® polymer and (b) poly(1-[(2-methacryloyloxy)ethyl]-3-butylimidazolium-hydroxide) (poly(MEBIM-OH)).

**FIG. 7.** (Color online) 2D x-ray scattering pattern (at 86% RH, 25 °C) of (a) Nafion® cast film and (b) Nafion® nanofiber (arrow indicating longitude direction of fibers). X-ray scattering profile as a function of relative humidity for (c) Nafion® cast film and (d) Nafion® nanofibers. The x-ray scattering data are shifted vertically for clarity.

**FIG. 8.** (Color online) X-ray scattering profile as a function of relative humidity for a polymerized ionic liquid. The x-ray scattering profiles are shifted vertically for clarity.
At 30 °C, the temperature shows a deviation of 0.6 °C from the set point and temperature precision of ±0.05 °C at relative humidities from 0% to 95% RH. The relative humidity shows a response time between 1 and 2 min with minimal overshoot and a precision of ±0.5 °C. The relative humidity shows a response time between 1 and 2 min with minimal overshoot and a precision of ±3% RH. The response time for changes in the RH is ~1 to 3 min and the overshoot is 5% of the set point above 60% RH. The incident beam intensity remains constant as a function of relative humidity indicating that attenuation due to nitrogen and water vapor is minimal due to the short beam path within the EC. Additionally, we demonstrated its potential by studying the morphologies of Nafion® membranes and a polymerized ionic liquid as a function of relative humidity. This environmental chamber extends the possibilities for a detailed understanding of the transformation of morphology of materials.

ACKNOWLEDGMENTS

We thank Professor Yossef A. Elabd, Dr. Bing Dong, and Dr. Yuesheng Ye for providing the Nafion® nanofibers and imidazolium-based polymerized ionic liquids. Also, we thank Thomas B. Schmiedel and Chris W. Gray from W. L. Gore and Associates Inc. for technical advice and assistance. Funding for the construction of this device was provided by the Laboratory for Research on the Structure of Matter (NSF-MRSEC-05-20020). D. Salas-de la Cruz was supported by the Ionic Liquids in Electro Active Devices (ILEAD) MURI grant (CR-19314-430286).