# THE REGULATION OF GENE EXPRESSION DURING MEMORY CONSOLIDATION IN THE HIPPOCAMPUS 

Shane Gary Poplawski
A DISSERTATION
in

## Pharmacology

Presented to the Faculties of the University of Pennsylvania
in
Partial Fulfillment of the Requirements for the
Degree of Doctor of Philosophy
2014

## Supervisor of Dissertation:

Edwin (Ted) Abel
Brush Family Professor of Biology

## Graduate Group Chairperson:

Julie Blendy
Professor of Pharmacology

## Dissertation Committee:

Jim Eberwine, Elmer Holmes Bobst Professor of Pharmacology
Olivier Berton, Assistant Professor of Neuroscience in Psychiatry
Julie Blendy, Professor of Pharmacology
Teresa Reyes, Research Assistant Professor of Pharmacology

## ACKNOWLEDGMENT

I would like to thank Dr. Ted Abel for letting me work on projects that interested me and valuing my input on projects. He has been a tremendous mentor and advisor throughout my graduate career. Any skill I have in science, from experimental design to grant writing, I can attribute in some way to Ted. I also want to sincerely thank Josh Hawk, the former Abel lab graduate student who taught me all the basic procedures I used in this dissertation. More importantly, discussions with Josh taught me how to think about science and the value of doing something you are passionate about. Finally, I am greatly indebted to all of the rest of the Abel lab members who have helped me throughout the way. Although everyone deserves my thanks, Robbert Havekes deserves a special acknowledgment. Robbert took me under his wing and has been a great friend and discussion board throughout my time in the Abel lab.

The members of my thesis committee - Dr. Jim Eberwine, Dr. Olivier Berton, Dr. Julie Blendy and Dr. Teresa Reyes - have been a great help. It is easy to get caught up in your own lab's ideas, so I always enjoyed having experts in related fields listen to (and correct) my ideas.

I would not be here without the support of my family. My parents, two brothers, and sister-in-law have been essential to any success I have had. I also need to acknowledge my grandparents. My grandfather's battle with Parkinson's disease was a huge motivation to research the brain and has continued to inspire my efforts in lab. My grandmother is always thrilled to hear about the research I am doing despite not understanding the scientific jargon I often use. I love and appreciate you all.

Graduate school would be an impossible endeavor without close friends who understand what you are going through. Everyone in my PGG class has helped make
this period of my life as enjoyable as it could be. I especially want to thank Melissa Love and Corey Cannon. Melissa shares my passion for sports (particularly Detroit teams) and has always been willing to chat for hours with me about mostly useless subjects. Corey is a fellow pessimist and scotch lover who was an invaluable daily distraction from any failures or frustrations in lab.

Finally, and most importantly, I need to thank my fiancée Tracy Trexler. Tracy and I have been together long enough to go through drastic life changes, but we always go through changes together and seem to be ever stronger. I can't wait for our future adventures as we figure out life together.


#### Abstract

THE REGULATION OF GENE EXPRESSION DURING MEMORY CONSOLIDATION IN THE HIPPOCAMPUS

Shane Gary Poplawski Edwin (Ted) Abel


Memory consolidation is the process through which short-term memories are stabilized for long-term retention. New gene expression is required for this process to occur successfully. Although gene expression is a necessary component for memory consolidation, the targets and regulation of this gene expression are not well understood. The advent of next-generation sequencing technologies has provided a tremendous resource to probe important questions genome-wide in ways that were previously impossible. In this dissertation, I use next-generation sequencing to investigate the transcriptional targets of learning in the hippocampus. Chapter 1 reviews the previous research on the regulation of gene expression during memory consolidation. Previous work has implicated histone acetylation as an epigenomic modification that regulates long-term memory. In Chapter 2, I use RNA-seq to investigate the gene expression changes that occur 30 minutes after contextual fear conditioning. I use recently developed analysis techniques to improve our ability to detect changes and study alternative splicing genome-wide for the first time after learning. Chapter 3 investigates whether these gene expression changes are specific to contextual fear conditioning or shared with other hippocampus-dependent learning tasks such as object-location memory. I find that the transcriptional targets are similar between training paradigms, but their temporal activation differs. In Chapter 4, we use ChIP-seq, Sono-seq and MNase-
seq to determine changes in histone acetylation, chromatin accessibility and nucleosome positioning that occur in response to learning. I find only small changes in H3K9/14ac, but large changes in chromatin accessibility. This may suggest that a multitude of histone modifications act in concert to regulate chromatin accessibility during memory consolidation.

## TABLE OF CONTENTS

ACKNOWLEDGMENT ..... ii
ABSTRACT ..... iv
LIST OF TABLES ..... ix
LIST OF FIGURES ..... x
CHAPTER 1: Epigenomic Regulation Of Gene Expression In Long-term Memory.. ..... 1
Abstract ..... 1
1.1 The hippocampus and contextual memory ..... 2
1.2 Long-term contextual memory and CREB-dependent transcription ..... 2
1.3 Epigenetic regulation of transcription ..... 4
1.4 The role of histone acetyltransferases in long-term contextual memory ..... 5
1.5 Acetylation of specific lysine residues on histone tails may be important during memory consolidation ..... 7
1.6 Histone deacetylase inhibitors increase long-term memory ..... 9
1.7 Hippocampal gene regulation after contextual fear conditioning ..... 10
1.8 Hippocampal gene regulation after spatial memory tasks ..... 11
1.9 Next-generation sequencing technology and gene regulation in the hippocampus ..... 12
Figure Legends ..... 15
Contributions ..... 17
Figures ..... 18
Tables ..... 21
CHAPTER 2: Determining the transcriptional targets of contextual fear conditioning ..... 26
Abstract ..... 26
Introduction ..... 28
Materials and Methods ..... 30
Results ..... 33
Discussion ..... 37
Figure Legends ..... 40
Contributions ..... 43
Figures ..... 44
Tables ..... 50
CHAPTER 3: Object-Location Training Elicits An Overlapping But Temporally Distinct Transcriptional Profile From Contextual Fear Conditioning ..... 72
Abstract ..... 72
Introduction ..... 73
Materials and Methods ..... 74
Results ..... 78
Discussion ..... 81
Figure Legends ..... 84
Contributions ..... 86
Figures ..... 87
Tables ..... 92
CHAPTER 4: Chromatin Accessibility Is Increased After Learning ..... 96
Abstract ..... 96
Introduction ..... 97
Materials and Methods ..... 98
Results ..... 104
Discussion ..... 109
Figure Legends ..... 111
Contributions ..... 114
Figures ..... 115
Tables ..... 121
CHAPTER 5: Conclusions And Future Directions ..... 165
5.1 Gene expression changes during memory consolidation ..... 166
5.2 Chromatin accessibility during memory and the histone code hypothesis ..... 169
5.3 Future Directions: The need for sorting technologies for brain epigenomic research 172
Figure Legends ..... 174
Contributions ..... 175
Figures ..... 176
APPENDIX: Cell-type Specific Epigenomics In The Brain ..... 177
Abstract ..... 177
Introduction ..... 179
Materials and Methods ..... 181
Results ..... 186
Discussion ..... 187
Figure Legends ..... 191
Contributions ..... 193
Figures ..... 194
BIBLIOGRAPHY ..... 199

## LIST OF TABLES

Table 1.1. Summary of hippocampus-dependent memory in HAT mutant mice ..... 21
Table 1.2. Summary of HAT homology and lysine modifications ..... 22
Table 1.3. Summary of HDAC classes and relation to long-term memory ..... 24
Table 2.1. Results of differential expression test after upper quantile
normalization ..... 50
Table 2.2. Results of differential expression test after RUV normalization ..... 52
Table 2.3. Results of diffSplice analysis ..... 69
Table 3.1. Taqman Assays Used In This Experiment ..... 92
Table 4.1. List of significantly regulated Sono-seq peaks after learning ..... 121
Table 4.2. Comparison of differential Sono-seq regions to gene lists that regulate memory ..... 163
Table 4.3. Comparison of publicly available ChIP-seq data to our differential Sono- seq regions ..... 164

## LIST OF FIGURES

Figure 1.1. Contextual Fear Conditioning Paradigm18Figure 1.2. Regulation of histone acetylation at promoters of genes necessary for
long-term memory ..... 19
Figure 1.3. Object-location memory paradigm ..... 20
Figure 2.1. RUV normalization allows proper grouping of replicates ..... 44
Figure 2.2. Normalization impacts differential expression after contextual fear
$\qquad$conditioning45
Figure 2.3. Comparison of gene lists after upper quantile or RUV normalization ..... 46
Figure 2.4. Exon-specific regulation of Homer1 (Ania-3) ..... 47
Figure 2.5. Exon-specific regulation of Las1I ..... 48
Figure 2.6. Exon-specific regulation of Rbm3 ..... 49

Figure 3.1. Classic IEGs Show Expected Expression Changes after OLM
$\qquad$Training87

Figure 3.2. Limited Expression Changes of Nuclear Receptors after OLM
training ..... 88

Figure 3.3. Modifiers of Histone Acetylation Display Limited Regulation after OLM
$\qquad$Training89

Figure 3.4. No Changes in Other Transcriptional Regulators after OLM Training .. 90

Figure 3.5. OLM Training Induces Long-Lasting Changes in Gene Expression Not
$\qquad$

Figure 4.1. DEScan flowthrough

Figure 4.2. H3K9/14ac shows the most promoters with >2 fold differences 116

Figure 4.3. H3K9/14ac signal is slightly increased around the TSS after
$\qquad$
learning 117

Figure 4.4. Sono-seq signal has a dramatic increase around the TSS after
$\qquad$
learning
118

Figure 4.5. Differences in Sono-seq signal are primarily located within gene
$\qquad$
bodies 119

Figure 4.6. No difference in nucleosome positioning around the TSS afterlearning120
Figure 5.1. Model for the regulation of chromatin accessibility after learning. ..... 176

Figure A.1. Using a tagged histone H 3.3 as a novel epigenetic tool to sort chromatin194

Figure A.2. Doxycyline suppresses expression of H3.3-HA 195

Figure A.3. Expression of H3.3-HA is limited to excitatory neurons.196

Figure A.4. ChIP for H3.3-HA enriches for promoters of genes active in excitatory neurons197

Figure A.5. ChIP for H3.3-HA enriches for promoters over gene bodies............... 198

# CHAPTER 1: Epigenomic Regulation Of Gene Expression In Long-term Memory 


#### Abstract

Long-term contextual memory is formed in the hippocampus through a transcriptiondependent process known as memory consolidation. This transcription is regulated by histone acetylation, an activating epigenetic mark. Determining the specific lysine residues and genetic targets of histone acetylation during learning is crucial to understanding the transcriptional regulation that occurs in the hippocampus. Chromatin accessibility, the result of histone acetylation changes, is also poorly studied during memory consolidation. Next-generation sequencing technology has fostered unprecedented discoveries in the field of gene expression and epigenetic regulation, but has yet to uncover the genetic regulation that occurs in response to learning. In this work, I use high-throughput techniques to investigate the gene expression and epigenetic changes that occur in the hippocampus during memory consolidation.


### 1.1 The hippocampus and contextual memory

A role for the hippocampus in forming episodic memories in humans was first described by Scoville and Milner over 50 years ago [1]. Our understanding of the role of the hippocampus in long-term memory has since been greatly supported by rodent studies. Long-term contextual memories are formed in the hippocampus of the rodent through a process involving distinct stages of acquisition, consolidation, maintenance and retrieval [2]. Contextual fear conditioning is a behavior that allows for dissection of particular phases of memory in the hippocampus due to the temporal resolution of using only one brief training session [3, 4]. Contextual fear conditioning consists of placing a mouse in a novel context that is paired with an aversive foot shock. The memory is then tested by measuring freezing, a species-specific fear response, at 1 hour after training for shortterm memory and 24 hours after training for long-term memory (Figure 1.1). Contextual fear memory formation depends on at least two brain systems: the hippocampus and the amygdala [4]. Hippocampal lesions impair acquisition [5] and produce temporally graded retrograde amnesia $[6,7]$ for contextual fear memory. Thus, components of contextual fear memory traces appear to be formed and stored in the hippocampus for weeks prior to systems-level consolidation into the cortex [8].

### 1.2 Long-term contextual memory and CREB-dependent transcription

Memory consolidation, the process by which memories become stored [9, 10], requires transcription to convert labile short-term memories into stable long-term memories. The need for transcription in this process was first demonstrated over 40 years ago through the ability of actinomycin D , a transcription inhibitor, to block long-term memory in goldfish [11]. This finding was extended by studies demonstrating that two necessary
waves of gene transcription and protein translation occur in the hippocampus after memory formation and correlate with previously determined waves of PKA activity [12, 13]. Although transcription has been known to be involved in memory formation for many years, the processes regulating this transcription have only recently begun to be unraveled [14].

In the hippocampus, there are two time windows after learning that exhibit increased phosphorylation of the transcription factor cAMP-response element-binding protein (CREB), which activates this protein and is thought to mediate memorypromoting transcription [15]. These two windows of CREB phosphorylation, 0-30 min and 3-6 hr after training, coincide strikingly well with the two time windows during which inhibition of transcription or translation impairs memory storage [13, 16]. These gene expression changes after learning occur in between 18-40\% of the neurons in the pyramidal cell layers of the hippocampus, depending on the specific subregion observed [17, 18]. Importantly, long-term memory is affected in both flies [19] and mice [20, 21] bearing mutations in CREB, highlighting the importance of this transcription factor. Although CREB is a necessary transcription factor regulating the transcription required for long-term memory formation, studies have shown that CREB phosphorylation alone is not sufficient to drive expression of target genes [22], indicating that additional coactivators of CREB are required for transcription of target genes. One of these coactivators, CREB-binding protein (CBP), has been shown by our lab and others to be crucial for long-term memory [23-28]. CBP is a histone acetyltransferase (HAT), an enzyme that adds acetyl groups to the N -terminal tails of histones to activate transcription.

### 1.3 Epigenetic regulation of transcription

Substantial advances in our understanding of the regulation of transcription have been made in the field of epigenetics, which is the study of stable alterations of gene expression that do not involve changes in the underlying DNA sequence. The fundamental unit of chromatin is the repeated nucleosome structure, which consists of the four core histones, each in duplicate, tightly encircled by genomic DNA. The aminoterminal tails of histone proteins protrude out of the nucleosome and are sites for posttranslational modifications that regulate the ability of the transcriptional machinery to bind to and transcribe the underlying genes. Specific amino acid residues on histone tails are targets for acetylation, methylation, phosphorylation, sumoylation, and/or ubiquitination among other modifications [29]. These modifications are postulated to form a combinatorial code that defines the transcription status of a given loci. This is known as the "histone code" hypothesis [30]. The epigenetic landscape at promoters can regulate accessibility of chromatin for current or future transcriptional activation [31, 32].

In general, histone acetylation and phosphorylation are activating marks while histone methylation can be activating or repressing depending on the lysine residue targeted [29]. Each modification is expected to have a "writer" that deposits the mark, a "reader" that binds to the mark for a given function, and an "eraser" that removes the mark [31, 33]. In the case of histone acetylation, histone acetyltransferases (HATs) are the "writers" that add acetyl marks, histone deacetylases (HDACs) are the "erasers" that remove acetyl marks, and bromodomain-containing proteins can target and bind histone acetylation as "readers" [33]. Histone modifications also often associate with another epigenomic mark, DNA methylation [34], to facilitate or repress transcription. The epigenetic code at a given locus is thought to regulate the accessibility of chromatin in that region. The accessibility of chromatin regulates whether transcriptional machinery
will bind [35, 36]. Given this complexity, it is important to determine which chromatin modifying enzymes are required for a process such as memory before studying the epigenetic code resulting from learning.

### 1.4 The role of histone acetyltransferases in long-term contextual memory

The concept of DNA-histone complexes regulating memory was first speculated as early as 1975 [37]. Several proteins that interact with phosphorylated CREB are histone acetyltransferases (HATs), including CBP, p300 and p300/CBP associated factor (PCAF). Histone acetylation appears to play a critical role in a number of psychiatric and neurological disorders including depression, schizophrenia and intellectual disability [38, 39]. Therefore, these histone acetyltransferases may be crucial regulators of the transcription necessary for long-term memory. Studies have shown that each of these HATs serves a role in distinct types of memory formation (Table 1.1). Mice lacking Pcaf display short-term memory deficits that gradually worsen with age [40]. Although this finding may appear to link PCAF histone acetyltransferase function with memory, shortterm memory is transcription-independent and therefore this deficit is likely due to developmental defects in the knockout animals. Morphological analysis of the mutant mice showed loss of cells in the CA1 and CA3 subfields of the hippocampus, further supporting the notion of developmental defects causing memory impairments in these mice [40]. Mice overexpressing a truncated form of $p 300$ [41] and mice with conditional p300 deletion [42] demonstrate selective long-term memory deficits in both contextual fear conditioning, which is hippocampus- and amygdala-dependent and object recognition memory, which is hippocampus-independent [43]. Hippocampus-dependent spatial memory is not affected in either of these mice, which indicates p300 is not
required for this form of memory and may suggest that the memory deficits in the p300 mutant mice are due to transcriptional effects in brain regions outside the hippocampus.

The most thoroughly studied histone acetyltransferase in memory is CREB binding protein (CBP). CBP is a transcriptional coactivator that is recruited to phosphorylated CREB [44], as well as other transcription factors [45]. Mutations of Cbp or $p 300$ in the human population cause a form of intellectual disability termed Rubinstein-Taybi syndrome [46]. Recent studies by our lab and others have demonstrated that the histone acetyltransferase CBP is required for efficient long-term memory consolidation [23-27]. Six independently generated Cbp alleles disrupt longterm memory without impairing short-term memory [23-27, 47]. We have produced mice in which CBP activity in neurons is reduced by the transgenic expression of an inhibitory form of CBP lacking the HAT domain [26]. These mice exhibit selective deficits in longterm contextual memory, underscoring the importance of the histone acetyltransferase activity of CBP for memory consolidation. As a coactivator with intrinsic histone acetyltransferase activity, CBP interacts with numerous transcription factors and contains multiple functional domains. Importantly, mice homozygous for a mutation in the kinase-inducible interaction (KIX) domain of CBP, in which CBP is unable to interact specifically with the CREB/ATF transcription factor family, are impaired in long-term memory formation [27, 48]. It was reported that mice with forebrain-specific full Cbp deletion displayed both short- and long-term memory deficits [49], a result that has not been observed with any previous Cbp mutant mice. However, deletion of CBP in adult mice using viral Cre expression causes deficits in synaptic plasticity and a long-term form of hippocampus-dependent memory selectively [28]. This finding would indicate that the deficits in short-term memory observed in mice with the CaMKIl $\alpha$-driven
forebrain deletion were probably due to developmental defects, rather than due to a role of CBP in short-term memory. Taken together, the studies of the role of the histone acetyltransferase CBP in long-term memory consolidation suggest that histone acetylation may be a crucial regulator of transcription during memory consolidation.

### 1.5 Acetylation of specific lysine residues on histone tails may be important during memory consolidation

Because individual HATs are known to acetylate specific lysine residues on histone tails, the possibility exists that acetylation of particular lysine residues controls the transcription necessary for long-term memory (Table 1.2). Although the first evidence for histone acetylation occurring during memory consolidation came by studying the incorporation of radioactive acetyl-CoA into histones in 1979 [50], the first study to demonstrate changes in particular histone acetylation marks was made in 2004 with the finding that acetylation of lysine 14 on histone H 3 increases in bulk histone extracts one hour after contextual learning [51]. This mark returns to baseline levels by 2 hours after fear conditioning [52]. A study by Peleg et al. [53] has found multiple acetylation marks upregulated one hour after contextual fear conditioning including lysine 9 of histone H3 and lysines 5,8 and 12 of histone H 4 . These changes were studied in both young mice, which are able to properly form long-term memories, and aged (16 month old) mice, which have long-term memory deficits. Lysine 12 of histone H 4 was the only mark specifically increased in young mice but not in aged mice, indicating that this acetylation mark may be a key regulator of the transcription necessary for long-term memory formation.

The most direct method to determine the histone acetylation marks pertinent to memory formation would be to study those affected by the histone acetyltransferases known to be required during memory consolidation, such as CBP. In vitro, CBP and p300 have multiple overlapping lysine targets on histone tails. However, recent in vivo work has suggested a more circumscribed role for individual HATs in regulating acetylation of specific lysine residues. Work by Jin et al. [54] has found that knockout of PCAF in mouse embryonic fibroblast cells only decreases acetylation at lysine 9 of histone H 3 , whereas loss of CBP/p300 showed remarkable specificity for decreasing acetylation of lysines 18 and 27 of histone H3. It is important to note that changes in acetylation of lysines 18 and 27 of histone H 3 were not examined in previous studies of bulk histone acetylation changes after learning. The observed specificity of CBP/p300 histone acetylation in fibroblasts could be due to compensation by other histone acetyltransferases at a subset of CBP/p300 target lysines or the guiding of HAT activity by accessory components that provide target specificity. In either scenario, the targets of specific HATs may be different in the hippocampus, the site where memory is consolidated. To bypass these problems, Barrett et al. have used a focal viral deletion of CBP to study the histone marks regulated by CBP in the adult mouse hippocampus [28]. Viral deletion has the benefit of temporal control, which limits compensation by other histone acetyltransferases, as well as stereotaxic control to only affect the brain region of interest. Using this technique, acetylation differences were observed on lysine 14 of histone H3, lysine 12 of histone H2B, and lysine 8 of histone H 4 , but not the mark decreased in aged mice, lysine 12 of histone H4 [28]. Lysines 18 and 27 of histone H3 were not studied in this work. Determining the genes regulated by these acetylation marks during memory consolidation promises to uncover interesting targets that are important for long-term memory, and future methods targeting these specific lysine
residues could hold potential for novel therapeutics that would improve memory formation while limiting side effects.

### 1.6 Histone deacetylase inhibitors increase long-term memory

The increases in histone acetylation during memory consolidation suggest that artificially increasing histone acetylation could enhance long-term memory. Histone acetylation at a promoter is controlled through a delicate balance of HATs, such as CBP, that add acetyl groups to specific lysine residues on histone tails, and histone deacetylases (HDACs), which remove acetyl groups from these lysines (Figure 1.2). Increasing histone acetylation could be achieved either through enhancing HAT activity or by reducing memory-suppressing HDAC activity. HDAC inhibitors, which increase histone acetylation, given during memory consolidation enhance long-term memory [55, 56]. Our work has shown that the memory enhancement by the HDAC inhibitor trichostatin A (TSA) requires the CREB-CBP interaction, indicating that CREB target genes are those required for the memory enhancement. Surprisingly, TSA increased expression of only two out of fourteen CREB target genes examined, the orphan nuclear receptors Nr4a1 and Nr4a2 [55]. It was previously shown that knock-down of Nr4a2 expression impairs long-term memory formation in a spatial discrimination task [57, 58], and global constitutive, heterozygous Nr4a2 knockout mice are impaired in long-term memory formation in the hippocampus-dependent passive avoidance task [59] indicating that this gene expression change may be important for the memory enhancing effects of HDAC inhibitors [60]. A study by our lab found that mice expressing a dominant negative form of NR4A to block all family members display long-term memory deficits [61]. Excitingly,
these mice also do not respond to TSA, indicating this gene family is a crucial regulator of memory enhancement by TSA.

Determining the molecular targets and pathways affected by HDAC inhibitor treatment promises to uncover new genes necessary for contextual memory as well as providing novel avenues for therapeutic intervention for diseases in which memory is affected. Studying the role of particular HDACs in memory formation is another strategy that could lead to development of selective pharmacological agents that cause memory enhancement. HDACs are classified into four families (I-IV) based on sequence homology and structure [62]. Current studies indicate a role for class I HDACs in longterm memory formation (Table 1.3). Class I HDACs act in the nucleus and include HDACs 1-3 and 8. Genetic evidence has demonstrated that both HDAC2 and HDAC3, but not HDAC1, are required for long-term memory consolidation [58, 63]. Targeting only those specific HDAC proteins that dampen memory formation could lead to the application of selective therapeutics with greatly reduced side effect profiles compared to broad-spectrum HDAC inhibitors. This is especially important considering that chronic treatment of broad-spectrum HDAC inhibitors, such as TSA, causes synaptic dysfunction [64].

### 1.7 Hippocampal gene regulation after contextual fear conditioning

The genes regulated by contextual fear conditioning have received tremendous attention as potential targets to modulate memory. Early investigations discovered a set of inducible transcription factors, known as immediate early genes (IEGs), regulated by learning [65]. These include the genes c-fos and Egr1 (zif268) that are induced by neuronal activity. Also, brain-derived neurotrophic factor (Bdnf) expression levels
increase in CA1 after contextual fear conditioning [66]. Mutant mice with any of these genes knocked out demonstrate they participate in long-term memory [67-69]. Microarray analyses after contextual fear conditioning discovered additional phosphatases (Dusp1) and nuclear receptors (Nr4a1) among other genes, but the genes regulated by learning did not show much overlap between labs [70, 71]. Our lab has focused on CREB-dependent genes [72], including Nr4a1 and Nr4a2, which both increase after contextual fear conditioning and are required for long-term memory [61]. We have also found that CREB-dependent Gadd45b and Gadd45g increase after contextual learning and that Gadd45b knockout mice show selective long-term memory deficits [73].

Despite this progress, discovery of the genes regulated by contextual learning remains challenging. This is because of the heterogeneity of the hippocampus, which dilutes real signal from activated excitatory neurons with signal from unactivated excitatory neurons, glia and inhibitory neurons. Thus, the gene expression changes observed are often small. Statistical methods for removing variance have begun to correct this problem, but have yet to be applied genome-wide to data after learning.

### 1.8 Hippocampal gene regulation after spatial memory tasks

Although the targets and temporal dynamics of gene expression after contextual fear conditioning have been well studied, transcription after other spatial memory tasks is less understood. Object-location memory is a hippocampus-dependent spatial memory task in which a mouse learns the spatial orientation of objects within a novel chamber (Figure 1.3). Unlike contextual fear conditioning, object-location memory does not rely on a noxious footshock to induce a learning event. This is important because there is a
concern that some gene expression changes in the hippocampus could be caused by fear instead of learning [74]. Because of the propensity for mice to explore novelty, when an object is moved to a new location in the box 24 hours later, the mouse will preferentially explore that object over the other non-displaced objects [43, 56, 75]. Spatial memory tasks such as object-location memory induce gene expression of many of the same genes observed to change after fear conditioning [76], but there have been few large-scale studies of the targets and temporal dynamics of gene expression after these tasks.

Histone acetylation has also been implicated in long-term spatial memory. Viral deletion of the histone acetyltransferase CBP in the adult hippocampus blocks long-term (but not short-term) object-location memory [28]. Acetylation of lysine 12 of histone H2B and lysine 12 of histone H 4 are both increased in bulk histone extracts during spatial memory consolidation [77]. Our lab has found that class I HDAC inhibition using MS275 enhances long-term object-location memory [56]. MS275 has highest affinity for HDACs 1 and 2, moderate affinity for HDAC3, and low affinity for HDAC 8. Another HDAC inhibitor that preferentially binds to HDAC3, RGFP136, has also been shown to enhance long-term memory using this task [58]. It is currently unclear whether HDAC2 and HDAC3 have overlapping genetic targets important for memory formation, or whether each regulates expression of distinct genes.

### 1.9 Next-generation sequencing technology and gene regulation in the

## hippocampus

Research into gene expression and chromatin regulation has been revolutionized by the advent of next-generation sequencing technology. RNA-seq takes advantage of high-
throughput sequencing to quantify gene expression levels of the entire transcriptome. RNA-seq provides increased sensitivity over microarrays, allows for discovery of novel gene isoforms, and can be used to study alternative splicing [78]. ChIP-seq couples chromatin immunoprecipitation (ChIP) with next generation high throughput sequencing to determine the location of chromatin bound proteins genome wide. ChIP-seq can be used to study histone modifications or transcription factor binding. DNase-seq [79], FAIRE [80], ATAC-seq [81], and Sono-seq [82] are all methods to isolate "open" regions of chromatin. Chromatin being described as "open" or "loosened" means that these regions are thought to be accessible for binding of transcription or epigenetic factors. It is expected that a change in epigenetic modifications would result in alterations in this accessibility, with activating marks such as histone acetylation causing more opening. These powerful sequencing techniques are only recently being applied to studies of long-term memory [53, 76, 83], but often with too few biological replicates to be statistically meaningful in the brain. Bioinformatic analysis of high-throughput sequencing data is a rapidly evolving field and results obtained are highly dependent on analysis method, so implementation of the proper algorithms is necessary to gain an understanding of biological phenomena.

In this work, I use next-generation sequencing technology to understand gene regulation in the hippocampus after learning and stress. Chapter 2 utilizes RNA-seq to determine the gene and exon-level gene expression changes that occur 30 minutes after contextual fear conditioning. Chapter 3 uses a microfluidic high-throughput qPCR system to study gene expression after object-location memory, a spatial memory task that does not rely on a noxious footshock to study memory. Chapter 4 utilizes ChIP-seq
and Sono-seq to study the chromatin regulation that occurs 30 minutes after fear conditioning to regulate gene expression.

Figure Legends

Figure 1.1. Contextual Fear Conditioning Paradigm. In contextual fear conditioning, a mouse is placed into a novel context, allowed to explore, and delivered a mild footshock. The mouse is then returned to the homecage. Upon being reintroduced to the training context, the mouse will exhibit the species-specific fear response of freezing. The time spent freezing can be measured as a correlate for how well the mouse associates the context with the footshock.

Figure 1.2. Regulation of histone acetylation at promoters of genes necessary for long-term memory. After learning, CREB is activated by phosphorylation, binds to CREB response elements (CREs) in the genome, and recruits the coactivator CBP to the region. Acetyl groups are added to lysine residues on histone tails by the histone acetyltransferase function of CBP. Acetylation is removed by class I histone deacetylase (HDAC) proteins.

Figure 1.3. Object-location memory paradigm. In object-location memory, mice are trained to learn the spatial configuration of 3 objects relative to a cue on one wall. After 3 training sessions, mice are removed to their homecage. 24 hours later, mice are placed back in the context with one object displaced. Because mice naturally prefer novelty, they will explore the displaced object more than non-displaced objects.

Table 1.1. Summary of hippocampus-dependent memory in HAT mutant mice. PCAF mutant mice had impaired shock sensitivity, so fear conditioning was not measured. Impaired short- and long-term spatial memory in these mice suggests a
transcription-independent mechanism. Both p300 mutant mice show impaired long-term contextual fear conditioning, but normal spatial memory suggesting a non-hippocampal effect. CBP mutant mice generally show selective hippocampal long-term memory deficits.

Table 1.2. Summary of HAT homology and lysine modifications. Each HAT in the mouse genome is shown within a homology family with known targeted lysines. If a predominant mark is known, it is shown in bold. Clock has homology to both the SRC and MYST family, but is shown in the SRC family in this table.

Table 1.3. Summary of HDAC classes and relation to long-term memory. Class I HDACs have been highly implicated in long-term memory. HDACs 4,5 , and 6 also have evidence linking them to memory but this may or may not be through regulating histones. HDAC3 was shown to regulate memory in a CBP-dependent manner.

## Contributions

This chapter was written by Shane G. Poplawski with suggestions by Ted Abel.

Portions of this chapter were published in: Poplawski SG and Abel T. (2012) "The Role of Histone Acetylation in Long-Term Memory Storage". In P. Sassone-Corsi \& Y. Christen (Eds.), Epigenetics, Brain and Behavior (pp 71-80). Berlin: Springer-Verlag.

Figures
Figure 1.1


Figure 1.2


Figure 1.3


## Tables

Table 1.1

| Mouse model | Short-term <br> Contextual FC | Long-term <br> Contextual FC | Spatial Memory |
| :---: | :---: | :---: | :---: |
| PCAF KO | $?$ | $?$ | Impaired (short and long- <br> term, progresses with age) |
| Truncated p300 | Normal | Impaired | Normal |
| Conditional p300 <br> KO | Normal | Impaired | Normal |
| Truncated CBP <br> (CBP+/-) | Normal | Impaired | $?$ |
| CBP-kix mutant | Normal | Impaired | Impaired (water maze) |
| Truncated CBP <br> (CBPD1) | Normal | Impaired | $?$ |
| CBP+/- | Normal | Impaired | Normal (water maze) |
| CBP HAT- | Normal | Normal | Impaired (water maze) |
| Floxed CBP <br> (CaMKIIa-cre) | Normal [48] <br> Impaired [50] | Normal [48] <br> Impaired [50] | Normal (water maze) [48] <br> Impaired [50] |
| Hippocampal <br> CBP knockout <br> (Cre virus) | Normal | Impaired | Impaired (OLM) |

Table 1.2

| GCN-related Family |  |  |
| :---: | :---: | :---: |
| Name | Other Names | Lysine Modified |
| Gcn5 | Kat2a | H3K14, H3K18 [84] |
| Pcaf | Kat2b | H3K9 [54] |
| Hat1 | Kat1 | H4K5, H4K12, H2AK5 [85] |
| Elp3 | Kat9 | H3K14, H4K8 [86] |
| Atf2 |  | H4, H2B [87] |
| MYST Family |  |  |
| Name | Other Names | Lysine Modified |
| Myst1 | MOF, Kat8 | H4K16 [88] |
| Myst2 | Hbo1, Kat7 | H3K14 [89] |
| Myst3 | MOZ, Kat6a | H3K9 [90] |
| Myst4 | Morf, Kat6b | H3/H4 [91] |
| TIP60 | Kat5 | H2AK5, H3K14, H4K5, H4K8, H4K12, H4K16 [92] |
| CBP/p300 family |  |  |
| Name | Other Names | Lysine Modified |
| Crebbp | CBP, Kat3a | H3K14, H3K18, H3K27, H4K5, H4K8, H4K12, H4K16 [54, 93] |
| EP300 | p300, Kat3b | H3K14, H3K18, H4K5, H4K8, H4K12, H4K16 [54, 93] |
| SRC (Nuclear Receptor Coactivators) |  |  |
| Name | Other Names | Lysine Modified |
| Ncoa1 | Src1, Kat13a | H3K9, H3K14, H4 [94] |
| Ncoa2 | Src2, Tif2, Kat13c | Unknown |


| Ncoa3 | Src3, Actr | Unkown |
| :---: | :---: | :---: |
| Clock | Kat13d | H3/H4 [95] |
| Other | Other Names | Lysine Modified |
| Name | TAFII250, Kat4 | H3K14, H4 [96] |
| TAF1 | TFIIIC90, Kat12 | H3K14 [97] |
| Gtf3c1 |  | H4, H2A [98] |
| Cdyl | NCOAT | H3K14, H4K8 [99] |
| Mgea5 |  | Unknown |
| Naa50 | Nat15, Hat4 | H4K20, H4K79, H4K91 [100] |
| Naa60 |  |  |

Table 1.3

| Class I |  |  |  |
| :---: | :---: | :---: | :---: |
| Name | Typical Location | Histone Target | Relation to Long-term Memory |
| HDAC1 | Nucleus | Yes | May regulate fear extinction [101] |
| HDAC2 | Nucleus | Yes | Negative regulator of fear memory [63] and extinction [102] |
| HDAC3 | Nucleus/cytoplasm | Yes | Negative regulator of spatial memory [58] |
| HDAC8 | Nucleus | Yes | Unknown |
| Class IIA |  |  |  |
| Name | Typical Location | Histone Target | Relation to Long-term Memory |
| HDAC4 | Nucleus/cytoplasm | Yes | Positive regulator of memory in Drosophilia [103] |
| HDAC5 | Nucleus/cytoplasm | Yes | HDAC5 -/- mice have impaired spatial memory [104] |
| HDAC7 | Nucleus/cytoplasm | Yes | Unknown |
| HDAC9 | Nucleus/cytoplasm | Yes | Unknown |
| Class IIB |  |  |  |
| Name | Typical Location | Histone Target | Relation to Long-term Memory |
| HDAC6 | Cytoplasm | No | HDAC6 reduction improves memory in Alzheimer's model [105, 106] |
| HDAC10 | Nucleus/cytoplasm | Yes | Unknown |
| Class III |  |  |  |


| Name | Typical Location | Histone Target | Relation to Long-term Memory |
| :---: | :---: | :---: | :---: |
| SIRT 1-7 | Nucleus/cytoplasm | Yes | SIRT1 loss impairs memory by <br> increasing miR-134 [107] |
| Class IV |  |  |  |
| Name | Typical Location | Histone Target | Relation to Long-term Memory |
| HDAC11 | Nucleus/cytoplasm | Yes | Unknown |

# CHAPTER 2: Determining The Transcriptional Targets Of Contextual Fear Conditioning 


#### Abstract

Memory consolidation requires transcription to form long-term memories. Significant effort has been dedicated to characterizing the genes that change expression in the hippocampus in response to contextual fear conditioning. However, the genome-wide targets of this transcription have not been thoroughly studied. Previous work studying gene expression on a genome-wide scale after contextual fear conditioning has identified only a small number of genes and has not revealed consistent results between labs. In this work, we use RNA-seq in combination with a novel normalization method that allows reliable and consistent expression differences to be discovered. Using this method, we identified 823 genes regulated after contextual fear memory. These genes cluster with transcriptional regulation, MAPK signaling, calcium signaling and synaptic transmission - functional classes known to be involved in learning and memory. In addition to gene expression differences, RNA-seq also allows exon-level transcription analysis. Studying alternative splicing by differential exon usage during memory consolidation has not been previously attempted genome-wide. Here, we use RNA-seq to determine exon-level changes in expression after contextual fear conditioning. We discovered that Ania-3, a short variant of Homer1 which has not been studied after learning, is regulated by contextual fear conditioning. The ribosome biogenesis regulator Las1l and the RNA-binding protein Rbm3 also change specific exon usage after fear


conditioning. This analysis discovered novel gene and exon regulation after learning that is unlikely to be detected by other methods.

## Introduction

Contextual fear conditioning requires two waves of transcription and protein synthesis to form long-term memory in the hippocampus [16, 108]. Our lab and others have focused on discovering the genetic targets of these transcriptional waves using both candidate gene and genome-wide approaches. Our research has indicated that the first wave of transcription induces the largest change in gene expression at 30 minutes after contextual learning [61, 109]. However, previous genome-wide efforts to study the gene expression during memory consolidation have led to largely inconsistent findings across labs [70, 71, 109-113]. This is due both to study design and restraints of microarray technology, which can introduce variability in hybridization between runs [78]. To resolve the genetic targets of the transcription that occur immediately after contextual fear conditioning, we used RNA-seq. RNA-seq is a recent technology that allows all polyadenylated mRNA within a sample to be sequenced, mapped to the transcriptome, and quantified. RNA-seq has a better dynamic range than microarrays and consistent technical reproducibility [78], so this technology provides the best opportunity to discover reliable transcriptional targets. In addition, we designed our study in such a way that mice from each group were dissected at the same time over multiple days, limiting the effects of circadian rhymicity and differences such as odors or loud noises that could occur on one particular day.

Contextual learning occurs throughout the lifetime of an animal, so isolating the effects of a specific contextual memory can be difficult. Standard RNA-seq normalization is unable to resolve small differences within a complex signal [114]. This is because the signal contains both variation caused by the condition being tested as well as variation caused by unwanted effects. Ideally, RNA-seq analysis would account for this unwanted variation to reveal the true signature of the condition being tested. To this end, we
applied a recently designed normalization method, termed remove unwanted variation (RUV) that accounts for unwanted variation within an RNA-seq data set [114]. Unwanted variation would be any variation in gene expression not caused by the treatment being tested. In the case of our experiment, this could include previous contextual information, individual effects of stress on mice, or even variation caused by RNA extraction and library preparation. This method is much better at resolving complex signals, such as those from the brain, than standard upper quantile normalization, which primarily normalizes for read count differences between samples.

Gene expression is a complex process with multiple layers of regulation. Expression levels of particular isoforms of transcripts can be regulated by alternative start sites, differential splicing including exon skipping and intron retention, and alternative $\operatorname{poly}(A)$ site selection [115]. Alternative splicing can lead to distinct protein function and interactions [116] or regulate mRNA localization [117-119], and thus is expected to be particularly important in neurons with long processes. Previous research studying genome-wide gene expression in the hippocampus after contextual learning has relied on microarray technology [70, 71, 109-113]. Although microarrays are good at identifying gene expression differences, they are often unable to distinguish exon-level effects that are indicative of alternative splicing. RNA-seq provides numerous advantages over microarrays, including the ability to study exon-level changes in gene expression. Isoform-specific gene expression changes are known to occur after contextual fear conditioning, including specific upregulation of Bdnf IV, but not other Bdnf isoforms [120], and Homer1a, but not Homer1c [121]. These examples indicate that transcriptional changes after learning may be more complex than gene-level differences and can be highly selective for particular exons of a gene.

In this study, we used RNA-seq to study gene expression 30 minutes after contextual fear conditioning in the hippocampus. Applying RUV, a recently designed normalization algorithm, to our data, we find that the number of differentially regulated novel and positive control genes dramatically increase. This results in interesting functional classes of genes being identified. We also tested differential alternative splicing 30 minutes after contextual fear conditioning. We discovered 118 exons across 97 genes that showed differential expression of an exon independent of gene-level expression differences. These differences confirmed changes in Snord14e, a small nucleolar RNA that our lab has previously shown to be regulated at this time point [109]. Sno-RNAs regulate RNA processing and have been implicated in memory consolidation [122]. Ania-3, an alternative short form of Homer1 that has not previously been linked to learning, was also shown to be regulated by fear conditioning. These findings demonstrate for the first time on a genome-wide scale that alternative splicing is regulated by contextual learning and also provide novel transcriptional targets that are regulated during memory consolidation.

## Materials and Methods

## Subjects

C57BL/6J mice were maintained under standard conditions with food and water available ad libitum. Adult male mice 2 months of age were kept on a 12-hr light/12-hr dark cycle with lights on at 7AM. All behavioral and biochemical experiments were performed during the light cycle with training starting at 10AM (ZT3).

## Behavior

Fear conditioning was performed as previously described [55, 61] with handling for 3 days prior to conditioning. Briefly, the conditioning protocol entailed a single 2-sec, 1.5 mA footshock terminating at 2.5 minutes after placement of the mouse in the novel chamber. Mice were left in the chamber for an additional 30 seconds and then returned to their homecage. One mouse per behavioral group (homecage and fear conditioned) was trained per day over 5 days. This allowed dissections to occur at the same time and reduce circadian effects.

## RNA isolation

Hippocampi were dissected from homecage mice or fear conditioned mice 30 minutes after the training into RNAlater (Qiagen, Valencia, CA) and frozen on dry ice. Tissue was homogenized using a TissueLyser system and RNA was extracted using the RNAeasy Microarray Tissue kit (Qiagen) according to the manufacturer's instructions. Samples were DNase treated using the RNase-Free DNase kit (Qiagen) off-column by incubating $5 \mu \mathrm{l}$ DNase and $35 \mu \mathrm{l}$ Buffer RDD for 25 min at RT with each sample. Samples were then ethanol precipitated and resuspended in water.

## RNA-seq Library Preparation and Sequencing

$2 \mu \mathrm{~g}$ of RNA from $\mathrm{n}=5$ homecage and fear conditioned mice was used in the TruSeq RNA Sample Prep Kit (Illumina, San Diego, CA) according to the manufacturer's instructions with polyA selection. Completed libraries were size-selected on an agarose gel to remove any high basepair fragments, quantified by qPCR (KAPA Biosystems, Boston, MA), and submitted to the PGFI sequencing core at UPENN. An Illumina HiSeq 2000 sequenced the libraries in paired-end 100bp reads. 3 libraries were sequenced per lane on an Illumina HiSeq 2000, resulting in an average of $67,011,105$ reads per sample
in the homecage mice and $62,115,805$ reads per sample after fear conditioning. Reads had good unique concordance ( $86.9 \%$ in homecage, $85.5 \%$ after fear conditioning) and mapping $(90.7 \%$ of unique concordant reads in homecage and $93.1 \%$ after fear conditioning).

## Data Analysis

Sequencing reads were aligned to the mouse mm9 genome using GSNAP [123] (http://share.gene.com/gmap). A gene count table was produced using HTSeq [124] (http://www-huber.embl.de/users/anders/HTSeq/doc/overview.html). Gene counts were normalized either using standard EdgeR normalization [125] or RUVneg, which corrects for unwanted variation using negative controls [114]. We used 625 genes identified as unchanged from a previous microarray experiment as negative controls for RUV [109]. We discovered that five principal components $(k=5)$ need to be normalized to resolve the differences caused by contextual fear. Significance was calculated using EdgeR [125]. Functional annotation was performed through DAVID [126, 127] (http://david.abcc.ncifcrf.gov/). The annotation was limited to the following sources: GO Biological process, GO Molecular Function, KEGG pathways, and SwissProt and Protein Information Resource keywords and an EASE score restriction of 0.1.

Exons were separated into unique, non-overlapping "bins" using Ensembl gene models and DEXseq [128]. A "bin" can either be part of an exon or an entire exon depending on the uniqueness of the region. A bin read count table was produced using HTSeq. Samples were normalized using RUV. We operated under the assumption that exons from housekeeper genes selected from a previous microarray study did not change. Exon-level analysis was performed using the Bioconductor package voom and diffSplice.

## $q P C R$ analysis

RNA was extracted from a separate cohort of fear conditioned mice as described above. RNA was converted to cDNA using the RETROscript kit (Ambion) according to the manufacturer's instructions. cDNA reactions were diluted to $200 \mu \mathrm{l}$ and $2.25 \mu \mathrm{l}$ was combined with $0.25 \mu \mathrm{l} 5 \mu \mathrm{M}$ primer mix and $2.5 \mu \mathrm{l}$ SYBR Select Master Mix (Life Technologies) and run on a Viia7 Real Time PCR system. The $\Delta \Delta C t$ method was used for analysis as previously described [55]. The primers used for exon-level analysis were: Ania-3 F: 5'- AGTGGCTGGTTTTCTTGGACT-3', Ania-3 R: 5'-GGGAGGTGGATTGGTGACAA-3', Homer1 Bin21 F: 5'-CTGGAGTCCACTGCCAATGT-3', Homer1 Bin21 R: 5’- CTCTGCTTCCTCCTGGTACG3', Las1l Bin15 F: 5'- TCAAAGTCAGAGGGGTCGGA-3', La1l Bin15 R: 5'-AGACTTCGCTCTTGCTGCTT-3', Las1I Bin17 F: 5'- TGCTGGAGAAACACAGGCAT-3', Las1l Bin17 R: 5'- ACATTGTACACGTGGGGAAAGA-3', Rbm3 Bin2 F: 5'-ACCTGAGTTTTGGAGGCTGG-3', Rbm3 Bin2 R: 5'- ACAACAGCGGACACCATAGG-3', Rbm3 Bin7 F: 5'-GGTGGCTATGACCGCTACTC-3', Rbm3 Bin7 R: 5'-TTTTGTGTGCATGCCCCATC-3', Rbm3 Bin22 F: 5'-TGCCCCTGGCAGACATAGAG-3', Rbm3 Bin22 R: 5'-GTCTGCCACTTTCTTCGTTCTTT-3'.

## Results

To study gene expression after learning using RNA-seq, we applied a recently created algorithm, termed remove unwanted variance (RUV) [114], that allows a user to correct for unwanted variance. RUV uses negative control genes (or samples) to test for sources of variation that do not correlate with the factor of interest and correct for that
variation. We used 625 negative control genes from a previous microarray study [109] as controls for this normalization and therefore will refer to the normalized data as RUVneg. There are a number of ways to test whether data normalization is performing its function, but two major quality control tests for RNA-seq data analysis are relative log expression (RLE) plots and principal component analysis. RLE plots measure the count data for a gene in a sample relative to the median of all samples and plot this over all genes. We would expect the mean to be close to 0 and variability between all samples to be minimal on this plot. RUVneg restores this uniformity, while upper quantile normalization does not (Figure 2.1A). Principal component analysis determines which components of the data are the major drivers of variability in a data set. Ideally, the difference being tested (in our case learning) would be the major driver of variability. However, upper quantile normalization shows no clustering of samples in the first two principal components (Figure 2.1B), indicating that learning is not the major driver of differences between groups. RUVneg with normalization of five principal components introduces proper clustering (Figure 2.1B), indicating that any differences observed in this data will primarily be the result of differences induced by contextual learning.

We then applied upper quantile and RUVneg normalization to our data to identify differentially regulated genes. We would expect that the p-value distribution of the results would have a uniform value, with peaks at either end for genes that are obviously changed or unchanged [129]. In other words, there is no logical reason that more genes would have a $p$-value of 0.8 than a $p$-value of 0.2 . However, we see that upper quantile normalization does not have a uniform p-value distribution, while RUVneg does (Figure
2.2A). RUVneg also introduces a large peak at the lowest $p$-value bin, indicating a greater power to detect changes. A major test of this normalization is that it is not introducing artifacts into our data. To test this, we produced a volcano plot showing p-
value vs. fold change ( $-\log _{10} \mathrm{P}$-value vs. $\log _{2} \mathrm{FC} / \mathrm{CC}$ ) (Figure 2.2B). While RUVneg produces many more significant genes than upper quantile normalization (blue), it also greatly improves the number of 18 positive control genes selected from the literature (red) that are detected as significant. This gives us confidence that the genes detected by RUVneg are indeed genes regulated by contextual fear conditioning.

Upper quantile normalization detected 56 genes as differentially regulated (Table 2.1), while RUVneg identified 823 genes (Table 2.2). The 823 genes identified by RUVneg include all 56 genes identified by upper quantile normalization (Figure 2.3). Functional annotation of these gene lists by DAVID [126, 127] also demonstrates the improved ability of RUVneg to identify changes important for learning. Upper quantile normalization produced 7 significant clusters mapping to transcriptional regulation, embryo/blood vessel development and vision. RUVneg produced 20 clusters mapping to transcriptional regulation, synaptic transmission, MAPK activity and calcium homeostasis among other functions (Figure 2.3). Because of the known importance of synaptic changes, MAPK signaling, and calcium signaling to memory [130, 131], this further demonstrates the power of RUVneg. RUVneg, therefore, is a useful tool that identified numerous genes and processes that are regulated during memory consolidation.

RNA-seq also has the advantage of distinguishing exon-level reads that are difficult to detect by any other method, and therefore is an ideal technique to study alternative splicing. We used GSNAP [123] to align reads to the mm9 mouse genome and HTSeq [124] to count reads by DEXSeq bins [128] using Ensembl gene models. Bins are separated based on overlap of Ensembl gene models, with exons that have variable sizes between different transcripts split into multiple bins. Therefore, a bin can represent either a whole exon or part of an exon. RUVneg normalization was performed as described [114], using removal of 5 principle components of variance, which we
discovered is optimal for this dataset. Bioconductor packages voom and diffSplice were then used to determine differential exon usage independent of gene-level changes. We identified 118 bins across 97 genes that displayed differential usage after contextual fear conditioning (Table 2.3). 87 of these exons were upregulated and 31 were downregulated, consistent with the general increase in gene expression after fear conditioning (Figure 2.2). We performed functional classification of genes showing at least 1 exon-specific change after fear conditioning. The SwissProt and Protein Information Resource keyword "alternative splicing" was enriched in our data set, indicating that our exon-level analysis discovers alternative splicing as expected. However, no functional clusters were enriched in either the upregulated or downregulated exon lists.

Upregulated exons included Snord14e, which reside in the introns of the Hspa8 gene. We have recently validated Snord14e upregulation after detecting differences by microarray [109]. We also discovered that a poorly studied short isoform of Homer1 known as Ania-3 is upregulated after contextual fear conditioning. Homer1a has previously been shown to be upregulated by fear conditioning, but Ania-3 has not been studied [121]. To validate our results, we performed qPCR in a separate cohort of mice, comparing the bins observed to change in these genes to an exon of the same gene that was unchanged. Ania-3 was found to be upregulated independently of Homer1 (Figure 2.4). Ribosome biogenesis protein Las1l exhibits bin-specific downregulation in response to contextual fear conditioning (Figure 2.5). RNA-binding protein $\mathrm{Rbm3}$, which our lab has shown to change in the hippocampus after sleep deprivation [132], displays complex regulation with both upregulated and downregulated bins after learning (Figure 2.6). In all cases, the exon predicted to change was significantly regulated while a
control exon in the same gene was unchanged. This indicates that alternative splicing does occur on a genome-wide scale in response to contextual fear conditioning.

## Discussion

Although gene expression has been recognized to be crucial for long-term memory for many years [11, 108], identifying the genome-wide targets of this transcription has proven difficult. This is due both to the limitations of technology used in previous experiments and the inability to distinguish signal from noise. The brain is a heterogenous tissue with only a fraction of excitatory neurons responding transcriptionally to a learning event [17]. This heterogeneity dilutes changes and makes it difficult to identify changes caused by contextual learning. In this study, we used RNAseq to study gene expression in the hippocampus 30 minutes after contextual fear conditioning, a time point our lab has previously determined to show the most expression changes after fear conditioning [61, 109]. We used a recently developed normalization algorithm, RUV, in combination with negative controls identified from a previous microarray experiment to reduce the effects of unwanted variation. We found that RUVneg normalization greatly improves our ability to detect both novel and expected changes with RNA-seq. We also provide the first evidence of genome-wide regulation of alternative splicing after learning in the hippocampus.

RUVneg normalization was able to normalize a number of factors that standard upper quantile normalization does not. This is an important because a major problem with studying gene expression after learning is the heterogeneity of the brain. The brain contains excitatory neurons, inhibitory neurons, glia, and epithelial cells. Only a fraction (18-35\% [17]) of excitatory neurons respond transcriptionally to any memory trace.

Therefore, the signal caused by contextual fear conditioning is highly diluted by surrounding cells, making it small and hard to observe. After RUVneg, the RLE plot was much less variable between samples and principal component analysis clustered samples according to treatment. These normalization measures are essential for producing a reliable list of genes that are caused by the learning event in question. The gene list produced by RUVneg showed a uniform p-value distribution, which would be expected, and identified more novel and positive control genes than standard upper quantile normalization. The larger gene list allowed for identification of more functional classes of genes being regulated that could be followed up for future study. Immediate shock and context only controls will be needed to test whether these gene expression changes are caused by the association between context and shock or caused by either stimulus alone. Importantly, because the goal of RUV is to normalize for sources of unwanted variation, this normalization should be broadly applicable to future studies of this type and greatly enhance consistency and reliability between experiments. We believe that the gene list produced from our data will be highly reproducible by other labs studying contextual fear conditioning.

Because RNA-seq provides the additional advantage of studying alternative splicing, we also used RUV to look at gene counts broken down by bins corresponding to whole or parts of exons. We were able to detect gene expression changes at particular bins occurring in response to contextual fear conditioning at 97 genes. Although individual examples of alternative splicing have been observed during memory consolidation [120, 121, 133], nothing has ever been discovered at this genome-wide level. We confirmed [109] that Snord14e, which exists within an intron of Hspa8, is regulated by fear conditioning. We also implicate the selective expression of particular bins of Homer1 isoform Ania-3, RNA-binding protein Rbm3 and ribosome biogenesis
regulator Las1/ in learning for the first time. This discovery would not have been possible looking at gene-level expression. The mechanism that drives this alternative splicing is unclear, although transcription of certain splicing proteins is known to change after fear conditioning [134]. Our data also indicates regulation of a specific isoform of splicing factor Sfpq (data not shown). However, it is unclear whether these transcriptional changes would have time to translate into protein and effect splicing by 30 minutes. It is possible that changes in epigenetic modifications are regulating this selective exon usage [135], including H3K36me3 and H4K20me1 [136, 137]. Future studies can determine whether the differential bins discovered in this study show differential histone modifications as well. Although the exact function of the exons discovered to be differentially regulated by fear conditioning in this study remains unclear, we hope that these findings drive further study into the mechanisms of isoform specific transcriptional effects during memory consolidation.

## Figure Legends

Figure 2.1. RUV normalization allows proper grouping of replicates. A) Relative log expression (RLE) plot of all samples following either traditional upper-quantile normalization or normalization with RUV using negative controls (RUVneg). This plot compares the ratio between the gene counts for each sample and the median gene counts for all samples and plots the data over all genes. We expect RLE distributions to be centered around zero and as similar as possible to each other. Red samples are controls matched for time of day (CC), blue samples were obtained 30 minutes after memory acquisition (FC), and green samples were obtained 30 minutes after memory retrieval (RT). The RLE boxplots clearly show the need for additional normalization following upper quantile normalization. B) Scatterplot of first two principal components (PC1 and PC2) log-scaled and centered following traditional upper-quantile normalization (left) or normalization with RUV using negative controls (right). Samples do not cluster according to treatment following UQ normalization but do after applying RUV normalization.

Figure 2.2. Normalization impacts differential expression after contextual fear conditioning. A) Distribution of edgeR uncorrected $p$-values for tests of differential expression between control (CC) and fear conditioned (FC) samples for upper quantile normalized counts and RUVneg normalized counts. The y-axis is frequency of each pvalue bin occurring and $x$-axis is increasing $p$-values. The distribution of upper quantile normalized counts is far from the expected uniform distribution. RUVneg returns uniformity to the p -value distribution and increases discovery of differentially expressed
genes (genes that have a low p-value). B) Volcano plot of differential expression (-log ${ }_{10}$ p -value vs $\log _{2}$ fold change) of upper quantile normalized samples and RUVneg normalized samples. Genes with and FDR $<0.1$ are highlighted in blue. Genes known to be regulated by FC (positive controls) are outlined in red. RUV increases the significance of differentially expressed positive controls.

Figure 2.3. Comparison of gene lists after upper quantile or RUV normalization. A) Upper quantile normalization identifies only 56 genes as significantly regulated by fear conditioning. Functional annotation of these genes identifies: Regulation of transcription, Embryo/blood vessel development and Vision. B) RUVneg results in 823 genes regulates by fear conditioning. Functional annotation of these genes identifies: Regulation of Transcription, Synaptic Transmission, Embryo/blood vessel Development, Metal Ion Binding, MAPK activity, Zinc fingers, Calcium homeostasis, Ear development, Cadherins, Fibronectins, Immune response, and Apoptosis.

Figure 2.4. Exon-specific regulation of Homer1 (Ania-3). A) diffSplice result showing the predicted significant bin changes in red. Bins 16-18 indicate the Ania-3 isoform. B) qPCR validation of the change in Bin18 in an independent cohort of mice. Bin 21 expression was compared as a control.

Figure 2.5. Exon-specific regulation of Las1I. A) diffSplice result showing the predicted significant bin changes in red. B) qPCR validation of the change in Bin 15 in an independent cohort of mice. Expression of Bin 17 was used as a control.

Figure 2.6. Exon-specific regulation of Rbm3. A) diffSplice result showing the predicted significant bin changes in red. B) qPCR validation of the changes in Bin 2 and Bin 22 an independent cohort of mice. Expression of Bin 7 was used as a control.

Table 2.1. Results of differential expression test after upper quantile normalization. Genes with an FDR <0.1 after upper quantile normalization are shown. 56 genes were identified. logFC is the fold change between fear conditioned and homecage in $\log _{2}$, $\log C P M$ is the $\log _{2}$ counts per million for that gene, $L R$ is the likelihood ratio (statistical test) and FDR is the multiple testing corrected false discovery rate.

Table 2.2. Results of differential expression test after RUV normalization. Genes with an FDR <0.1 after upper quantile normalization are shown. 823 genes were identified, including all 56 identified using upper quantile normalization. logFC is the fold change between fear conditioned and homecage in $\log _{2}, \log C P M$ is the $\log _{2}$ counts per million for that gene, LR is the likelihood ratio (statistical test) and FDR is the multiple testing corrected false discovery rate.

Table 2.3. Results of diffSplice analysis. diffSplice tests whether the logFC of a particular bin differs from the average logFC of that gene. Ensembl Gene IDs were used for gene classification and broken into bins. 118 bins corresponding to 97 genes were identified. $\log F C$ is the fold change between fear conditioned and home cage animals in $\log _{2}$ format. $t$ represents the t-statistic for the difference. FDR is a multiple-testing corrected false discovery rate. Only exons with an FDR $<0.1$ are shown.

## Contributions

This chapter was written by Shane G. Poplawski with suggestions by Ted Abel, Lucia Peixoto, and Giulia Porcari. Experiments were planned by Shane Poplawski, Lucia Peixoto, and Mathieu Wimmer. Behavior and experiments were carried out by Shane Poplawski, Lucia Peixoto, Mathieu Wimmer, and Giulia Porcari. Data analysis was performed by Lucia Peixoto, Shane Poplawski and Davide Risso (Terry Speed laboratory, UC Berkeley). We thank Giulia Porcari for constructive discussions and critical reading of the chapter.

Figures
Figure 2.1


Figure 2.2


Figure 2.3

A Upper Quantile


Regulation of Transcription Embryo/blood vessel Development Vision

B RUVneg


Regulation of Transcription
Synaptic Transmission
Embryo/blood vessel Development
Metal Ion Binding MAPK activity
Zinc fingers
Calcium homeostasis
Ear development
Cadherins
Fibronectins
Immune response
Apoptosis

Figure 2.4



Figure 2.5


Figure 2.6



## Tables

Table 2.1

| GenelD | Gene Name | logFC | logCPM | LR | p-value | FDR |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000021250 | Fos | 2.33 | 4.13 | 195.21 | 2.3E-44 | 4.2E-40 |
| ENSMUSG00000037868 | Egr2 | 2.54 | 2.04 | 110.33 | 8.3E-26 | 7.5E-22 |
| ENSMUSG00000022602 | Arc | 1.47 | 7.48 | 92.73 | 6.0E-22 | 3.6E-18 |
| ENSMUSG00000028195 | Cyr61 | 1.08 | 2.49 | 53.34 | $2.8 \mathrm{E}-13$ | 1.3E-09 |
| ENSMUSG00000052837 | Junb | 1.06 | 5.94 | 45.64 | 1.4E-11 | 5.1E-08 |
| ENSMUSG00000023034 | Nr4a1 | 1.02 | 6.43 | 43.57 | 4.1E-11 | 1.2E-07 |
| ENSMUSG00000024042 | Sik1 | 0.99 | 3.89 | 41.44 | 1.2E-10 | 3.1E-07 |
| ENSMUSG00000024190 | Dusp1 | 0.89 | 4.37 | 40.82 | 1.7E-10 | 3.7E-07 |
| ENSMUSG00000085609 | 1700016P03Rik | 1.42 | 2.34 | 40.63 | 1.8E-10 | 3.7E-07 |
| ENSMUSG00000020423 | Btg2 | 0.76 | 4.65 | 38.76 | $4.8 \mathrm{E}-10$ | 8.6E-07 |
| ENSMUSG00000061808 | Ttr | 4.09 | 7.15 | 37.94 | 7.3E-10 | 1.2E-06 |
| ENSMUSG00000003545 | Fosb | 0.90 | 3.45 | 36.82 | 1.3E-09 | 2.0E-06 |
| ENSMUSG00000065537 | Mir132 | 1.21 | 0.27 | 30.32 | 3.7E-08 | 5.0E-05 |
| ENSMUSG00000071341 | Egr4 | 1.01 | 5.02 | 29.34 | 6.0E-08 | 7.8E-05 |
| ENSMUSG00000090698 | Apold1 | 0.88 | 2.39 | 28.99 | 7.3E-08 | 8.8E-05 |
| ENSMUSG00000036151 | Tm6sf2 | 0.87 | 2.44 | 28.35 | 1.0E-07 | 1.1E-04 |
| ENSMUSG00000022949 | Clic6 | 2.39 | 2.25 | 26.85 | 2.2E-07 | 2.3E-04 |
| ENSMUSG00000001827 | Folr1 | 1.90 | 1.14 | 26.28 | 3.0E-07 | 3.0E-04 |
| ENSMUSG00000017723 | Wfdc2 | 2.31 | -1.27 | 25.76 | 3.9E-07 | 3.5E-04 |
| ENSMUSG00000053560 | ler2 | 1.23 | 3.11 | 25.79 | 3.8E-07 | 3.5E-04 |
| ENSMUSG00000034739 | Mfrp | 3.30 | 0.41 | 24.15 | 8.9E-07 | 7.7E-04 |
| ENSMUSG00000025488 | Cox8b | 1.86 | -1.50 | 24.03 | 9.5E-07 | 7.8E-04 |
| ENSMUSG00000068323 | Slc4a5 | 3.37 | 0.48 | 23.83 | 1.1E-06 | 8.2E-04 |
| ENSMUSG00000079436 | Kcnj13 | 2.68 | -1.25 | 23.73 | 1.1E-06 | 8.3E-04 |
| ENSMUSG00000037086 | 1110059M19Rik | 2.71 | -1.05 | 23.61 | 1.2E-06 | 8.5E-04 |
| ENSMUSG00000021848 | Otx2 | 1.94 | 0.42 | 22.48 | 2.1E-06 | 0.001 |
| ENSMUSG00000039672 | Kcne2 | 2.96 | 0.33 | 22.96 | 1.7E-06 | 0.001 |
| ENSMUSG00000028348 | Murc | -0.97 | -0.06 | 21.93 | 2.8E-06 | 0.002 |
| ENSMUSG00000026051 | 1500015010Rik | 1.50 | 1.71 | 20.84 | 5.0E-06 | 0.003 |
| ENSMUSG00000004655 | Aqp1 | 2.21 | 0.69 | 20.28 | 6.7E-06 | 0.004 |
| ENSMUSG00000015652 | Steap1 | 1.92 | -1.11 | 19.21 | 1.2E-05 | 0.007 |
| ENSMUSG00000046470 | Sox18 | -0.68 | 3.20 | 18.53 | 1.7E-05 | 0.009 |
| ENSMUSG00000048489 | 8430408G22Rik | 1.14 | -0.04 | 18.57 | 1.6E-05 | 0.009 |
| ENSMUSG00000055148 | Klf2 | 0.94 | 3.37 | 18.53 | 1.7E-05 | 0.009 |
| ENSMUSG00000034936 | Arl4d | 0.78 | 3.37 | 17.95 | 2.3E-05 | 0.011 |
| ENSMUSG00000044595 | Dnd1 | 0.73 | 1.20 | 18.06 | 2.1E-05 | 0.011 |
| ENSMUSG00000034765 | Dusp5 | 0.81 | 4.84 | 17.76 | 2.5E-05 | 0.012 |
| ENSMUSG00000049382 | Krt8 | 1.35 | -0.41 | 17.50 | 2.8E-05 | 0.014 |
| ENSMUSG00000020907 | Rcvrn | 1.18 | -1.05 | 17.20 | 3.4E-05 | 0.016 |
| ENSMUSG00000024793 | Tnfrsf25 | 0.69 | 5.54 | 16.72 | 4.3E-05 | 0.020 |
| ENSMUSG00000040287 | Stac3 | 0.63 | 1.95 | 16.60 | 4.6E-05 | 0.020 |
| ENSMUSG00000037447 | Arid5a | 0.58 | 3.72 | 16.51 | 4.8E-05 | 0.021 |
| ENSMUSG00000079681 | Zglp1 | 0.89 | 0.79 | 16.44 | 5.0E-05 | 0.021 |


| ENSMUSG00000028125 | Abca4 | 1.10 | 1.78 | 15.81 | $7.0 \mathrm{E}-05$ | 0.029 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000030450 | Oca2 | 1.10 | -1.09 | 15.61 | $7.8 \mathrm{E}-05$ | 0.031 |
| ENSMUSG00000026579 | F5 | 2.09 | 1.45 | 15.38 | $8.8 \mathrm{E}-05$ | 0.034 |
| ENSMUSG00000084381 | AA413626 | -0.98 | -0.97 | 15.26 | $9.3 \mathrm{E}-05$ | 0.036 |
| ENSMUSG00000073437 | D330041H03Rik | -1.02 | -1.39 | 14.84 | $1.2 \mathrm{E}-04$ | 0.044 |
| ENSMUSG00000023043 | Krt18 | 1.55 | 0.13 | 14.68 | $1.3 \mathrm{E}-04$ | 0.047 |
| ENSMUSG00000000182 | Fgf23 | -1.13 | -1.42 | 14.48 | $1.4 \mathrm{E}-04$ | 0.051 |
| ENSMUSG00000015467 | Egfl8 | 1.09 | -0.18 | 13.32 | $2.6 \mathrm{E}-04$ | 0.086 |
| ENSMUSG00000026628 | Atf3 | 0.83 | 0.57 | 13.39 | $2.5 \mathrm{E}-04$ | 0.086 |
| ENSMUSG00000030742 | Lat | 0.88 | 0.05 | 13.43 | $2.5 \mathrm{E}-04$ | 0.086 |
| ENSMUSG0000048109 | Rbm15 | 0.58 | 2.76 | 13.33 | $2.6 \mathrm{E}-04$ | 0.086 |
| ENSMUSG00000050370 | Ch25h | 1.33 | -1.74 | 13.45 | $2.5 \mathrm{E}-04$ | 0.086 |
| ENSMUSG0000032265 | Fam46a | 0.52 | 2.63 | 13.15 | $2.9 \mathrm{E}-04$ | 0.093 |

Table 2.2

| GenelD | Gene Name | logFC | logCPM | LR | p-value | FDR |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000021250 | Fos | 2.34 | 4.13 | 374.87 | 1.6E-83 | 2.9E-79 |
| ENSMUSG00000037868 | Egr2 | 2.62 | 2.04 | 266.52 | 6.5E-60 | 5.8E-56 |
| ENSMUSG00000020423 | Btg2 | 0.78 | 4.65 | 231.73 | 2.5E-52 | 1.5E-48 |
| ENSMUSG00000085609 | 1700016P03Rik | 1.24 | 2.34 | 189.85 | 3.4E-43 | 1.5E-39 |
| ENSMUSG00000024042 | Sik1 | 1.05 | 3.89 | 155.56 | 1.1E-35 | 3.8E-32 |
| ENSMUSG00000022602 | Arc | 1.48 | 7.48 | 147.89 | 5.0E-34 | 1.5E-30 |
| ENSMUSG00000034640 | Tiparp | 0.61 | 4.08 | 144.43 | 2.9E-33 | 7.4E-30 |
| ENSMUSG00000028195 | Cyr61 | 1.06 | 2.49 | 143.49 | 4.6E-33 | 1.0E-29 |
| ENSMUSG00000052837 | Junb | 0.97 | 5.94 | 134.12 | 5.2E-31 | 1.0E-27 |
| ENSMUSG00000003545 | Fosb | 0.93 | 3.45 | 127.11 | 1.8E-29 | 3.2E-26 |
| ENSMUSG00000079436 | Kcnj13 | 2.87 | -1.25 | 120.87 | 4.1E-28 | 6.7E-25 |
| ENSMUSG00000053560 | ler2 | 0.91 | 3.11 | 119.73 | 7.3E-28 | 1.1E-24 |
| ENSMUSG00000046470 | Sox18 | -0.69 | 3.20 | 117.07 | 2.8E-27 | 3.8E-24 |
| ENSMUSG00000036151 | Tm6sf2 | 0.81 | 2.44 | 110.24 | 8.7E-26 | 1.1E-22 |
| ENSMUSG00000023034 | Nr4a1 | 0.94 | 6.43 | 104.47 | 1.6E-24 | 1.9E-21 |
| ENSMUSG00000034936 | Arl4d | 0.68 | 3.37 | 87.74 | 7.4E-21 | 8.4E-18 |
| ENSMUSG00000024190 | Dusp1 | 0.90 | 4.37 | 86.70 | 1.3E-20 | 1.3E-17 |
| ENSMUSG00000061808 | Ttr | 3.88 | 7.15 | 85.27 | 2.6E-20 | 2.6E-17 |
| ENSMUSG00000071341 | Egr4 | 0.83 | 5.02 | 85.12 | 2.8E-20 | 2.7E-17 |
| ENSMUSG00000037447 | Arid5a | 0.50 | 3.72 | 72.76 | 1.5E-17 | 1.3E-14 |
| ENSMUSG00000090698 | Apold1 | 0.82 | 2.39 | 71.24 | 3.2E-17 | 2.7E-14 |
| ENSMUSG00000024793 | Tnfrsf25 | 0.69 | 5.54 | 69.63 | 7.2E-17 | 5.9E-14 |
| ENSMUSG00000015652 | Steap1 | 1.88 | -1.11 | 68.96 | 1.0E-16 | 7.8E-14 |
| ENSMUSG00000065537 | Mir132 | 1.21 | 0.27 | 66.12 | 4.2E-16 | 3.2E-13 |
| ENSMUSG00000055148 | Klf2 | 0.71 | 3.37 | 63.78 | 1.4E-15 | 1.0E-12 |
| ENSMUSG00000048546 | Tob2 | 0.49 | 4.75 | 60.79 | 6.4E-15 | 4.4E-12 |
| ENSMUSG00000034739 | Mfrp | 3.06 | 0.41 | 53.33 | 2.8E-13 | 1.9E-10 |
| ENSMUSG00000068323 | Slc4a5 | 3.09 | 0.48 | 50.60 | 1.1E-12 | 7.3E-10 |
| ENSMUSG00000032265 | Fam46a | 0.53 | 2.63 | 50.19 | 1.4E-12 | 8.7E-10 |
| ENSMUSG00000038612 | Mcl1 | 0.25 | 6.24 | 49.78 | 1.7E-12 | 1.0E-09 |
| ENSMUSG00000019970 | Sgk1 | 0.35 | 6.04 | 47.48 | 5.6E-12 | 3.2E-09 |
| ENSMUSG00000017418 | Arl5b | 0.32 | 4.59 | 47.36 | 5.9E-12 | 3.3E-09 |
| ENSMUSG00000020893 | Per1 | 0.46 | 6.52 | 47.22 | 6.4E-12 | 3.5E-09 |
| ENSMUSG00000040287 | Stac3 | 0.65 | 1.95 | 47.01 | 7.0E-12 | 3.7E-09 |
| ENSMUSG00000034765 | Dusp5 | 0.64 | 4.84 | 45.51 | 1.5E-11 | 7.8E-09 |
| ENSMUSG00000026051 | 1500015O10Rik | 1.47 | 1.71 | 44.61 | $2.4 \mathrm{E}-11$ | 1.2E-08 |
| ENSMUSG00000053819 | Camk2d | -0.39 | 5.31 | 44.15 | 3.0E-11 | 1.5E-08 |
| ENSMUSG00000039672 | Kcne2 | 2.81 | 0.33 | 43.84 | 3.6E-11 | 1.7E-08 |
| ENSMUSG00000020108 | Ddit4 | 0.42 | 5.89 | 43.12 | 5.1E-11 | 2.4E-08 |
| ENSMUSG00000048109 | Rbm15 | 0.47 | 2.76 | 42.48 | 7.1E-11 | 3.2E-08 |
| ENSMUSG00000028967 | Errfi1 | 0.32 | 5.45 | 42.13 | 8.5E-11 | 3.7E-08 |
| ENSMUSG00000017723 | Wfdc2 | 2.20 | -1.27 | 41.99 | 9.2E-11 | 3.9E-08 |
| ENSMUSG00000049516 | Spty2d1 | 0.28 | 4.40 | 41.74 | 1.0E-10 | 4.4E-08 |
| ENSMUSG00000004655 | Aqp1 | 2.09 | 0.69 | 40.85 | 1.6E-10 | 6.7E-08 |
| ENSMUSG00000084088 | Gm12941 | 0.99 | -0.02 | 40.62 | 1.8E-10 | 7.4E-08 |
| ENSMUSG00000047230 | Cldn2 | 1.51 | 0.46 | 40.47 | 2.0E-10 | 7.8E-08 |
| ENSMUSG00000037086 | Prr32 | 2.66 | -1.05 | 39.74 | $2.9 \mathrm{E}-10$ | 1.1E-07 |
| ENSMUSG00000025019 | Lcor | 0.65 | 1.14 | 38.76 | $4.8 \mathrm{E}-10$ | 1.8E-07 |


| ENSMUSG00000040536 | Necab1 | -0.37 | 5.34 | 38.66 | 5.0E-10 | 1.9E-07 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000024136 | Dnase112 | 0.45 | 3.02 | 37.61 | 8.6E-10 | 3.1E-07 |
| ENSMUSG00000025488 | Cox8b | 1.65 | -1.50 | 37.63 | 8.6E-10 | 3.1E-07 |
| ENSMUSG00000061436 | Hipk2 | 0.46 | 4.18 | 37.39 | 9.7E-10 | 3.4E-07 |
| ENSMUSG00000001827 | Folr1 | 1.85 | 1.14 | 37.16 | 1.1E-09 | 3.7E-07 |
| ENSMUSG00000067833 | 2900097C17Rik | -0.28 | 4.57 | 37.12 | 1.1E-09 | 3.7E-07 |
| ENSMUSG00000021848 | Otx2 | 1.95 | 0.42 | 36.54 | 1.5E-09 | 4.9E-07 |
| ENSMUSG00000035828 | Pim3 | 0.26 | 5.10 | 36.07 | 1.9E-09 | 6.1E-07 |
| ENSMUSG00000021025 | Nfkbia | 0.33 | 4.34 | 36.03 | 1.9E-09 | 6.1E-07 |
| ENSMUSG00000028125 | Abca4 | 1.15 | 1.78 | 35.92 | 2.1E-09 | 6.4E-07 |
| ENSMUSG00000021453 | Gadd45g | 0.35 | 4.05 | 35.20 | 3.0E-09 | 9.1E-07 |
| ENSMUSG00000028348 | Murc | -0.90 | -0.06 | 35.17 | 3.0E-09 | 9.1E-07 |
| ENSMUSG00000016024 | Lbp | 0.72 | 2.51 | 33.70 | 6.4E-09 | 1.9E-06 |
| ENSMUSG00000026579 | F5 | 2.05 | 1.45 | 33.71 | 6.4E-09 | 1.9E-06 |
| ENSMUSG00000008384 | Sertad1 | 0.43 | 2.92 | 33.51 | 7.1E-09 | 2.0E-06 |
| ENSMUSG00000025450 | Gm9752 | -0.48 | 1.94 | 32.93 | 9.5E-09 | 2.7E-06 |
| ENSMUSG00000038268 | Ovca2 | 0.36 | 3.70 | 32.77 | 1.0E-08 | 2.8E-06 |
| ENSMUSG00000044595 | Dnd1 | 0.63 | 1.20 | 32.79 | 1.0E-08 | 2.8E-06 |
| ENSMUSG00000022949 | Clic6 | 2.22 | 2.25 | 31.79 | 1.7E-08 | 4.6E-06 |
| ENSMUSG00000048489 | 8430408G22Rik | 0.95 | -0.04 | 31.47 | 2.0E-08 | 5.4E-06 |
| ENSMUSG00000038418 | Egr1 | 0.57 | 7.19 | 31.07 | 2.5E-08 | 6.5E-06 |
| ENSMUSG00000037573 | Tob1 | 0.33 | 5.11 | 30.80 | 2.9E-08 | 7.3E-06 |
| ENSMUSG00000056364 | Six3os1 | 1.13 | -1.11 | 30.80 | 2.9E-08 | 7.3E-06 |
| ENSMUSG00000045314 | Sowahb | -0.46 | 2.00 | 30.45 | 3.4E-08 | 8.4E-06 |
| ENSMUSG00000047648 | Fbxo30 | 0.40 | 2.46 | 30.47 | 3.4E-08 | 8.4E-06 |
| ENSMUSG00000014747 | Ankrd53 | 0.59 | 0.93 | 29.90 | 4.6E-08 | 1.1E-05 |
| ENSMUSG00000049907 | Rasl11b | 0.24 | 6.01 | 29.77 | 4.9E-08 | 1.2E-05 |
| ENSMUSG00000032501 | Trib1 | 0.38 | 3.57 | 29.68 | 5.1E-08 | 1.2E-05 |
| ENSMUSG00000058626 | Capn11 | 2.17 | 1.39 | 29.62 | 5.3E-08 | 1.2E-05 |
| ENSMUSG00000026628 | Atf3 | 0.74 | 0.57 | 29.28 | 6.3E-08 | 1.4E-05 |
| ENSMUSG00000090622 | A930033H14Rik | -0.55 | 1.09 | 28.89 | 7.7E-08 | 1.7E-05 |
| ENSMUSG00000030450 | Oca2 | 1.14 | -1.09 | 28.76 | 8.2E-08 | 1.8E-05 |
| ENSMUSG00000047867 | Gimap6 | 0.48 | 1.69 | 28.19 | 1.1E-07 | 2.4E-05 |
| ENSMUSG00000074825 | Itpripl1 | 0.58 | 1.12 | 28.14 | 1.1E-07 | 2.5E-05 |
| ENSMUSG00000034342 | Cbl | 0.38 | 3.17 | 28.08 | 1.2E-07 | 2.5E-05 |
| ENSMUSG00000062116 | Zfp954 | 0.29 | 3.78 | 28.04 | 1.2E-07 | 2.5E-05 |
| ENSMUSG00000090338 | Gm17081 | -0.89 | -0.39 | 28.06 | 1.2E-07 | 2.5E-05 |
| ENSMUSG00000066150 | Slc31a1 | 0.27 | 4.50 | 27.92 | 1.3E-07 | 2.6E-05 |
| ENSMUSG00000048001 | Hes5 | -0.43 | 2.32 | 27.87 | 1.3E-07 | 2.7E-05 |
| ENSMUSG00000049382 | Krt8 | 1.21 | -0.41 | 27.76 | 1.4E-07 | 2.8E-05 |
| ENSMUSG00000070780 | Rbm47 | 0.98 | -0.66 | 27.54 | 1.5E-07 | 3.1E-05 |
| ENSMUSG00000005268 | Prlr | 0.94 | 1.79 | 27.49 | 1.6E-07 | 3.2E-05 |
| ENSMUSG00000047604 | Frat2 | 0.32 | 3.60 | 27.36 | 1.7E-07 | 3.3E-05 |
| ENSMUSG00000015312 | Gadd45b | 0.47 | 3.49 | 27.32 | 1.7E-07 | 3.3E-05 |
| ENSMUSG00000021367 | Edn1 | -0.65 | 0.85 | 27.32 | 1.7E-07 | 3.3E-05 |
| ENSMUSG00000025350 | Rdh5 | 0.67 | 1.60 | 27.28 | 1.8E-07 | 3.4E-05 |
| ENSMUSG00000090986 | Gm17275 | 0.45 | 1.79 | 26.99 | 2.0E-07 | 3.9E-05 |
| ENSMUSG00000084917 | Gm17477 | 1.19 | -1.12 | 26.97 | 2.1E-07 | 3.9E-05 |
| ENSMUSG00000025049 | Taf5 | 0.36 | 2.89 | 26.88 | 2.2E-07 | 4.0E-05 |
| ENSMUSG00000020482 | Ccdc117 | 0.33 | 4.19 | 26.85 | 2.2E-07 | 4.0E-05 |
| ENSMUSG00000025255 | Zfhx4 | 0.34 | 3.29 | 26.08 | 3.3E-07 | 6.0E-05 |


| ENSMUSG00000034538 | Zfp418 | 0.45 | 1.73 | 26.04 | 3.3E-07 | 6.0E-05 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000022893 | Adamts1 | 0.32 | 4.49 | 25.96 | 3.5E-07 | 6.2E-05 |
| ENSMUSG00000038775 | Vill | -0.59 | 1.23 | 25.89 | 3.6E-07 | 6.4E-05 |
| ENSMUSG00000001467 | Cyp51 | 0.16 | 6.01 | 25.42 | 4.6E-07 | 8.1E-05 |
| ENSMUSG00000074890 | Lcmt2 | -0.32 | 3.05 | 25.09 | 5.5E-07 | 9.5E-05 |
| ENSMUSG00000086331 | Gm16310 | 1.07 | -1.14 | 24.79 | 6.4E-07 | 1.1E-04 |
| ENSMUSG00000085779 | Atcayos | 0.44 | 1.99 | 24.71 | 6.7E-07 | 1.1E-04 |
| ENSMUSG00000020385 | Clk4 | 0.19 | 6.11 | 24.30 | 8.2E-07 | 1.4E-04 |
| ENSMUSG00000091448 | Gm17388 | 0.37 | 2.28 | 24.31 | 8.2E-07 | 1.4E-04 |
| ENSMUSG00000020641 | Rsad2 | -0.62 | 0.49 | 24.28 | 8.4E-07 | 1.4E-04 |
| ENSMUSG00000021986 | Amer2 | 0.29 | 3.90 | 24.02 | 9.5E-07 | 1.6E-04 |
| ENSMUSG00000050930 | Map10 | -0.35 | 2.39 | 24.00 | 9.6E-07 | 1.6E-04 |
| ENSMUSG00000034295 | Fhod3 | -0.27 | 3.79 | 23.84 | 1.0E-06 | 1.7E-04 |
| ENSMUSG00000024383 | Map3k2 | 0.22 | 4.77 | 23.64 | 1.2E-06 | 1.9E-04 |
| ENSMUSG00000032515 | Csrnp1 | 0.48 | 3.40 | 23.60 | 1.2E-06 | 1.9E-04 |
| ENSMUSG00000026565 | Pou2f1 | 0.26 | 3.70 | 23.50 | 1.3E-06 | 2.0E-04 |
| ENSMUSG00000091070 | Gm3764 | -0.34 | 3.62 | 23.42 | 1.3E-06 | 2.0E-04 |
| ENSMUSG00000034161 | Scx | 0.63 | 0.50 | 23.30 | 1.4E-06 | 2.1E-04 |
| ENSMUSG00000034855 | Cxcl10 | -1.35 | -1.46 | 23.26 | 1.4E-06 | 2.2E-04 |
| ENSMUSG00000018604 | Tbx3 | -0.49 | 1.56 | 23.12 | 1.5E-06 | 2.3E-04 |
| ENSMUSG00000048379 | Socs4 | 0.26 | 3.61 | 22.99 | 1.6E-06 | 2.4E-04 |
| ENSMUSG00000030031 | Kbtbd8 | 0.35 | 2.46 | 22.87 | 1.7E-06 | 2.6E-04 |
| ENSMUSG00000022425 | Enpp2 | 0.85 | 7.78 | 22.80 | 1.8E-06 | 2.6E-04 |
| ENSMUSG00000079681 | Zglp1 | 0.62 | 0.79 | 22.79 | 1.8E-06 | 2.6E-04 |
| ENSMUSG00000074170 | Plekhf1 | 0.46 | 1.76 | 22.71 | 1.9E-06 | 2.7E-04 |
| ENSMUSG00000044177 | Wfikkn2 | 0.91 | 0.78 | 22.62 | 2.0E-06 | 2.8E-04 |
| ENSMUSG00000023043 | Krt18 | 1.44 | 0.13 | 22.54 | 2.1E-06 | 2.9E-04 |
| ENSMUSG00000041308 | Sntb2 | 0.33 | 2.58 | 22.43 | 2.2E-06 | 3.1E-04 |
| ENSMUSG00000010492 | Uckl1os | 0.71 | -0.11 | 22.32 | 2.3E-06 | 3.2E-04 |
| ENSMUSG00000052387 | Trpm3 | 0.29 | 4.65 | 22.12 | 2.6E-06 | 3.6E-04 |
| ENSMUSG00000042256 | Ptchd4 | 0.38 | 2.03 | 22.10 | 2.6E-06 | 3.6E-04 |
| ENSMUSG00000083929 | Gm10600 | -0.91 | 1.70 | 22.06 | 2.6E-06 | 3.6E-04 |
| ENSMUSG00000091542 | Gm17167 | -0.77 | 0.76 | 21.64 | 3.3E-06 | 4.5E-04 |
| ENSMUSG00000000392 | Fap | 0.79 | -0.52 | 21.50 | 3.5E-06 | 4.8E-04 |
| ENSMUSG00000037490 | Slc2a12 | 0.51 | 2.03 | 21.31 | 3.9E-06 | 5.3E-04 |
| ENSMUSG00000020681 | Ace | 0.72 | 3.67 | 21.29 | 3.9E-06 | 5.3E-04 |
| ENSMUSG00000025981 | Coq10b | 0.24 | 3.78 | 21.27 | 4.0E-06 | 5.3E-04 |
| ENSMUSG00000046962 | Zbtb21 | 0.22 | 4.56 | 21.25 | 4.0E-06 | 5.3E-04 |
| ENSMUSG00000034320 | Slc26a2 | 0.30 | 3.15 | 21.22 | 4.1E-06 | 5.3E-04 |
| ENSMUSG00000036834 | Plch1 | -0.38 | 2.85 | 21.21 | 4.1E-06 | 5.3E-04 |
| ENSMUSG00000002325 | Irf9 | 0.27 | 3.98 | 21.13 | 4.3E-06 | 5.5E-04 |
| ENSMUSG00000026034 | Clk1 | 0.21 | 6.92 | 21.06 | 4.4E-06 | 5.7E-04 |
| ENSMUSG00000057716 | Tmem178b | 0.34 | 3.12 | 21.04 | 4.5E-06 | 5.7E-04 |
| ENSMUSG00000024232 | Bambi | 0.29 | 3.09 | 20.92 | 4.8E-06 | 6.0E-04 |
| ENSMUSG00000021670 | Hmgcr | 0.16 | 6.66 | 20.79 | 5.1E-06 | 6.4E-04 |
| ENSMUSG00000022507 | 1810013L24Rik | 0.20 | 5.47 | 20.76 | 5.2E-06 | 6.5E-04 |
| ENSMUSG00000043953 | Ccrl2 | -0.71 | -0.03 | 20.69 | 5.4E-06 | 6.7E-04 |
| ENSMUSG00000036169 | Sostdc1 | 0.95 | 2.05 | 20.45 | 6.1E-06 | 7.5E-04 |
| ENSMUSG00000063354 | Slc39a4 | 0.65 | 0.14 | 20.30 | 6.6E-06 | 8.0E-04 |
| ENSMUSG00000055235 | Wdr86 | 1.26 | 0.10 | 20.24 | 6.8E-06 | 8.2E-04 |
| ENSMUSG00000056553 | Ptprn2 | -0.13 | 8.54 | 20.17 | 7.1E-06 | 8.5E-04 |


| ENSMUSG00000072568 | Fam84b | 0.31 | 4.17 | 20.15 | 7.1E-06 | 8.5E-04 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000043872 | Zmym1 | 0.28 | 4.01 | 19.89 | 8.2E-06 | 9.7E-04 |
| ENSMUSG00000082984 | RP23-304I1.8 | -0.86 | -0.84 | 19.85 | 8.4E-06 | 9.9E-04 |
| ENSMUSG00000039634 | Zfp189 | 0.36 | 4.47 | 19.68 | 9.1E-06 | 0.001 |
| ENSMUSG00000046922 | Gpr6 | -0.70 | -0.15 | 19.69 | 9.1E-06 | 0.001 |
| ENSMUSG00000048450 | Msx1 | 0.62 | 2.67 | 19.69 | 9.1E-06 | 0.001 |
| ENSMUSG00000032840 | 2410131K14Rik | 0.17 | 5.03 | 19.66 | 9.2E-06 | 0.001 |
| ENSMUSG00000027895 | Kcnc4 | -0.18 | 6.38 | 19.64 | 9.3E-06 | 0.001 |
| ENSMUSG00000089857 | Zfp882 | 0.34 | 2.28 | 19.60 | 9.5E-06 | 0.001 |
| ENSMUSG00000025402 | Nab2 | 0.25 | 4.85 | 19.56 | 9.7E-06 | 0.001 |
| ENSMUSG00000056592 | Zfp658 | 0.29 | 3.27 | 19.48 | 1.0E-05 | 0.001 |
| ENSMUSG00000020173 | Cobl | -0.38 | 4.47 | 19.33 | 1.1E-05 | 0.001 |
| ENSMUSG00000019960 | Dusp6 | 0.29 | 5.91 | 19.29 | 1.1E-05 | 0.001 |
| ENSMUSG00000047777 | Phf13 | 0.32 | 4.21 | 19.20 | 1.2E-05 | 0.001 |
| ENSMUSG00000044641 | Pard6b | 0.34 | 2.23 | 19.10 | 1.2E-05 | 0.001 |
| ENSMUSG00000079737 | 3110001I22Rik | 0.56 | 0.56 | 19.09 | 1.2E-05 | 0.001 |
| ENSMUSG00000049420 | Tmem200a | -0.27 | 3.59 | 19.07 | 1.3E-05 | 0.001 |
| ENSMUSG00000024298 | Zfp871 | 0.34 | 3.24 | 19.03 | 1.3E-05 | 0.001 |
| ENSMUSG00000078651 | Aoc2 | 0.29 | 3.07 | 18.90 | $1.4 \mathrm{E}-05$ | 0.001 |
| ENSMUSG00000051951 | Xkr4 | 0.29 | 3.06 | 18.89 | 1.4E-05 | 0.001 |
| ENSMUSG00000001763 | Tspan33 | 0.20 | 5.34 | 18.80 | 1.5E-05 | 0.002 |
| ENSMUSG00000084381 | AA413626 | -0.83 | -0.97 | 18.77 | $1.5 \mathrm{E}-05$ | 0.002 |
| ENSMUSG00000040035 | Disp2 | -0.09 | 9.15 | 18.73 | 1.5E-05 | 0.002 |
| ENSMUSG00000053137 | Mapk11 | -0.18 | 5.28 | 18.64 | $1.6 \mathrm{E}-05$ | 0.002 |
| ENSMUSG00000067629 | Syngap1 | -0.16 | 7.52 | 18.63 | 1.6E-05 | 0.002 |
| ENSMUSG00000042608 | Stk40 | 0.20 | 5.21 | 18.57 | 1.6E-05 | 0.002 |
| ENSMUSG00000038393 | Txnip | 0.32 | 4.14 | 18.52 | 1.7E-05 | 0.002 |
| ENSMUSG00000015467 | Egfl8 | 0.75 | -0.18 | 18.40 | 1.8E-05 | 0.002 |
| ENSMUSG00000033191 | Tie1 | -0.22 | 3.98 | 18.41 | 1.8E-05 | 0.002 |
| ENSMUSG00000068196 | Col8a1 | 1.24 | 0.44 | 18.41 | 1.8E-05 | 0.002 |
| ENSMUSG00000073437 | D330041H03Rik | -1.02 | -1.39 | 18.41 | 1.8E-05 | 0.002 |
| ENSMUSG00000030409 | Dmpk | 0.21 | 4.74 | 18.35 | 1.8E-05 | 0.002 |
| ENSMUSG00000031659 | Adcy7 | 0.31 | 2.81 | 18.36 | 1.8E-05 | 0.002 |
| ENSMUSG00000022376 | Adcy8 | -0.21 | 4.63 | 18.26 | $1.9 \mathrm{E}-05$ | 0.002 |
| ENSMUSG00000087598 | Zfp111 | 0.25 | 3.12 | 18.26 | $1.9 \mathrm{E}-05$ | 0.002 |
| ENSMUSG00000064125 | BC068157 | -0.18 | 5.72 | 18.14 | 2.1E-05 | 0.002 |
| ENSMUSG00000030742 | Lat | 0.68 | 0.05 | 18.05 | 2.1E-05 | 0.002 |
| ENSMUSG00000022537 | Tmem44 | -0.23 | 5.97 | 17.98 | 2.2E-05 | 0.002 |
| ENSMUSG00000026686 | Lmx1a | 1.08 | -1.58 | 17.91 | 2.3E-05 | 0.002 |
| ENSMUSG00000005917 | Otx1 | 0.36 | 2.03 | 17.84 | $2.4 \mathrm{E}-05$ | 0.002 |
| ENSMUSG00000045648 | Vwc2l | -0.51 | 3.01 | 17.80 | $2.5 \mathrm{E}-05$ | 0.002 |
| ENSMUSG00000030775 | Trat1 | 0.94 | -1.45 | 17.78 | 2.5E-05 | 0.002 |
| ENSMUSG00000043091 | Tuba1c | 0.45 | 1.21 | 17.77 | 2.5E-05 | 0.002 |
| ENSMUSG00000030123 | Plxnd1 | -0.27 | 5.13 | 17.65 | 2.7E-05 | 0.002 |
| ENSMUSG00000050370 | Ch25h | 1.31 | -1.74 | 17.62 | 2.7E-05 | 0.002 |
| ENSMUSG00000067158 | Col4a4 | 0.73 | -0.67 | 17.54 | 2.8E-05 | 0.003 |
| ENSMUSG00000092492 | B230208B08Rik | -0.59 | 0.03 | 17.54 | $2.8 \mathrm{E}-05$ | 0.003 |
| ENSMUSG00000048442 | Smim5 | 0.57 | 0.35 | 17.53 | $2.8 \mathrm{E}-05$ | 0.003 |
| ENSMUSG00000037434 | Slc30a1 | 0.18 | 5.20 | 17.45 | 3.0E-05 | 0.003 |
| ENSMUSG00000000182 | Fgf23 | -0.99 | -1.42 | 17.38 | 3.1E-05 | 0.003 |
| ENSMUSG00000092558 | Med20 | 0.19 | 4.42 | 17.31 | 3.2E-05 | 0.003 |


| ENSMUSG00000056771 | Gm10010 | 0.42 | 1.22 | 17.24 | 3.3E-05 | 0.003 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000068452 | Duox2 | 1.07 | -1.66 | 17.20 | 3.4E-05 | 0.003 |
| ENSMUSG00000036432 | Siah2 | 0.24 | 3.99 | 17.18 | 3.4E-05 | 0.003 |
| ENSMUSG00000039853 | Trim14 | -1.05 | -1.45 | 17.17 | 3.4E-05 | 0.003 |
| ENSMUSG00000053113 | Socs3 | -0.51 | 0.61 | 17.16 | 3.4E-05 | 0.003 |
| ENSMUSG00000078739 | CT868723.17-2 | -0.73 | -0.32 | 17.16 | 3.4E-05 | 0.003 |
| ENSMUSG00000007812 | Zfp655 | 0.18 | 4.87 | 17.12 | 3.5E-05 | 0.003 |
| ENSMUSG00000040511 | Pvr | 0.24 | 3.37 | 17.08 | 3.6E-05 | 0.003 |
| ENSMUSG00000083579 | Gm15538 | 1.12 | -1.58 | 16.98 | 3.8E-05 | 0.003 |
| ENSMUSG00000037580 | Gch1 | 0.46 | 1.01 | 16.86 | 4.0E-05 | 0.003 |
| ENSMUSG00000020941 | Map3k14 | 0.37 | 2.60 | 16.84 | 4.1E-05 | 0.003 |
| ENSMUSG00000054893 | Zfp667 | 0.18 | 4.44 | 16.80 | 4.1E-05 | 0.003 |
| ENSMUSG00000055407 | Map6 | -0.13 | 6.98 | 16.81 | 4.1E-05 | 0.003 |
| ENSMUSG00000065485 | Mir219a-2 | -0.69 | -0.35 | 16.82 | 4.1E-05 | 0.003 |
| ENSMUSG00000052748 | Swt1 | 0.28 | 3.26 | 16.68 | 4.4E-05 | 0.004 |
| ENSMUSG00000054146 | Krt15 | 0.62 | 0.37 | 16.65 | 4.5E-05 | 0.004 |
| ENSMUSG00000034009 | Rxfp1 | -0.54 | 1.65 | 16.64 | 4.5E-05 | 0.004 |
| ENSMUSG00000040490 | Lrfn2 | -0.17 | 5.07 | 16.59 | 4.6E-05 | 0.004 |
| ENSMUSG00000001506 | Col1a1 | -0.28 | 3.51 | 16.56 | 4.7E-05 | 0.004 |
| ENSMUSG00000074221 | Zfp568 | 0.42 | 2.08 | 16.51 | 4.8E-05 | 0.004 |
| ENSMUSG00000060530 | A930017M01Rik | -0.28 | 2.40 | 16.48 | 4.9E-05 | 0.004 |
| ENSMUSG00000027204 | Fbn1 | 0.30 | 3.57 | 16.41 | 5.1E-05 | 0.004 |
| ENSMUSG00000027570 | Col9a3 | 0.50 | 4.18 | 15.98 | 6.4E-05 | 0.005 |
| ENSMUSG00000032640 | Chsy1 | 0.30 | 2.33 | 15.98 | 6.4E-05 | 0.005 |
| ENSMUSG00000073879 | Gm5859 | -0.77 | 0.46 | 15.99 | 6.4E-05 | 0.005 |
| ENSMUSG00000054493 | Gm9947 | -0.86 | -1.03 | 15.96 | 6.5E-05 | 0.005 |
| ENSMUSG00000020907 | Rcvrn | 0.96 | -1.05 | 15.91 | 6.6E-05 | 0.005 |
| ENSMUSG00000054986 | Sec14I3 | -0.56 | 0.58 | 15.91 | 6.6E-05 | 0.005 |
| ENSMUSG00000031786 | Ccdc135 | 0.78 | 2.10 | 15.87 | 6.8E-05 | 0.005 |
| ENSMUSG00000089896 | Hsn2 | 0.71 | -0.69 | 15.84 | 6.9E-05 | 0.005 |
| ENSMUSG00000056174 | Col8a2 | 0.77 | 1.97 | 15.78 | 7.1E-05 | 0.006 |
| ENSMUSG00000007877 | Tcap | 0.67 | -0.11 | 15.68 | 7.5E-05 | 0.006 |
| ENSMUSG00000035692 | Isg15 | -0.53 | 1.18 | 15.69 | 7.5E-05 | 0.006 |
| ENSMUSG00000024127 | Prepl | -0.14 | 7.90 | 15.67 | 7.5E-05 | 0.006 |
| ENSMUSG00000043993 | 2900052L18Rik | -0.31 | 2.15 | 15.66 | 7.6E-05 | 0.006 |
| ENSMUSG00000032712 | 2810474O19Rik | 0.22 | 3.50 | 15.64 | 7.7E-05 | 0.006 |
| ENSMUSG00000017417 | Plxdc1 | -0.34 | 3.87 | 15.48 | 8.3E-05 | 0.006 |
| ENSMUSG00000024812 | Tjp2 | -0.15 | 5.21 | 15.48 | 8.3E-05 | 0.006 |
| ENSMUSG00000087064 | Sap30bpos | 0.55 | 0.17 | 15.42 | 8.6E-05 | 0.006 |
| ENSMUSG00000010067 | Rassf1 | 0.30 | 2.43 | 15.38 | 8.8E-05 | 0.007 |
| ENSMUSG00000039252 | Lgi2 | -0.19 | 5.84 | 15.29 | 9.2E-05 | 0.007 |
| ENSMUSG00000049112 | Oxtr | -0.24 | 3.52 | 15.24 | 9.4E-05 | 0.007 |
| ENSMUSG00000020522 | Mfap3 | 0.18 | 4.94 | 15.23 | 9.5E-05 | 0.007 |
| ENSMUSG00000000214 | Th | 0.60 | 0.00 | 15.20 | 9.7E-05 | 0.007 |
| ENSMUSG00000032131 | Abcg4 | -0.12 | 6.54 | 15.20 | 9.6E-05 | 0.007 |
| ENSMUSG00000037211 | Spry1 | 0.21 | 3.68 | 15.11 | 1.0E-04 | 0.007 |
| ENSMUSG00000028268 | Gbp3 | -0.35 | 2.22 | 15.08 | 1.0E-04 | 0.007 |
| ENSMUSG00000043131 | Mob1a | 0.16 | 4.88 | 15.07 | 1.0E-04 | 0.007 |
| ENSMUSG00000045954 | Sdpr | 0.28 | 2.93 | 15.06 | 1.0E-04 | 0.008 |
| ENSMUSG00000035164 | Zc3h12c | 0.26 | 2.82 | 15.05 | 1.0E-04 | 0.008 |
| ENSMUSG00000020638 | Cmpk2 | -0.23 | 4.26 | 14.99 | 1.1E-04 | 0.008 |


| ENSMUSG00000053117 | E330013P04Rik | -0.45 | 0.89 | 14.99 | 1.1E-04 | 0.008 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000038587 | Akap12 | -0.20 | 4.65 | 14.95 | 1.1E-04 | 0.008 |
| ENSMUSG00000046341 | Gm11223 | -0.19 | 5.88 | 14.96 | 1.1E-04 | 0.008 |
| ENSMUSG00000049739 | Zfp646 | 0.15 | 4.98 | 14.90 | 1.1E-04 | 0.008 |
| ENSMUSG00000029718 | Pcolce | 0.30 | 3.05 | 14.87 | 1.1E-04 | 0.008 |
| ENSMUSG00000026322 | Htr4 | 0.29 | 3.60 | 14.86 | 1.2E-04 | 0.008 |
| ENSMUSG00000037171 | Nodal | -0.71 | -0.52 | 14.84 | 1.2E-04 | 0.008 |
| ENSMUSG00000055421 | Pcdh9 | -0.19 | 5.64 | 14.81 | 1.2E-04 | 0.008 |
| ENSMUSG00000057969 | Sema3b | 0.50 | 1.83 | 14.78 | 1.2E-04 | 0.008 |
| ENSMUSG00000086456 | Nudt16I1 | 0.33 | 1.75 | 14.77 | 1.2E-04 | 0.008 |
| ENSMUSG00000017400 | Stac2 | -0.14 | 6.66 | 14.75 | 1.2E-04 | 0.008 |
| ENSMUSG00000041112 | Elmo1 | -0.15 | 5.78 | 14.61 | 1.3E-04 | 0.009 |
| ENSMUSG00000092067 | A230107N01Rik | -0.44 | 0.80 | 14.58 | 1.3E-04 | 0.009 |
| ENSMUSG00000033731 | 3300002A11Rik | 0.65 | -0.34 | 14.52 | $1.4 \mathrm{E}-04$ | 0.009 |
| ENSMUSG00000019301 | Hsd17b1 | 0.87 | -1.04 | 14.50 | $1.4 \mathrm{E}-04$ | 0.009 |
| ENSMUSG00000046159 | Chrm3 | -0.19 | 4.88 | 14.50 | $1.4 \mathrm{E}-04$ | 0.009 |
| ENSMUSG00000051354 | Samd3 | 1.35 | -1.27 | 14.46 | $1.4 \mathrm{E}-04$ | 0.010 |
| ENSMUSG00000032009 | Sesn3 | 0.22 | 3.58 | 14.43 | 1.5E-04 | 0.010 |
| ENSMUSG00000039323 | Igfbp2 | 0.25 | 4.83 | 14.40 | 1.5E-04 | 0.010 |
| ENSMUSG00000026797 | Stxbp1 | -0.10 | 9.52 | 14.32 | 1.5E-04 | 0.010 |
| ENSMUSG00000036052 | Dnajb5 | 0.17 | 7.61 | 14.32 | 1.5E-04 | 0.010 |
| ENSMUSG00000063919 | Srrm4 | -0.15 | 5.67 | 14.33 | 1.5E-04 | 0.010 |
| ENSMUSG00000078202 | Nrarp | 0.22 | 4.45 | 14.26 | 1.6E-04 | 0.010 |
| ENSMUSG00000067017 | Gm3608 | -0.27 | 2.26 | 14.19 | 1.7E-04 | 0.011 |
| ENSMUSG00000023025 | Larp4 | 0.19 | 5.05 | 14.18 | 1.7E-04 | 0.011 |
| ENSMUSG00000021478 | Drd1a | -0.32 | 2.29 | 14.14 | 1.7E-04 | 0.011 |
| ENSMUSG00000030790 | Adm | 0.58 | -0.02 | 14.08 | 1.8E-04 | 0.011 |
| ENSMUSG00000051029 | Serpinb1b | 0.61 | -0.43 | 14.08 | 1.7E-04 | 0.011 |
| ENSMUSG00000033594 | Spata2l | 0.16 | 6.03 | 13.99 | 1.8E-04 | 0.012 |
| ENSMUSG00000065336 | Snora34 | 0.43 | 0.78 | 13.99 | $1.8 \mathrm{E}-04$ | 0.012 |
| ENSMUSG00000025986 | Slc39a10 | -0.16 | 6.88 | 13.97 | $1.9 \mathrm{E}-04$ | 0.012 |
| ENSMUSG00000032503 | Arpp21 | -0.20 | 7.41 | 13.98 | $1.9 \mathrm{E}-04$ | 0.012 |
| ENSMUSG00000041216 | Clvs1 | -0.21 | 3.56 | 13.96 | $1.9 \mathrm{E}-04$ | 0.012 |
| ENSMUSG00000087141 | Plcxd2 | -0.26 | 4.94 | 13.95 | $1.9 \mathrm{E}-04$ | 0.012 |
| ENSMUSG00000061894 | Zscan20 | 0.28 | 2.75 | 13.91 | $1.9 \mathrm{E}-04$ | 0.012 |
| ENSMUSG00000056749 | Nfil3 | 0.22 | 3.74 | 13.89 | 1.9E-04 | 0.012 |
| ENSMUSG00000028952 | Zbtb48 | 0.23 | 4.02 | 13.85 | 2.0E-04 | 0.012 |
| ENSMUSG00000005220 | Corin | 0.93 | -1.37 | 13.84 | 2.0E-04 | 0.012 |
| ENSMUSG00000014158 | Trpv4 | 0.67 | 1.36 | 13.83 | 2.0E-04 | 0.012 |
| ENSMUSG00000046402 | Rbp1 | 0.28 | 3.58 | 13.83 | $2.0 \mathrm{E}-04$ | 0.012 |
| ENSMUSG00000024014 | Pim1 | -0.44 | 0.70 | 13.80 | 2.0E-04 | 0.012 |
| ENSMUSG00000028211 | Trp53inp1 | 0.23 | 3.43 | 13.80 | 2.0E-04 | 0.012 |
| ENSMUSG00000045671 | Spred2 | -0.14 | 6.70 | 13.80 | 2.0E-04 | 0.012 |
| ENSMUSG00000052684 | Jun | 0.14 | 6.95 | 13.77 | 2.1E-04 | 0.013 |
| ENSMUSG00000050640 | Tmem150c | -0.12 | 6.31 | 13.75 | 2.1E-04 | 0.013 |
| ENSMUSG00000050840 | Cdh20 | -0.23 | 4.24 | 13.74 | 2.1E-04 | 0.013 |
| ENSMUSG00000081787 | Gm13991 | -0.25 | 2.72 | 13.75 | 2.1E-04 | 0.013 |
| ENSMUSG00000066647 | Gm5113 | -0.20 | 3.59 | 13.73 | 2.1E-04 | 0.013 |
| ENSMUSG00000007817 | Zmiz1 | -0.18 | 6.28 | 13.71 | 2.1E-04 | 0.013 |
| ENSMUSG00000044337 | Ackr3 | 0.23 | 4.10 | 13.69 | 2.2E-04 | 0.013 |
| ENSMUSG00000046562 | Unc119b | 0.16 | 5.38 | 13.66 | 2.2E-04 | 0.013 |


| ENSMUSG00000041193 | Pla2g5 | 0.93 | -0.61 | 13.65 | 2.2E-04 | 0.013 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000005583 | Mef2c | -0.15 | 6.86 | 13.59 | 2.3E-04 | 0.013 |
| ENSMUSG00000034390 | Cmip | -0.12 | 8.43 | 13.58 | 2.3E-04 | 0.013 |
| ENSMUSG00000042997 | Nhlrc3 | -0.20 | 3.70 | 13.57 | 2.3E-04 | 0.013 |
| ENSMUSG00000018932 | Map2k3 | 0.20 | 3.85 | 13.56 | 2.3E-04 | 0.013 |
| ENSMUSG00000044690 | NR_002887 | -0.29 | 2.10 | 13.53 | 2.3E-04 | 0.014 |
| ENSMUSG00000049657 | Zbtb5 | 0.19 | 3.73 | 13.53 | 2.3E-04 | 0.014 |
| ENSMUSG00000021448 | Shc3 | 0.22 | 3.07 | 13.49 | 2.4E-04 | 0.014 |
| ENSMUSG00000020654 | Adcy3 | -0.14 | 5.53 | 13.48 | 2.4E-04 | 0.014 |
| ENSMUSG00000029335 | Bmp3 | -0.43 | 0.81 | 13.46 | 2.4E-04 | 0.014 |
| ENSMUSG00000039953 | Clstn1 | -0.10 | 10.14 | 13.46 | 2.4E-04 | 0.014 |
| ENSMUSG00000050335 | Lgals3 | 0.51 | 0.36 | 13.46 | 2.4E-04 | 0.014 |
| ENSMUSG00000022861 | Dgkg | -0.15 | 7.07 | 13.42 | 2.5E-04 | 0.014 |
| ENSMUSG00000085667 | Gm12992 | -0.68 | -0.62 | 13.42 | 2.5E-04 | 0.014 |
| ENSMUSG00000037857 | Nufip2 | 0.20 | 4.48 | 13.40 | 2.5E-04 | 0.014 |
| ENSMUSG00000065608 | Mirlet7c-2 | -0.51 | 0.27 | 13.40 | 2.5E-04 | 0.014 |
| ENSMUSG00000091421 | Gm4202 | -0.16 | 6.40 | 13.40 | 2.5E-04 | 0.014 |
| ENSMUSG00000020846 | Fam101b | -0.23 | 4.01 | 13.30 | 2.6E-04 | 0.015 |
| ENSMUSG00000025946 | Pth2r | -0.72 | -0.83 | 13.31 | 2.6E-04 | 0.015 |
| ENSMUSG00000085941 | Gm11201 | 0.74 | -0.93 | 13.30 | 2.7E-04 | 0.015 |
| ENSMUSG00000028051 | Hen3 | -0.18 | 3.98 | 13.28 | 2.7E-04 | 0.015 |
| ENSMUSG00000028341 | Nr4a3 | 0.18 | 6.65 | 13.25 | 2.7E-04 | 0.015 |
| ENSMUSG00000030533 | Unc45a | 0.14 | 5.45 | 13.23 | 2.8E-04 | 0.015 |
| ENSMUSG00000087424 | 5730405015Rik | -0.56 | -0.22 | 13.19 | 2.8E-04 | 0.015 |
| ENSMUSG00000060427 | Zfp868 | 0.19 | 3.69 | 13.17 | 2.9E-04 | 0.016 |
| ENSMUSG00000026655 | Fam107b | 0.19 | 3.88 | 13.09 | 3.0E-04 | 0.016 |
| ENSMUSG00000042216 | Sgsm1 | -0.15 | 6.60 | 13.07 | 3.0E-04 | 0.016 |
| ENSMUSG00000033987 | Dnah17 | 0.33 | 1.56 | 13.04 | 3.0E-04 | 0.017 |
| ENSMUSG00000072663 | Spef2 | 0.32 | 1.64 | 13.04 | 3.0E-04 | 0.017 |
| ENSMUSG00000028354 | Fmn2 | -0.11 | 6.23 | 13.01 | 3.1E-04 | 0.017 |
| ENSMUSG00000032193 | Ldir | 0.24 | 4.84 | 13.01 | 3.1E-04 | 0.017 |
| ENSMUSG00000016918 | Sulf1 | 0.56 | 2.87 | 12.99 | 3.1E-04 | 0.017 |
| ENSMUSG00000036452 | Arhgap26 | -0.13 | 6.32 | 12.95 | 3.2E-04 | 0.017 |
| ENSMUSG00000065453 | Mirlet7d | -0.60 | -0.36 | 12.96 | 3.2E-04 | 0.017 |
| ENSMUSG00000024033 | Rsph1 | 0.42 | 2.48 | 12.92 | 3.3E-04 | 0.017 |
| ENSMUSG00000091318 | Gm5415 | -0.38 | 2.33 | 12.92 | 3.2E-04 | 0.017 |
| ENSMUSG00000026072 | Il1r1 | 0.26 | 3.90 | 12.88 | 3.3E-04 | 0.018 |
| ENSMUSG00000081989 | RP23-388P16.1 | -0.59 | -0.35 | 12.89 | 3.3E-04 | 0.018 |
| ENSMUSG00000025888 | Casp1 | -0.44 | 0.67 | 12.88 | 3.3E-04 | 0.018 |
| ENSMUSG00000036687 | Tmem184a | 0.97 | -1.91 | 12.87 | 3.3E-04 | 0.018 |
| ENSMUSG00000049625 | Tifab | -0.34 | 1.56 | 12.83 | 3.4E-04 | 0.018 |
| ENSMUSG00000020190 | Mknk2 | 0.18 | 5.66 | 12.82 | 3.4E-04 | 0.018 |
| ENSMUSG00000029202 | Pds5a | -0.14 | 4.86 | 12.81 | 3.4E-04 | 0.018 |
| ENSMUSG00000027568 | Ntsr1 | -0.27 | 2.41 | 12.78 | 3.5E-04 | 0.018 |
| ENSMUSG00000028060 | 2810403A07Rik | -0.16 | 5.82 | 12.79 | 3.5E-04 | 0.018 |
| ENSMUSG00000050069 | Grem2 | -0.17 | 4.15 | 12.79 | 3.5E-04 | 0.018 |
| ENSMUSG00000052558 | Gm9884 | -0.55 | -0.07 | 12.78 | 3.5E-04 | 0.018 |
| ENSMUSG00000023915 | Tnfrsf21 | -0.11 | 7.06 | 12.75 | 3.6E-04 | 0.018 |
| ENSMUSG00000026312 | Cdh7 | -0.30 | 1.93 | 12.74 | 3.6E-04 | 0.018 |
| ENSMUSG00000035262 | Amh | 0.37 | 1.39 | 12.71 | 3.6E-04 | 0.018 |
| ENSMUSG00000047878 | A4galt | -0.40 | 1.24 | 12.72 | 3.6E-04 | 0.018 |


| ENSMUSG00000017167 | Cntnap1 | -0.10 | 8.22 | 12.70 | 3.7E-04 | 0.019 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000025816 | Sec61a2 | -0.13 | 5.33 | 12.66 | 3.7E-04 | 0.019 |
| ENSMUSG00000090444 | D930048G16 | -0.42 | 0.65 | 12.65 | 3.8E-04 | 0.019 |
| ENSMUSG00000029605 | Oas1b | -0.72 | -0.86 | 12.64 | 3.8E-04 | 0.019 |
| ENSMUSG00000028182 | Lrriq3 | 0.70 | -0.86 | 12.61 | 3.8E-04 | 0.019 |
| ENSMUSG00000058488 | KI | 0.59 | 4.57 | 12.58 | 3.9E-04 | 0.019 |
| ENSMUSG00000039081 | Zfp503 | 0.30 | 2.24 | 12.58 | 3.9E-04 | 0.019 |
| ENSMUSG00000005533 | Igf1r | 0.20 | 3.49 | 12.49 | 4.1E-04 | 0.020 |
| ENSMUSG00000015484 | Fam163a | -0.17 | 4.10 | 12.51 | 4.0E-04 | 0.020 |
| ENSMUSG00000019737 | Syne4 | 0.27 | 2.28 | 12.51 | 4.1E-04 | 0.020 |
| ENSMUSG00000021303 | Gng4 | -0.17 | 4.89 | 12.49 | 4.1E-04 | 0.020 |
| ENSMUSG00000028172 | Tacr3 | -0.33 | 1.79 | 12.51 | 4.0E-04 | 0.020 |
| ENSMUSG00000034006 | Pqlc1 | 0.16 | 4.88 | 12.53 | 4.0E-04 | 0.020 |
| ENSMUSG00000035969 | Rusc2 | -0.11 | 7.16 | 12.52 | 4.0E-04 | 0.020 |
| ENSMUSG00000038415 | Foxq1 | -0.32 | 1.64 | 12.52 | 4.0E-04 | 0.020 |
| ENSMUSG00000044548 | Dact1 | 0.30 | 2.28 | 12.50 | 4.1E-04 | 0.020 |
| ENSMUSG00000051495 | Irf2bp2 | 0.18 | 5.22 | 12.53 | 4.0E-04 | 0.020 |
| ENSMUSG00000055200 | Sertad3 | 0.34 | 1.52 | 12.49 | 4.1E-04 | 0.020 |
| ENSMUSG00000067279 | Ppp1r3c | 0.20 | 5.88 | 12.48 | 4.1E-04 | 0.020 |
| ENSMUSG00000092526 | Gm17907 | -0.50 | 0.43 | 12.49 | 4.1E-04 | 0.020 |
| ENSMUSG00000027520 | Zdbf2 | 0.19 | 4.05 | 12.46 | 4.2E-04 | 0.020 |
| ENSMUSG00000085527 | Gm15535 | -0.37 | 1.08 | 12.45 | 4.2E-04 | 0.020 |
| ENSMUSG00000062380 | Tubb3 | -0.14 | 8.14 | 12.44 | 4.2E-04 | 0.020 |
| ENSMUSG00000041483 | Zfp281 | 0.17 | 5.26 | 12.43 | 4.2E-04 | 0.020 |
| ENSMUSG00000038132 | Rbm24 | 0.18 | 4.09 | 12.43 | 4.2E-04 | 0.020 |
| ENSMUSG00000025577 | Cbx2 | 0.33 | 2.03 | 12.42 | 4.3E-04 | 0.020 |
| ENSMUSG00000085269 | Gm15777 | -0.47 | 0.29 | 12.42 | 4.3E-04 | 0.020 |
| ENSMUSG00000072763 | 5430403G16Rik | 0.35 | 1.13 | 12.37 | 4.4E-04 | 0.021 |
| ENSMUSG00000085906 | Gm16882 | -0.23 | 3.36 | 12.37 | 4.4E-04 | 0.021 |
| ENSMUSG00000047045 | Tmem164 | -0.18 | 4.34 | 12.36 | 4.4E-04 | 0.021 |
| ENSMUSG00000007682 | Dio2 | 0.22 | 5.62 | 12.34 | 4.4E-04 | 0.021 |
| ENSMUSG00000072294 | Klf12 | 0.21 | 3.31 | 12.34 | 4.4E-04 | 0.021 |
| ENSMUSG00000092060 | Bend4 | 0.20 | 3.44 | 12.34 | 4.4E-04 | 0.021 |
| ENSMUSG00000036766 | Dner | -0.12 | 7.12 | 12.31 | 4.5E-04 | 0.021 |
| ENSMUSG00000030157 | Clec2d | -0.34 | 1.85 | 12.30 | 4.5E-04 | 0.021 |
| ENSMUSG00000029352 | Crybb3 | 0.39 | 1.20 | 12.28 | 4.6E-04 | 0.021 |
| ENSMUSG00000039298 | Cdk5rap2 | 0.15 | 4.76 | 12.26 | 4.6E-04 | 0.021 |
| ENSMUSG00000014301 | Pam16 | 0.35 | 1.46 | 12.24 | 4.7E-04 | 0.021 |
| ENSMUSG00000034275 | Igsf9b | 0.45 | 0.55 | 12.23 | 4.7E-04 | 0.021 |
| ENSMUSG00000033253 | Szt2 | 0.16 | 5.98 | 12.23 | 4.7E-04 | 0.022 |
| ENSMUSG00000053693 | Mast1 | -0.14 | 5.91 | 12.22 | 4.7E-04 | 0.022 |
| ENSMUSG00000039431 | Mtmr7 | -0.12 | 5.64 | 12.19 | 4.8E-04 | 0.022 |
| ENSMUSG00000074394 | Vmn2r29 | -0.38 | 0.92 | 12.18 | 4.8E-04 | 0.022 |
| ENSMUSG00000092622 | Khdc3 | 0.49 | 0.48 | 12.17 | 4.8E-04 | 0.022 |
| ENSMUSG00000032172 | Olfm2 | -0.21 | 5.49 | 12.15 | 4.9E-04 | 0.022 |
| ENSMUSG00000090223 | Pcp4 | 0.40 | 2.84 | 12.14 | 4.9E-04 | 0.022 |
| ENSMUSG00000024304 | Cdh2 | -0.11 | 6.33 | 12.08 | 5.1E-04 | 0.023 |
| ENSMUSG00000032135 | Mcam | -0.14 | 5.04 | 12.07 | 5.1E-04 | 0.023 |
| ENSMUSG00000046191 | Pcdhb20 | -0.21 | 3.44 | 12.07 | 5.1E-04 | 0.023 |
| ENSMUSG00000026222 | Sp100 | -0.31 | 1.96 | 12.05 | 5.2E-04 | 0.023 |
| ENSMUSG00000021217 | Tshz3 | -0.27 | 3.68 | 12.03 | 5.2E-04 | 0.023 |


| ENSMUSG00000025902 | Sox17 | -0.25 | 2.39 | 12.02 | 5.3E-04 | 0.023 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000081855 | Rpl17-ps5 | -0.48 | 0.43 | 12.03 | 5.2E-04 | 0.023 |
| ENSMUSG00000020887 | A230052G05Rik | -0.96 | -1.57 | 12.01 | 5.3E-04 | 0.023 |
| ENSMUSG00000091969 | Pcdha3 | -0.34 | 1.25 | 12.01 | 5.3E-04 | 0.023 |
| ENSMUSG00000022377 | Asap1 | -0.15 | 5.75 | 11.98 | 5.4E-04 | 0.024 |
| ENSMUSG00000032246 | Calm14 | 0.76 | 0.88 | 11.98 | 5.4E-04 | 0.024 |
| ENSMUSG00000031167 | Rbm3 | 0.16 | 5.07 | 11.92 | 5.6E-04 | 0.024 |
| ENSMUSG00000027750 | Postn | 0.54 | 1.09 | 11.90 | 5.6E-04 | 0.024 |
| ENSMUSG00000026100 | Mstn | -0.83 | -1.58 | 11.89 | 5.7E-04 | 0.025 |
| ENSMUSG00000073875 | AL824709.35-2 | -1.05 | -1.95 | 11.87 | 5.7E-04 | 0.025 |
| ENSMUSG00000033685 | Ucp2 | 0.25 | 4.22 | 11.86 | 5.7E-04 | 0.025 |
| ENSMUSG00000039661 | Dusp26 | -0.19 | 5.70 | 11.82 | 5.9E-04 | 0.025 |
| ENSMUSG00000039813 | Tbc1d2 | 0.32 | 2.60 | 11.76 | 6.0E-04 | 0.026 |
| ENSMUSG00000048482 | Bdnf | 0.15 | 5.55 | 11.77 | 6.0E-04 | 0.026 |
| ENSMUSG00000063239 | Grm4 | -0.22 | 3.97 | 11.76 | 6.0E-04 | 0.026 |
| ENSMUSG00000018470 | Kcnab3 | -0.17 | 4.74 | 11.73 | $6.1 \mathrm{E}-04$ | 0.026 |
| ENSMUSG00000091793 | Rian | -0.16 | 8.88 | 11.73 | 6.1E-04 | 0.026 |
| ENSMUSG00000020032 | Nuak1 | -0.15 | 6.19 | 11.72 | 6.2E-04 | 0.026 |
| ENSMUSG00000016619 | Nup50 | 0.14 | 5.55 | 11.70 | 6.2E-04 | 0.026 |
| ENSMUSG00000018822 | Sfrp5 | 0.91 | -1.49 | 11.70 | 6.2E-04 | 0.026 |
| ENSMUSG00000075327 | Zbtb2 | 0.17 | 3.83 | 11.68 | 6.3E-04 | 0.027 |
| ENSMUSG00000029726 | Mepce | 0.15 | 5.71 | 11.65 | 6.4E-04 | 0.027 |
| ENSMUSG00000030748 | 114 ra | -0.27 | 2.36 | 11.63 | 6.5E-04 | 0.027 |
| ENSMUSG00000043929 | KIhl15 | 0.21 | 3.80 | 11.62 | 6.5E-04 | 0.027 |
| ENSMUSG00000061578 | Ksr2 | 0.35 | 1.98 | 11.62 | 6.5E-04 | 0.027 |
| ENSMUSG00000044676 | Zfp612 | -0.17 | 3.95 | 11.60 | 6.6E-04 | 0.027 |
| ENSMUSG00000055485 | Soga1 | 0.17 | 4.52 | 11.60 | 6.6E-04 | 0.027 |
| ENSMUSG00000087885 | Gm23600 | -0.50 | -0.05 | 11.60 | 6.6E-04 | 0.027 |
| ENSMUSG00000079362 | Gbp6 | -0.50 | 0.30 | 11.59 | 6.6E-04 | 0.027 |
| ENSMUSG00000049353 | Rd3 | 0.67 | -0.76 | 11.58 | 6.6E-04 | 0.028 |
| ENSMUSG00000028760 | Eif4g3 | -0.09 | 8.14 | 11.54 | 6.8E-04 | 0.028 |
| ENSMUSG00000063297 | Luzp2 | -0.21 | 4.72 | 11.51 | 6.9E-04 | 0.029 |
| ENSMUSG00000035711 | Dok3 | 0.20 | 3.83 | 11.50 | 7.0E-04 | 0.029 |
| ENSMUSG00000047330 | Kcne4 | 0.48 | 0.13 | 11.50 | 7.0E-04 | 0.029 |
| ENSMUSG00000022053 | Ebf2 | 1.03 | -1.85 | 11.49 | 7.0E-04 | 0.029 |
| ENSMUSG00000018412 | Kansl1 | -0.12 | 6.59 | 11.44 | 7.2E-04 | 0.029 |
| ENSMUSG00000027806 | Tsc22d2 | 0.14 | 5.12 | 11.43 | 7.2E-04 | 0.029 |
| ENSMUSG00000028645 | Slc2a1 | 0.13 | 6.32 | 11.41 | 7.3E-04 | 0.030 |
| ENSMUSG00000050164 | Mchr1 | 0.18 | 4.38 | 11.41 | 7.3E-04 | 0.030 |
| ENSMUSG00000071203 | Naip5 | -0.43 | 0.45 | 11.41 | 7.3E-04 | 0.030 |
| ENSMUSG00000022111 | Uchl3 | -0.15 | 4.42 | 11.39 | 7.4E-04 | 0.030 |
| ENSMUSG00000028803 | Nipal3 | -0.10 | 6.76 | 11.39 | 7.4E-04 | 0.030 |
| ENSMUSG00000038194 | Lhb | 0.58 | -0.32 | 11.34 | 7.6E-04 | 0.030 |
| ENSMUSG00000025006 | Sorbs1 | -0.12 | 5.79 | 11.33 | 7.6E-04 | 0.030 |
| ENSMUSG00000092099 | Gm17530 | 0.73 | -1.10 | 11.34 | 7.6E-04 | 0.030 |
| ENSMUSG00000052572 | Dlg2 | -0.11 | 7.19 | 11.31 | 7.7E-04 | 0.031 |
| ENSMUSG00000027079 | Clp1 | 0.22 | 2.94 | 11.31 | 7.7E-04 | 0.031 |
| ENSMUSG00000067578 | Cbln4 | -0.39 | 3.25 | 11.30 | 7.8E-04 | 0.031 |
| ENSMUSG00000086996 | 4933416E14Rik | 0.69 | -0.96 | 11.30 | 7.8E-04 | 0.031 |
| ENSMUSG00000087130 | D230004N17Rik | -0.27 | 3.71 | 11.30 | 7.7E-04 | 0.031 |
| ENSMUSG00000025404 | R3hdm2 | -0.12 | 7.61 | 11.28 | 7.8E-04 | 0.031 |


| ENSMUSG00000060733 | Ipmk | 0.16 | 5.66 | 11.29 | 7.8E-04 | 0.031 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000039834 | Zfp335 | -0.15 | 4.80 | 11.27 | 7.9E-04 | 0.031 |
| ENSMUSG00000018844 | Fndc8 | 0.29 | 2.37 | 11.26 | 7.9E-04 | 0.031 |
| ENSMUSG00000028420 | Tmem38b | 0.20 | 3.80 | 11.25 | 8.0E-04 | 0.031 |
| ENSMUSG00000035314 | Gdpd5 | -0.16 | 4.43 | 11.26 | 7.9E-04 | 0.031 |
| ENSMUSG00000026384 | Ptpn4 | 0.22 | 2.99 | 11.25 | 8.0E-04 | 0.031 |
| ENSMUSG00000059898 | Dsc3 | -0.57 | 1.20 | 11.24 | 8.0E-04 | 0.031 |
| ENSMUSG00000063179 | Pstk | -0.19 | 3.95 | 11.20 | 8.2E-04 | 0.032 |
| ENSMUSG00000038011 | Dnah10 | 0.31 | 1.84 | 11.19 | 8.2E-04 | 0.032 |
| ENSMUSG00000041263 | Rusc1 | -0.10 | 7.40 | 11.18 | 8.3E-04 | 0.032 |
| ENSMUSG00000073752 | Gm10570 | -0.72 | -1.05 | 11.16 | 8.3E-04 | 0.032 |
| ENSMUSG00000019865 | Nmbr | -0.36 | 0.96 | 11.15 | 8.4E-04 | 0.032 |
| ENSMUSG00000041378 | Cldn5 | -0.27 | 5.80 | 11.14 | 8.4E-04 | 0.032 |
| ENSMUSG00000085565 | Gm15721 | -0.45 | 0.54 | 11.12 | 8.5E-04 | 0.033 |
| ENSMUSG00000082329 | Gm14287 | -0.44 | 0.59 | 11.11 | 8.6E-04 | 0.033 |
| ENSMUSG00000029641 | Rasi11a | 0.24 | 3.37 | 11.09 | 8.7E-04 | 0.033 |
| ENSMUSG00000087478 | 4930506C21Rik | 0.36 | 0.99 | 11.08 | 8.7E-04 | 0.033 |
| ENSMUSG00000051331 | Cacna1c | 0.15 | 5.63 | 11.05 | 8.9E-04 | 0.034 |
| ENSMUSG00000031227 | Magee1 | -0.10 | 7.43 | 11.00 | 9.1E-04 | 0.035 |
| ENSMUSG00000042372 | Dmrt3 | 0.48 | 0.11 | 10.99 | 9.2E-04 | 0.035 |
| ENSMUSG00000039116 | Gpr126 | -0.42 | 0.51 | 10.98 | 9.2E-04 | 0.035 |
| ENSMUSG00000032122 | Slc37a2 | 0.33 | 1.59 | 10.98 | 9.2E-04 | 0.035 |
| ENSMUSG00000024044 | Epb4.113 | -0.10 | 7.34 | 10.95 | 9.4E-04 | 0.035 |
| ENSMUSG00000029401 | Rilpl2 | 0.20 | 3.88 | 10.95 | 9.3E-04 | 0.035 |
| ENSMUSG00000038135 | Crygn | 0.56 | -0.06 | 10.95 | 9.4E-04 | 0.035 |
| ENSMUSG00000064345 | mt-Nd2 | -0.14 | 11.70 | 10.95 | 9.4E-04 | 0.035 |
| ENSMUSG00000066798 | Zbtb6 | 0.17 | 3.82 | 10.96 | 9.3E-04 | 0.035 |
| ENSMUSG00000087674 | 4930447M23Rik | -0.26 | 2.29 | 10.96 | 9.3E-04 | 0.035 |
| ENSMUSG00000047141 | Zfp654 | 0.22 | 3.88 | 10.94 | 9.4E-04 | 0.035 |
| ENSMUSG00000048138 | Dmrt2 | -0.49 | 0.01 | 10.93 | 9.4E-04 | 0.035 |
| ENSMUSG00000013089 | Etv5 | -0.14 | 6.22 | 10.92 | 9.5E-04 | 0.035 |
| ENSMUSG00000031292 | Cdkl5 | 0.21 | 3.45 | 10.90 | 9.6E-04 | 0.035 |
| ENSMUSG00000036197 | Gxylt1 | 0.19 | 4.00 | 10.91 | 9.6E-04 | 0.035 |
| ENSMUSG00000062151 | Unc13c | -0.37 | 3.56 | 10.91 | 9.6E-04 | 0.035 |
| ENSMUSG00000035493 | Tgfbi | 0.31 | 2.26 | 10.87 | 9.8E-04 | 0.036 |
| ENSMUSG00000031609 | Sap30 | 0.27 | 2.58 | 10.85 | 9.9E-04 | 0.036 |
| ENSMUSG00000071035 | Gm5499 | -0.14 | 5.10 | 10.84 | 1.0E-03 | 0.036 |
| ENSMUSG00000051413 | Plagl2 | 0.19 | 3.46 | 10.80 | 0.001 | 0.037 |
| ENSMUSG00000025498 | Irf7 | -0.48 | 1.26 | 10.79 | 0.001 | 0.037 |
| ENSMUSG00000079164 | Tlr5 | -0.72 | -1.12 | 10.79 | 0.001 | 0.037 |
| ENSMUSG00000068966 | Zbtb34 | 0.18 | 3.83 | 10.78 | 0.001 | 0.037 |
| ENSMUSG00000019278 | Dpep1 | 1.00 | -1.91 | 10.76 | 0.001 | 0.037 |
| ENSMUSG00000021908 | Gm6768 | -0.21 | 2.98 | 10.76 | 0.001 | 0.037 |
| ENSMUSG00000005483 | Dnajb1 | 0.18 | 5.78 | 10.75 | 0.001 | 0.038 |
| ENSMUSG00000022307 | Oxr1 | -0.15 | 7.81 | 10.71 | 0.001 | 0.038 |
| ENSMUSG00000051212 | Gpr183 | -0.42 | 0.43 | 10.71 | 0.001 | 0.038 |
| ENSMUSG00000024277 | Mapre2 | -0.09 | 8.02 | 10.70 | 0.001 | 0.038 |
| ENSMUSG00000050567 | Maml1 | 0.19 | 3.95 | 10.70 | 0.001 | 0.038 |
| ENSMUSG00000018541 | Cwc25 | 0.16 | 4.22 | 10.70 | 0.001 | 0.038 |
| ENSMUSG00000044211 | Gm7887 | -0.18 | 3.70 | 10.67 | 0.001 | 0.039 |
| ENSMUSG00000047228 | BC048546 | -0.22 | 2.92 | 10.68 | 0.001 | 0.039 |


| ENSMUSG00000085440 | Sorbs2os | -0.32 | 1.19 | 10.67 | 0.001 | 0.039 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000045519 | Zfp560 | 0.22 | 3.05 | 10.66 | 0.001 | 0.039 |
| ENSMUSG00000025921 | Rdh10 | 0.17 | 3.68 | 10.64 | 0.001 | 0.039 |
| ENSMUSG00000085894 | Gm15832 | 0.30 | 1.75 | 10.63 | 0.001 | 0.039 |
| ENSMUSG00000089872 | Rps6kc1 | -0.13 | 5.64 | 10.63 | 0.001 | 0.039 |
| ENSMUSG00000063663 | Brwd3 | 0.22 | 2.99 | 10.59 | 0.001 | 0.040 |
| ENSMUSG00000051246 | Msantd1 | 0.27 | 1.98 | 10.58 | 0.001 | 0.040 |
| ENSMUSG00000050271 | D8Ertd82e | -0.17 | 5.00 | 10.56 | 0.001 | 0.040 |
| ENSMUSG00000041161 | Otud3 | 0.23 | 2.95 | 10.47 | 0.001 | 0.042 |
| ENSMUSG00000007655 | Cav1 | -0.19 | 3.81 | 10.45 | 0.001 | 0.043 |
| ENSMUSG00000032119 | Hinfp | 0.19 | 3.18 | 10.45 | 0.001 | 0.043 |
| ENSMUSG00000090061 | Nwd2 | -0.13 | 5.11 | 10.45 | 0.001 | 0.043 |
| ENSMUSG00000044636 | Csrnp2 | 0.15 | 5.02 | 10.44 | 0.001 | 0.043 |
| ENSMUSG00000085944 | 1700003D09Rik | 1.08 | -2.24 | 10.43 | 0.001 | 0.043 |
| ENSMUSG00000053153 | Spag16 | 0.61 | 0.42 | 10.42 | 0.001 | 0.043 |
| ENSMUSG00000086725 | A630052C17Rik | -0.25 | 2.25 | 10.39 | 0.001 | 0.044 |
| ENSMUSG00000080759 | Gm15573 | 0.73 | -1.27 | 10.37 | 0.001 | 0.044 |
| ENSMUSG00000025432 | Avil | 0.54 | -0.24 | 10.36 | 0.001 | 0.044 |
| ENSMUSG00000031027 | Stk33 | 0.37 | 0.85 | 10.36 | 0.001 | 0.044 |
| ENSMUSG00000081605 | Gm15953 | 0.87 | -1.72 | 10.30 | 0.001 | 0.046 |
| ENSMUSG00000045008 | 9030612E09Rik | -0.40 | 0.60 | 10.28 | 0.001 | 0.046 |
| ENSMUSG00000039830 | Olig2 | -0.21 | 5.04 | 10.26 | 0.001 | 0.046 |
| ENSMUSG00000049791 | Fzd4 | 0.23 | 2.20 | 10.26 | 0.001 | 0.046 |
| ENSMUSG00000090673 | Gm340 | 0.53 | -0.34 | 10.26 | 0.001 | 0.046 |
| ENSMUSG00000038702 | Dsel | -0.17 | 4.07 | 10.24 | 0.001 | 0.046 |
| ENSMUSG00000060441 | Trim5 | -0.59 | -0.74 | 10.23 | 0.001 | 0.047 |
| ENSMUSG00000007888 | Crlf1 | 0.13 | 5.44 | 10.20 | 0.001 | 0.047 |
| ENSMUSG00000044772 | Sntn | 0.51 | -0.11 | 10.16 | 0.001 | 0.048 |
| ENSMUSG00000092300 | Cdk3-ps | 0.32 | 1.40 | 10.09 | 0.001 | 0.050 |
| ENSMUSG00000069114 | Zbtb10 | 0.23 | 2.71 | 10.07 | 0.002 | 0.050 |
| ENSMUSG00000021196 | Pfkp | -0.11 | 7.90 | 10.07 | 0.002 | 0.051 |
| ENSMUSG00000045903 | Npas4 | 0.49 | 3.94 | 10.05 | 0.002 | 0.051 |
| ENSMUSG00000032579 | Hemk1 | 0.16 | 4.09 | 10.04 | 0.002 | 0.051 |
| ENSMUSG00000061397 | Krt79 | 0.78 | -1.23 | 10.01 | 0.002 | 0.052 |
| ENSMUSG00000038331 | Satb2 | -0.21 | 3.89 | 10.00 | 0.002 | 0.052 |
| ENSMUSG00000041351 | Rap1gap | -0.11 | 7.85 | 10.00 | 0.002 | 0.052 |
| ENSMUSG00000019850 | Tnfaip3 | 0.23 | 2.97 | 9.99 | 0.002 | 0.052 |
| ENSMUSG00000032860 | P2ry2 | 0.47 | -0.09 | 9.97 | 0.002 | 0.052 |
| ENSMUSG00000061532 | Zfp955b | -0.20 | 2.73 | 9.97 | 0.002 | 0.052 |
| ENSMUSG00000024049 | Myom1 | -0.29 | 1.67 | 9.96 | 0.002 | 0.053 |
| ENSMUSG00000069874 | Irgm2 | -0.27 | 2.07 | 9.95 | 0.002 | 0.053 |
| ENSMUSG00000000901 | Mmp11 | -0.32 | 1.26 | 9.94 | 0.002 | 0.053 |
| ENSMUSG00000027210 | Meis2 | -0.23 | 5.56 | 9.93 | 0.002 | 0.053 |
| ENSMUSG00000032556 | Bfsp2 | -0.27 | 2.02 | 9.93 | 0.002 | 0.053 |
| ENSMUSG00000065037 | Rn7sk | -1.37 | 0.61 | 9.93 | 0.002 | 0.053 |
| ENSMUSG00000056258 | Kcnq3 | 0.20 | 3.52 | 9.91 | 0.002 | 0.053 |
| ENSMUSG00000083889 | E530001F21Rik | -0.38 | 0.78 | 9.91 | 0.002 | 0.053 |
| ENSMUSG00000070639 | Lrrc8b | -0.14 | 4.71 | 9.87 | 0.002 | 0.054 |
| ENSMUSG00000078185 | Chml | -0.46 | 0.05 | 9.87 | 0.002 | 0.054 |
| ENSMUSG00000038291 | Snx25 | -0.14 | 5.73 | 9.86 | 0.002 | 0.055 |
| ENSMUSG00000024300 | Myo1f | -0.27 | 1.75 | 9.85 | 0.002 | 0.055 |


| ENSMUSG00000046062 | Ppp1r15b | 0.13 | 5.87 | 9.84 | 0.002 | 0.055 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000021483 | Cdk20 | 0.18 | 3.39 | 9.83 | 0.002 | 0.055 |
| ENSMUSG00000029309 | Sparcl1 | -0.11 | 10.84 | 9.82 | 0.002 | 0.055 |
| ENSMUSG00000038550 | Ciart | 0.19 | 3.22 | 9.81 | 0.002 | 0.056 |
| ENSMUSG00000032744 | Heyl | -0.25 | 2.53 | 9.78 | 0.002 | 0.057 |
| ENSMUSG00000022864 | D16Ertd472e | 0.25 | 2.07 | 9.77 | 0.002 | 0.057 |
| ENSMUSG00000040412 | 5330417C22Rik | -0.14 | 5.45 | 9.77 | 0.002 | 0.057 |
| ENSMUSG00000031748 | Gnao1 | -0.09 | 9.41 | 9.74 | 0.002 | 0.057 |
| ENSMUSG00000066956 | SC-144776 | 0.95 | -2.10 | 9.73 | 0.002 | 0.057 |
| ENSMUSG00000023017 | Asic1 | -0.13 | 5.35 | 9.72 | 0.002 | 0.058 |
| ENSMUSG00000021071 | Trim9 | -0.08 | 8.18 | 9.71 | 0.002 | 0.058 |
| ENSMUSG00000056962 | Jmjd6 | 0.15 | 4.97 | 9.71 | 0.002 | 0.058 |
| ENSMUSG00000057722 | Lepr | 0.35 | 0.74 | 9.70 | 0.002 | 0.058 |
| ENSMUSG00000050931 | Sgms2 | 0.29 | 1.79 | 9.68 | 0.002 | 0.059 |
| ENSMUSG00000021696 | Elovl7 | 0.23 | 3.98 | 9.67 | 0.002 | 0.059 |
| ENSMUSG00000045817 | Zfp3612 | -0.23 | 4.35 | 9.66 | 0.002 | 0.059 |
| ENSMUSG00000063281 | Zfp35 | 0.18 | 4.48 | 9.63 | 0.002 | 0.060 |
| ENSMUSG00000042589 | Cux2 | -0.18 | 4.77 | 9.62 | 0.002 | 0.060 |
| ENSMUSG00000049521 | Cdc42ep1 | -0.14 | 4.61 | 9.63 | 0.002 | 0.060 |
| ENSMUSG00000030500 | Slc17a6 | -0.37 | 4.00 | 9.62 | 0.002 | 0.060 |
| ENSMUSG00000020052 | Ascl1 | -0.23 | 2.90 | 9.61 | 0.002 | 0.060 |
| ENSMUSG00000001128 | Cfp | 0.16 | 4.29 | 9.60 | 0.002 | 0.060 |
| ENSMUSG00000017667 | Zfp334 | 0.13 | 4.77 | 9.59 | 0.002 | 0.060 |
| ENSMUSG00000028229 | Rmdn1 | -0.19 | 3.03 | 9.59 | 0.002 | 0.060 |
| ENSMUSG00000030330 | Ing4 | -0.11 | 5.54 | 9.59 | 0.002 | 0.060 |
| ENSMUSG00000036667 | Fam115a | -0.09 | 8.26 | 9.60 | 0.002 | 0.060 |
| ENSMUSG00000046572 | Zfp518b | 0.15 | 3.84 | 9.59 | 0.002 | 0.060 |
| ENSMUSG00000058922 | Gm10052 | -0.19 | 5.41 | 9.60 | 0.002 | 0.060 |
| ENSMUSG00000082665 | Gm11470 | 0.56 | -0.62 | 9.58 | 0.002 | 0.060 |
| ENSMUSG00000028753 | Vwa5b1 | 0.34 | 1.86 | 9.57 | 0.002 | 0.060 |
| ENSMUSG00000031562 | Dctd | -0.22 | 2.88 | 9.58 | 0.002 | 0.060 |
| ENSMUSG00000048696 | Mex3d | 0.22 | 3.73 | 9.57 | 0.002 | 0.060 |
| ENSMUSG00000050405 | Tmem151b | -0.17 | 5.35 | 9.57 | 0.002 | 0.060 |
| ENSMUSG00000020184 | Mdm2 | 0.11 | 5.67 | 9.54 | 0.002 | 0.061 |
| ENSMUSG00000019897 | Ccdc59 | -0.13 | 4.70 | 9.49 | 0.002 | 0.063 |
| ENSMUSG00000014773 | DII1 | 0.21 | 2.86 | 9.49 | 0.002 | 0.063 |
| ENSMUSG00000021962 | Dcp1a | 0.14 | 4.77 | 9.47 | 0.002 | 0.063 |
| ENSMUSG00000078611 | Gm5901 | 0.28 | 1.56 | 9.46 | 0.002 | 0.063 |
| ENSMUSG00000038457 | Tmem255b | -0.35 | 0.88 | 9.44 | 0.002 | 0.064 |
| ENSMUSG00000052076 | Gm9866 | -0.14 | 4.61 | 9.44 | 0.002 | 0.064 |
| ENSMUSG00000057387 | 4922502B01Rik | -0.34 | 0.86 | 9.44 | 0.002 | 0.064 |
| ENSMUSG00000049387 | Cox7b2 | -0.86 | -1.54 | 9.42 | 0.002 | 0.064 |
| ENSMUSG00000032988 | Slc16a8 | 0.47 | -0.08 | 9.40 | 0.002 | 0.065 |
| ENSMUSG00000029420 | Rimbp2 | -0.12 | 6.58 | 9.39 | 0.002 | 0.065 |
| ENSMUSG00000025064 | Col17a1 | 0.86 | -1.66 | 9.39 | 0.002 | 0.065 |
| ENSMUSG00000042846 | Lrrtm3 | -0.21 | 4.27 | 9.38 | 0.002 | 0.065 |
| ENSMUSG00000050234 | Gja4 | -0.28 | 1.74 | 9.38 | 0.002 | 0.065 |
| ENSMUSG00000041809 | Efhc1 | 0.35 | 0.79 | 9.36 | 0.002 | 0.066 |
| ENSMUSG00000028214 | Gem | 0.37 | 0.63 | 9.35 | 0.002 | 0.066 |
| ENSMUSG00000020836 | Coro6 | -0.25 | 5.15 | 9.34 | 0.002 | 0.066 |
| ENSMUSG00000028197 | Col24a1 | -0.48 | 0.07 | 9.34 | 0.002 | 0.066 |


| ENSMUSG00000042707 | Dnali1 | 0.37 | 1.58 | 9.34 | 0.002 | 0.066 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000046585 | Ccdc147 | -0.52 | -0.33 | 9.34 | 0.002 | 0.066 |
| ENSMUSG00000042605 | Atxn2 | -0.09 | 6.67 | 9.33 | 0.002 | 0.066 |
| ENSMUSG00000010080 | Epn3 | 0.59 | 1.42 | 9.32 | 0.002 | 0.066 |
| ENSMUSG00000023927 | Satb1 | -0.18 | 6.35 | 9.31 | 0.002 | 0.066 |
| ENSMUSG00000036578 | Fxyd7 | -0.19 | 5.51 | 9.31 | 0.002 | 0.066 |
| ENSMUSG00000038141 | Tmem181a | -0.15 | 3.82 | 9.31 | 0.002 | 0.066 |
| ENSMUSG00000075224 | Lrrc55 | -0.18 | 3.70 | 9.31 | 0.002 | 0.066 |
| ENSMUSG00000085929 | Gm13421 | -0.65 | -1.14 | 9.31 | 0.002 | 0.066 |
| ENSMUSG00000091270 | Gm17642 | 0.61 | -0.98 | 9.31 | 0.002 | 0.066 |
| ENSMUSG00000042115 | Klhdc8a | -0.26 | 3.25 | 9.29 | 0.002 | 0.067 |
| ENSMUSG00000041439 | Mfsd6 | -0.12 | 6.71 | 9.28 | 0.002 | 0.067 |
| ENSMUSG00000032422 | Snx14 | -0.13 | 5.41 | 9.27 | 0.002 | 0.067 |
| ENSMUSG00000050587 | Lrrc4c | -0.14 | 6.33 | 9.27 | 0.002 | 0.067 |
| ENSMUSG00000052229 | Gpr17 | -0.15 | 5.70 | 9.27 | 0.002 | 0.067 |
| ENSMUSG00000071533 | Pcnp | -0.12 | 5.93 | 9.26 | 0.002 | 0.067 |
| ENSMUSG00000025855 | Prkar1b | -0.09 | 9.28 | 9.24 | 0.002 | 0.068 |
| ENSMUSG00000028758 | Kif17 | -0.14 | 4.95 | 9.23 | 0.002 | 0.068 |
| ENSMUSG00000003418 | St8sia6 | 0.24 | 1.96 | 9.23 | 0.002 | 0.068 |
| ENSMUSG00000035356 | Nfkbiz | 0.18 | 3.27 | 9.22 | 0.002 | 0.069 |
| ENSMUSG00000038623 | Tm6sf1 | -0.20 | 2.95 | 9.22 | 0.002 | 0.069 |
| ENSMUSG00000035033 | Tbr1 | -0.17 | 5.53 | 9.21 | 0.002 | 0.069 |
| ENSMUSG00000053025 | Sv2b | -0.07 | 8.86 | 9.21 | 0.002 | 0.069 |
| ENSMUSG00000038806 | Sde2 | 0.14 | 4.62 | 9.20 | 0.002 | 0.069 |
| ENSMUSG00000045322 | TIr9 | -0.36 | 0.58 | 9.19 | 0.002 | 0.069 |
| ENSMUSG00000036362 | P2ry13 | -0.19 | 3.67 | 9.19 | 0.002 | 0.069 |
| ENSMUSG00000050945 | Zfp438 | 0.23 | 2.55 | 9.19 | 0.002 | 0.069 |
| ENSMUSG00000040016 | Ptger3 | -0.34 | 0.77 | 9.17 | 0.002 | 0.069 |
| ENSMUSG00000030235 | Slco1c1 | 0.13 | 5.88 | 9.16 | 0.002 | 0.070 |
| ENSMUSG00000048416 | MIf1 | 0.41 | 0.43 | 9.15 | 0.002 | 0.070 |
| ENSMUSG00000021177 | Tdp1 | -0.15 | 4.21 | 9.14 | 0.003 | 0.070 |
| ENSMUSG00000063415 | Cyp26b1 | -0.20 | 4.03 | 9.13 | 0.003 | 0.071 |
| ENSMUSG00000055980 | Irs1 | 0.19 | 3.00 | 9.13 | 0.003 | 0.071 |
| ENSMUSG00000028337 | Coro2a | -0.14 | 4.43 | 9.11 | 0.003 | 0.071 |
| ENSMUSG00000026383 | Epb4.115 | 0.19 | 2.90 | 9.10 | 0.003 | 0.071 |
| ENSMUSG00000070601 | Vmn2r84 | -0.43 | 0.14 | 9.10 | 0.003 | 0.071 |
| ENSMUSG00000089832 | Shkbp1 | 0.17 | 4.66 | 9.09 | 0.003 | 0.072 |
| ENSMUSG00000034227 | Foxj1 | 0.29 | 3.63 | 9.08 | 0.003 | 0.072 |
| ENSMUSG00000026725 | Tnn | -0.97 | -2.10 | 9.07 | 0.003 | 0.072 |
| ENSMUSG00000068696 | Gpr88 | -0.51 | 2.46 | 9.07 | 0.003 | 0.072 |
| ENSMUSG00000081382 | Rpl18-ps1 | 0.47 | 0.20 | 9.06 | 0.003 | 0.072 |
| ENSMUSG00000028522 | Mier1 | 0.13 | 4.98 | 9.04 | 0.003 | 0.073 |
| ENSMUSG00000078493 | A930039A15Rik | 0.36 | 0.50 | 9.04 | 0.003 | 0.073 |
| ENSMUSG00000046207 | Pik3r6 | -0.29 | 1.49 | 9.02 | 0.003 | 0.073 |
| ENSMUSG00000082160 | Gm11578 | -0.33 | 0.82 | 9.02 | 0.003 | 0.073 |
| ENSMUSG00000047657 | Crry-ps | -0.61 | -0.96 | 9.02 | 0.003 | 0.074 |
| ENSMUSG00000087018 | 2900072N19Rik | -0.29 | 1.18 | 9.01 | 0.003 | 0.074 |
| ENSMUSG00000020363 | Gfpt2 | 0.23 | 2.68 | 9.00 | 0.003 | 0.074 |
| ENSMUSG00000050671 | Ism2 | 0.73 | -1.30 | 8.99 | 0.003 | 0.074 |
| ENSMUSG00000032715 | Trib3 | -0.54 | -0.57 | 8.98 | 0.003 | 0.075 |
| ENSMUSG00000043015 | Tmem194b | -0.22 | 2.94 | 8.97 | 0.003 | 0.075 |


| ENSMUSG00000086782 | E130102H24Rik | -0.25 | 1.97 | 8.97 | 0.003 | 0.075 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000026167 | Wnt10a | -0.26 | 2.01 | 8.96 | 0.003 | 0.075 |
| ENSMUSG00000035248 | Zcchc6 | -0.11 | 5.35 | 8.96 | 0.003 | 0.075 |
| ENSMUSG00000039725 | 2810408M09Rik | 0.14 | 4.14 | 8.96 | 0.003 | 0.075 |
| ENSMUSG00000066189 | Cacng3 | -0.13 | 5.92 | 8.96 | 0.003 | 0.075 |
| ENSMUSG00000092210 | A930009A15Rik | -0.62 | -0.86 | 8.96 | 0.003 | 0.075 |
| ENSMUSG00000028943 | Espn | 0.22 | 3.02 | 8.94 | 0.003 | 0.075 |
| ENSMUSG00000028030 | Tbck | -0.16 | 3.57 | 8.92 | 0.003 | 0.076 |
| ENSMUSG00000054945 | Gm9958 | -0.41 | 0.20 | 8.91 | 0.003 | 0.076 |
| ENSMUSG00000045034 | Ankrd34b | -0.17 | 3.92 | 8.91 | 0.003 | 0.076 |
| ENSMUSG00000021913 | Ogdhl | -0.11 | 6.64 | 8.90 | 0.003 | 0.076 |
| ENSMUSG00000031627 | Iff2 | -0.13 | 4.67 | 8.90 | 0.003 | 0.076 |
| ENSMUSG00000078923 | Ube2v1 | 0.70 | -1.34 | 8.90 | 0.003 | 0.076 |
| ENSMUSG00000042045 | SIn | 0.58 | -0.34 | 8.89 | 0.003 | 0.076 |
| ENSMUSG00000071192 | Wfikkn1 | 0.58 | -0.64 | 8.89 | 0.003 | 0.076 |
| ENSMUSG00000062691 | Cebpzos | 0.27 | 1.76 | 8.87 | 0.003 | 0.077 |
| ENSMUSG00000038195 | Rilp | 0.36 | 0.76 | 8.87 | 0.003 | 0.077 |
| ENSMUSG00000046441 | Cmtr2 | 0.19 | 2.92 | 8.84 | 0.003 | 0.078 |
| ENSMUSG00000070565 | Rasal2 | 0.17 | 6.24 | 8.84 | 0.003 | 0.078 |
| ENSMUSG00000032334 | Lox11 | 0.22 | 2.52 | 8.83 | 0.003 | 0.078 |
| ENSMUSG00000023882 | Zfp54 | 0.42 | 0.12 | 8.83 | 0.003 | 0.078 |
| ENSMUSG00000081885 | Gm13231 | 0.67 | -1.46 | 8.83 | 0.003 | 0.078 |
| ENSMUSG00000001494 | Sost | 0.84 | -1.65 | 8.81 | 0.003 | 0.079 |
| ENSMUSG00000033715 | Akr1c14 | -0.37 | 0.43 | 8.80 | 0.003 | 0.079 |
| ENSMUSG00000021966 | Prss52 | 0.66 | -1.17 | 8.79 | 0.003 | 0.080 |
| ENSMUSG00000050295 | Foxc1 | -0.24 | 2.42 | 8.78 | 0.003 | 0.080 |
| ENSMUSG00000068566 | Myadm | -0.11 | 6.25 | 8.79 | 0.003 | 0.080 |
| ENSMUSG00000022816 | Fst11 | -0.15 | 5.29 | 8.78 | 0.003 | 0.080 |
| ENSMUSG00000044795 | Cyb5d1 | -0.17 | 3.69 | 8.77 | 0.003 | 0.080 |
| ENSMUSG00000047227 | Gm527 | 0.27 | 1.67 | 8.77 | 0.003 | 0.080 |
| ENSMUSG00000028840 | Zfp593 | 0.54 | -0.34 | 8.76 | 0.003 | 0.080 |
| ENSMUSG00000038893 | Fam117a | 0.22 | 3.15 | 8.76 | 0.003 | 0.080 |
| ENSMUSG00000037096 | Gm9762 | -0.14 | 4.06 | 8.76 | 0.003 | 0.080 |
| ENSMUSG00000046182 | Gsg11 | -0.23 | 5.36 | 8.75 | 0.003 | 0.080 |
| ENSMUSG00000030987 | Stim1 | -0.09 | 6.32 | 8.75 | 0.003 | 0.080 |
| ENSMUSG00000034751 | Mast4 | 0.16 | 5.84 | 8.74 | 0.003 | 0.080 |
| ENSMUSG00000019890 | Nts | -0.33 | 2.43 | 8.74 | 0.003 | 0.081 |
| ENSMUSG00000019861 | Gopc | -0.10 | 5.73 | 8.73 | 0.003 | 0.081 |
| ENSMUSG00000030111 | A2m | 0.31 | 2.22 | 8.72 | 0.003 | 0.081 |
| ENSMUSG00000030674 | Qprt | 0.57 | -0.82 | 8.71 | 0.003 | 0.081 |
| ENSMUSG00000047643 | Gm5454 | -0.22 | 3.34 | 8.70 | 0.003 | 0.082 |
| ENSMUSG00000008153 | Clstn3 | -0.08 | 8.66 | 8.69 | 0.003 | 0.082 |
| ENSMUSG00000027547 | Sall4 | 0.54 | -0.71 | 8.69 | 0.003 | 0.082 |
| ENSMUSG00000020420 | Zfp607 | 0.46 | -0.25 | 8.66 | 0.003 | 0.083 |
| ENSMUSG00000021260 | Hhipl1 | -0.29 | 1.26 | 8.66 | 0.003 | 0.083 |
| ENSMUSG00000025407 | Gli1 | -0.38 | 1.37 | 8.65 | 0.003 | 0.083 |
| ENSMUSG00000029636 | Wasf3 | -0.14 | 6.60 | 8.65 | 0.003 | 0.083 |
| ENSMUSG00000035852 | Misp | 0.59 | -0.61 | 8.65 | 0.003 | 0.083 |
| ENSMUSG00000022211 | Lrrc16b | -0.11 | 6.38 | 8.65 | 0.003 | 0.083 |
| ENSMUSG00000028804 | Csmd2 | 0.14 | 5.94 | 8.64 | 0.003 | 0.084 |
| ENSMUSG00000042182 | Bend6 | -0.15 | 6.48 | 8.62 | 0.003 | 0.084 |


| ENSMUSG00000002885 | Cd97 | -0.17 | 3.33 | 8.61 | 0.003 | 0.084 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000020014 | 4930485B16Rik | 0.26 | 2.67 | 8.62 | 0.003 | 0.084 |
| ENSMUSG00000026185 | Igfbp5 | -0.15 | 7.79 | 8.61 | 0.003 | 0.084 |
| ENSMUSG00000027177 | Hipk3 | 0.10 | 5.94 | 8.61 | 0.003 | 0.084 |
| ENSMUSG00000031750 | II34 | -0.14 | 5.57 | 8.61 | 0.003 | 0.084 |
| ENSMUSG00000045775 | Slc16a5 | -0.44 | 0.13 | 8.61 | 0.003 | 0.084 |
| ENSMUSG00000019359 | Gdpd2 | -0.20 | 2.67 | 8.58 | 0.003 | 0.085 |
| ENSMUSG00000036560 | Lgi4 | -0.15 | 3.78 | 8.58 | 0.003 | 0.085 |
| ENSMUSG00000059839 | Zfp874b | 0.16 | 4.03 | 8.58 | 0.003 | 0.085 |
| ENSMUSG00000072494 | Ppp1r3e | -0.18 | 3.20 | 8.59 | 0.003 | 0.085 |
| ENSMUSG00000093149 | Gm23374 | -0.60 | -0.97 | 8.58 | 0.003 | 0.085 |
| ENSMUSG00000046192 | Iqub | 0.32 | 1.11 | 8.56 | 0.003 | 0.085 |
| ENSMUSG00000030666 | Calcb | -0.85 | -1.76 | 8.56 | 0.003 | 0.085 |
| ENSMUSG00000059040 | Eno1b | -0.18 | 3.94 | 8.56 | 0.003 | 0.086 |
| ENSMUSG00000040495 | Chrm4 | 0.20 | 4.65 | 8.55 | 0.003 | 0.086 |
| ENSMUSG00000022607 | Ptk2 | -0.11 | 6.93 | 8.54 | 0.003 | 0.086 |
| ENSMUSG00000034917 | Tjp3 | 0.35 | 0.71 | 8.54 | 0.003 | 0.086 |
| ENSMUSG00000038605 | Samd10 | -0.16 | 4.13 | 8.52 | 0.004 | 0.086 |
| ENSMUSG00000048004 | Tmem196 | -0.30 | 1.17 | 8.52 | 0.004 | 0.086 |
| ENSMUSG00000074406 | Zfp628 | -0.17 | 3.85 | 8.52 | 0.004 | 0.086 |
| ENSMUSG00000091641 | Gm17593 | -0.49 | -0.44 | 8.52 | 0.004 | 0.086 |
| ENSMUSG00000039737 | Prkrip1 | 0.15 | 4.64 | 8.47 | 0.004 | 0.088 |
| ENSMUSG00000026377 | Nifk | 0.12 | 4.98 | 8.47 | 0.004 | 0.089 |
| ENSMUSG00000004105 | Angpt12 | 0.28 | 1.78 | 8.46 | 0.004 | 0.089 |
| ENSMUSG00000005148 | Klf5 | -0.26 | 1.72 | 8.46 | 0.004 | 0.089 |
| ENSMUSG00000026141 | Col19a1 | 0.19 | 2.95 | 8.44 | 0.004 | 0.089 |
| ENSMUSG00000030098 | Grip2 | -0.15 | 4.07 | 8.45 | 0.004 | 0.089 |
| ENSMUSG00000033871 | Ppargc1b | 0.23 | 2.05 | 8.44 | 0.004 | 0.089 |
| ENSMUSG00000038072 | Galnt11 | -0.13 | 5.23 | 8.45 | 0.004 | 0.089 |
| ENSMUSG00000041911 | Dlx1 | 0.13 | 5.16 | 8.45 | 0.004 | 0.089 |
| ENSMUSG00000050157 | Gm867 | 0.38 | 0.43 | 8.46 | 0.004 | 0.089 |
| ENSMUSG00000052861 | Dnah6 | 0.29 | 2.06 | 8.45 | 0.004 | 0.089 |
| ENSMUSG00000073176 | Zfp449 | 0.22 | 2.37 | 8.45 | 0.004 | 0.089 |
| ENSMUSG00000018599 | Mief2 | -0.13 | 4.43 | 8.44 | 0.004 | 0.089 |
| ENSMUSG00000030446 | Zfp273 | -0.19 | 2.69 | 8.44 | 0.004 | 0.089 |
| ENSMUSG00000041515 | Irf8 | -0.22 | 2.45 | 8.43 | 0.004 | 0.089 |
| ENSMUSG00000074671 | Tspyl3 | -0.13 | 4.93 | 8.43 | 0.004 | 0.089 |
| ENSMUSG00000042364 | Snx18 | 0.12 | 4.71 | 8.41 | 0.004 | 0.090 |
| ENSMUSG00000037253 | Mex3c | 0.13 | 5.26 | 8.40 | 0.004 | 0.090 |
| ENSMUSG00000031971 | Ccsap | -0.14 | 5.68 | 8.39 | 0.004 | 0.091 |
| ENSMUSG00000026437 | Cdk18 | -0.12 | 5.05 | 8.38 | 0.004 | 0.091 |
| ENSMUSG00000091890 | A830073021Rik | -0.36 | 0.38 | 8.37 | 0.004 | 0.091 |
| ENSMUSG00000023852 | Chd1 | 0.12 | 4.48 | 8.37 | 0.004 | 0.091 |
| ENSMUSG00000054044 | Gm9933 | -0.72 | -1.57 | 8.37 | 0.004 | 0.091 |
| ENSMUSG00000033055 | Ankrd54 | 0.14 | 4.12 | 8.36 | 0.004 | 0.091 |
| ENSMUSG00000024176 | Sox8 | 0.17 | 5.54 | 8.36 | 0.004 | 0.091 |
| ENSMUSG00000022099 | Dmtn | -0.07 | 8.53 | 8.35 | 0.004 | 0.091 |
| ENSMUSG00000046010 | Zfp830 | 0.16 | 3.47 | 8.35 | 0.004 | 0.091 |
| ENSMUSG00000044252 | Osbpl1a | -0.12 | 7.17 | 8.33 | 0.004 | 0.092 |
| ENSMUSG00000065968 | Ifitm7 | 0.76 | -1.72 | 8.33 | 0.004 | 0.092 |
| ENSMUSG00000030201 | Lrp6 | 0.12 | 5.57 | 8.33 | 0.004 | 0.092 |


| ENSMUSG00000024501 | Dpysl3 | -0.10 | 6.08 | 8.31 | 0.004 | 0.093 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000036904 | Fzd8 | -0.20 | 2.49 | 8.30 | 0.004 | 0.093 |
| ENSMUSG00000045294 | Insig1 | 0.13 | 6.03 | 8.29 | 0.004 | 0.094 |
| ENSMUSG00000021891 | Mettl6 | 0.18 | 4.04 | 8.28 | 0.004 | 0.094 |
| ENSMUSG00000028744 | Pqlc2 | 0.16 | 3.72 | 8.28 | 0.004 | 0.094 |
| ENSMUSG00000024043 | Arhgap28 | 0.49 | -0.50 | 8.27 | 0.004 | 0.094 |
| ENSMUSG00000033454 | Zbtb1 | 0.14 | 4.27 | 8.26 | 0.004 | 0.094 |
| ENSMUSG00000044835 | Ankrd45 | -0.14 | 4.75 | 8.26 | 0.004 | 0.094 |
| ENSMUSG00000084098 | Gm13422 | 0.22 | 1.98 | 8.27 | 0.004 | 0.094 |
| ENSMUSG00000007440 | Pcdha10 | -0.08 | 7.26 | 8.24 | 0.004 | 0.095 |
| ENSMUSG00000021573 | Tppp | -0.07 | 9.04 | 8.24 | 0.004 | 0.095 |
| ENSMUSG00000027399 | II1a | -0.59 | -0.97 | 8.25 | 0.004 | 0.095 |
| ENSMUSG00000028149 | Rap1gds1 | -0.09 | 8.27 | 8.24 | 0.004 | 0.095 |
| ENSMUSG00000055831 | Gm9982 | 0.51 | -0.59 | 8.25 | 0.004 | 0.095 |
| ENSMUSG00000060510 | Zfp266 | 0.13 | 5.03 | 8.26 | 0.004 | 0.095 |
| ENSMUSG00000087081 | 6430590A07Rik | -0.57 | -0.83 | 8.25 | 0.004 | 0.095 |
| ENSMUSG00000090194 | Gm16161 | 0.66 | -1.24 | 8.25 | 0.004 | 0.095 |
| ENSMUSG00000091166 | Tstd1 | 0.40 | 0.29 | 8.25 | 0.004 | 0.095 |
| ENSMUSG00000060240 | Cend1 | -0.09 | 8.12 | 8.24 | 0.004 | 0.095 |
| ENSMUSG00000030699 | Tbx6 | 0.35 | 0.80 | 8.23 | 0.004 | 0.095 |
| ENSMUSG00000017740 | SIc12a5 | -0.09 | 9.15 | 8.21 | 0.004 | 0.096 |
| ENSMUSG00000020134 | Peli1 | 0.12 | 5.31 | 8.20 | 0.004 | 0.096 |
| ENSMUSG00000020377 | Ltc4s | 0.55 | -0.49 | 8.20 | 0.004 | 0.096 |
| ENSMUSG00000030898 | Cckbr | -0.24 | 3.45 | 8.20 | 0.004 | 0.096 |
| ENSMUSG00000036882 | Arhgap33 | -0.14 | 7.28 | 8.20 | 0.004 | 0.096 |
| ENSMUSG00000042106 | Fam212a | 0.22 | 2.45 | 8.19 | 0.004 | 0.096 |
| ENSMUSG00000026301 | Iqca | 0.35 | 1.02 | 8.18 | 0.004 | 0.096 |
| ENSMUSG00000031351 | Zfp185 | 0.30 | 1.79 | 8.19 | 0.004 | 0.096 |
| ENSMUSG00000031431 | Tsc22d3 | 0.23 | 5.84 | 8.18 | 0.004 | 0.096 |
| ENSMUSG00000074657 | Kif5a | -0.08 | 11.29 | 8.17 | 0.004 | 0.097 |
| ENSMUSG00000025196 | Cpn1 | 0.71 | -1.44 | 8.16 | 0.004 | 0.097 |
| ENSMUSG00000021457 | Syk | -0.21 | 2.29 | 8.13 | 0.004 | 0.098 |
| ENSMUSG00000021763 | BC067074 | 0.46 | 1.15 | 8.15 | 0.004 | 0.098 |
| ENSMUSG00000029343 | Crybb1 | -0.24 | 2.12 | 8.14 | 0.004 | 0.098 |
| ENSMUSG00000037709 | Fam13a | 0.16 | 4.76 | 8.13 | 0.004 | 0.098 |
| ENSMUSG00000042508 | Dmtf1 | -0.11 | 5.81 | 8.13 | 0.004 | 0.098 |
| ENSMUSG00000052142 | Rasal3 | -0.24 | 1.87 | 8.15 | 0.004 | 0.098 |
| ENSMUSG00000053004 | Hrh1 | -0.34 | 0.74 | 8.14 | 0.004 | 0.098 |
| ENSMUSG00000054604 | Cggbp1 | 0.12 | 5.89 | 8.14 | 0.004 | 0.098 |
| ENSMUSG00000058013 | Sept11 | -0.10 | 6.59 | 8.14 | 0.004 | 0.098 |
| ENSMUSG00000064293 | Cntn4 | -0.18 | 3.99 | 8.13 | 0.004 | 0.098 |
| ENSMUSG00000005718 | Tfap4 | -0.24 | 2.04 | 8.12 | 0.004 | 0.098 |
| ENSMUSG00000001034 | Mapk7 | 0.15 | 5.23 | 8.12 | 0.004 | 0.098 |
| ENSMUSG00000001441 | Npepps | -0.11 | 7.41 | 8.11 | 0.004 | 0.098 |
| ENSMUSG00000024855 | Pacs1 | -0.10 | 6.64 | 8.11 | 0.004 | 0.098 |
| ENSMUSG00000028955 | Vamp3 | 0.13 | 4.91 | 8.10 | 0.004 | 0.098 |
| ENSMUSG00000029189 | Sel113 | -0.16 | 5.84 | 8.11 | 0.004 | 0.098 |
| ENSMUSG00000031391 | L1cam | -0.13 | 7.20 | 8.10 | 0.004 | 0.098 |
| ENSMUSG00000074227 | Spint2 | 0.19 | 4.38 | 8.10 | 0.004 | 0.098 |
| ENSMUSG00000082585 | Gm15387 | -0.34 | 0.88 | 8.10 | 0.004 | 0.098 |
| ENSMUSG00000024897 | Apba1 | -0.11 | 7.62 | 8.09 | 0.004 | 0.098 |


| ENSMUSG00000030019 | Fbxl14 | 0.15 | 4.93 | 8.09 | 0.004 | 0.098 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000031520 | Vegfc | -0.26 | 1.99 | 8.09 | 0.004 | 0.098 |
| ENSMUSG00000032637 | Atxn2l | -0.08 | 7.82 | 8.09 | 0.004 | 0.098 |
| ENSMUSG00000035621 | Midn | 0.14 | 5.76 | 8.09 | 0.004 | 0.098 |
| ENSMUSG00000029861 | Fam131b | -0.10 | 6.87 | 8.07 | 0.004 | 0.099 |
| ENSMUSG00000032908 | Sgpp2 | -0.23 | 3.25 | 8.07 | 0.004 | 0.099 |
| ENSMUSG00000044352 | Sowaha | 0.09 | 7.28 | 8.07 | 0.005 | 0.099 |
| ENSMUSG00000031077 | Fadd | -0.29 | 1.34 | 8.06 | 0.005 | 0.099 |
| ENSMUSG00000027811 | 4930579G24Rik | 0.34 | 0.65 | 8.05 | 0.005 | 0.100 |
| ENSMUSG00000049892 | Rasd1 | 0.20 | 5.33 | 8.05 | 0.005 | 0.100 |

Table 2.3

| GenelD | Gene Name | BinID | logFC | $t$ | p-value | FDR |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000007617 | Homer1 | 18 | 2.0 | 13.3 | 5.0E-30 | 1.3E-24 |
| ENSMUSG00000029657 | Hsph1 | 12 | 1.0 | 9.2 | 1.7E-17 | 2.3E-12 |
| ENSMUSG00000039801 | 2410089E03Rik | 53 | -1.1 | -8.5 | 2.2E-16 | 2.0E-11 |
| ENSMUSG00000007617 | Homer1 | 17 | 1.8 | 8.0 | 9.3E-14 | 6.2E-09 |
| ENSMUSG00000031167 | Rbm3 | 2 | 0.9 | 7.8 | 1.8E-13 | 9.8E-09 |
| ENSMUSG00000034083 | Ccdc174 | 9 | 1.0 | 7.2 | 1.7E-11 | 7.7E-07 |
| ENSMUSG00000025372 | Baiap2 | 27 | 0.5 | 7.0 | 2.9E-11 | 1.1E-06 |
| ENSMUSG00000008153 | Clstn3 | 4 | 1.2 | 6.6 | 2.4E-10 | 7.7E-06 |
| ENSMUSG00000057421 | Las11 | 15 | -1.0 | -6.6 | 2.6E-10 | 7.7E-06 |
| ENSMUSG00000020287 | Mpg | 10 | -0.6 | -6.9 | $4.9 \mathrm{E}-10$ | 1.3E-05 |
| ENSMUSG00000063077 | Kif1b | 41 | 0.5 | 6.3 | 7.3E-10 | 1.8E-05 |
| ENSMUSG00000043872 | Zmym1 | 1 | 1.0 | 6.5 | 1.1E-09 | 2.4E-05 |
| ENSMUSG00000035206 | Sppl2b | 16 | 0.4 | 6.1 | 7.6E-09 | 1.6E-04 |
| ENSMUSG00000024576 | Csnk1a1 | 27 | 0.6 | 5.9 | 1.4E-08 | $2.5 \mathrm{E}-04$ |
| ENSMUSG00000025372 | Baiap2 | 26 | 0.6 | 5.9 | 1.4E-08 | 2.5E-04 |
| ENSMUSG00000041879 | Ipo9 | 36 | -0.8 | -5.8 | 1.6E-08 | 2.7E-04 |
| ENSMUSG00000024576 | Csnk1a1 | 28 | 0.5 | 5.7 | 3.1E-08 | 4.7E-04 |
| ENSMUSG000000063160 | Numbl/Adck4 | 37 | 0.4 | 5.7 | 3.1E-08 | 4.7E-04 |
| ENSMUSG00000028782 | Bai2 | 46 | 0.3 | 5.6 | 3.9E-08 | 5.4E-04 |
| ENSMUSG00000023952 | Gtpbp2 | 44 | 0.5 | 5.6 | 5.2E-08 | 7.0E-04 |
| ENSMUSG00000028826 | Tmem57 | 2 | -0.5 | -5.6 | 7.7E-08 | 9.8E-04 |
| ENSMUSG00000022710 | Usp7 | 20 | 0.7 | 5.4 | 8.4E-08 | 0.001 |
| ENSMUSG00000034171 | Faah | 14 | 0.4 | 5.5 | 1.1E-07 | 0.001 |
| ENSMUSG00000024826 | Dpf2 | 11 | 0.5 | 5.5 | 1.3E-07 | 0.001 |
| ENSMUSG00000038383 | Pigu | 4 | 0.4 | 5.4 | 2.6E-07 | 0.003 |
| ENSMUSG00000060216 | Arrb2 | 11 | 0.8 | 5.3 | 2.7E-07 | 0.003 |
| ENSMUSG00000023952 | Gtpbp2 | 35 | 0.9 | 5.2 | 3.1E-07 | 0.003 |
| ENSMUSG00000027569 | Mrgbp | 10 | 0.5 | 5.4 | 3.3E-07 | 0.003 |
| ENSMUSG00000036052 | Dnajb5 | 11 | 0.4 | 5.4 | 3.4E-07 | 0.003 |
| ENSMUSG00000014873 | Surf2 | 9 | 0.4 | 5.3 | 3.5E-07 | 0.003 |
| ENSMUSG00000024777 | Ppp2r5b | 6 | 0.5 | 5.3 | 4.3E-07 | 0.004 |
| ENSMUSG00000071984 | Fndc1 | 1 | 0.4 | 5.2 | 4.2E-07 | 0.004 |
| ENSMUSG00000035202 | Lars2 | 21 | -0.8 | -5.2 | 5.3E-07 | 0.004 |
| ENSMUSG00000039219 | Arid4b | 44 | -0.6 | -5.1 | 5.4E-07 | 0.004 |
| ENSMUSG00000031878 | Nae1 | 15 | 1.0 | 5.0 | 8.3E-07 | 0.006 |
| ENSMUSG00000044308 | Ubr3 | 54 | -0.7 | -5.0 | 8.1E-07 | 0.006 |
| ENSMUSG00000006676 | Usp19 | 27 | 0.3 | 5.1 | 9.5E-07 | 0.007 |
| ENSMUSG000000092679 | Mir5129/Zeb2 | 1 | -0.2 | -5.0 | 9.4E-07 | 0.007 |
| ENSMUSG00000075003 | Jmjd1c | 41 | 0.7 | 5.0 | 9.8E-07 | 0.007 |
| ENSMUSG00000038664 | Herc1 | 95 | -0.6 | -4.9 | 1.1E-06 | 0.007 |
| ENSMUSG00000038538 | Ubn2 | 38 | 0.3 | 5.0 | 1.1E-06 | 0.007 |
| ENSMUSG00000040479 | Dgkz | 11 | 0.6 | 4.9 | 1.2E-06 | 0.008 |
| ENSMUSG00000063160 | Numbl | 35 | 0.5 | 4.9 | 1.4E-06 | 0.008 |
| ENSMUSG00000084896 | Gm11632/Fbxl20 | 14 | -0.7 | -4.9 | 1.4E-06 | 0.009 |


| ENSMUSG00000020883 |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000050357 | Rltpr | 40 | 0.4 | 4.9 | 1.8E-06 | 0.011 |
| ENSMUSG00000075876 |  |  |  |  |  |  |
| ENSMUSG00000064791 |  |  |  |  |  |  |
| ENSMUSG00000075924 |  |  |  |  |  |  |
| ENSMUSG00000015656 | Hspa8/Snord14e | 38 | 1.4 | 4.9 | 1.8E-06 | 0.011 |
| ENSMUSG00000020258 | Glyctk | 3 | -1.8 | -5.0 | 2.1E-06 | 0.012 |
| ENSMUSG00000034739 |  |  |  |  |  |  |
| ENSMUSG00000079592 | Mfrp/C1qtnf5 | 30 | -1.2 | -4.8 | 2.1E-06 | 0.012 |
| ENSMUSG00000034656 | Cacna1a | 68 | 1.0 | 4.7 | 2.8E-06 | 0.015 |
| ENSMUSG00000074247 | Dda1 | 13 | 0.8 | 4.9 | 3.4E-06 | 0.018 |
| ENSMUSG00000027429 | Sec23b | 30 | 0.8 | 4.7 | 3.7E-06 | 0.019 |
| ENSMUSG00000042605 | Atxn2 | 51 | 0.6 | 4.7 | 3.8E-06 | 0.019 |
| ENSMUSG00000007617 | Homer1 | 16 | 1.7 | 4.7 | 3.9E-06 | 0.020 |
| ENSMUSG00000075876 |  |  |  |  |  |  |
| ENSMUSG00000064791 |  |  |  |  |  |  |
| ENSMUSG00000075924 |  |  |  |  |  |  |
| ENSMUSG00000015656 | Hspa8/Snord14e | 37 | 1.3 | 4.7 | 4.0E-06 | 0.020 |
| ENSMUSG00000078789 |  |  |  |  |  |  |
| ENSMUSG00000038268 | Dph1/Ovca2 | 1 | 0.3 | 4.7 | 4.2E-06 | 0.020 |
| ENSMUSG00000023952 | Gtpbp2 | 31 | 0.4 | 4.6 | 4.8E-06 | 0.023 |
| ENSMUSG00000031167 | Rbm3 | 22 | -0.6 | -4.7 | 4.9E-06 | 0.023 |
| ENSMUSG00000039515 | Ppp2r4 | 26 | 0.6 | 4.7 | 5.2E-06 | 0.024 |
| ENSMUSG00000027185 | Nat10 | 27 | 0.9 | 4.6 | 5.3E-06 | 0.024 |
| ENSMUSG00000035640 | Dos | 14 | 1.7 | 4.6 | 5.7E-06 | 0.025 |
| ENSMUSG00000071646 | Mta2 | 3 | 1.0 | 4.7 | 5.8E-06 | 0.025 |
| ENSMUSG00000038664 | Herc1 | 93 | -0.5 | -4.6 | 6.2E-06 | 0.027 |
| ENSMUSG00000019790 | Stxbp5 | 17 | -0.7 | -4.6 | 6.4E-06 | 0.027 |
| ENSMUSG00000027893 | Ahcyl1 | 7 | 0.8 | 4.6 | 7.2E-06 | 0.030 |
| ENSMUSG00000040479 | Dgkz | 10 | 0.6 | 4.5 | 7.2E-06 | 0.030 |
| ENSMUSG00000026918 | Brd3 | 13 | -1.2 | -4.6 | 7.5E-06 | 0.030 |
| ENSMUSG00000009073 | Nf2 | 14 | 0.8 | 4.5 | 1.1E-05 | 0.041 |
| ENSMUSG00000079737 | 3110001I22Rik/Bfa |  |  |  |  |  |
| ENSMUSG00000022684 | - r | 15 | -0.8 | -4.6 | 1.0E-05 | 0.041 |
| ENSMUSG00000046709 | Mapk10 | 14 | -0.4 | -4.5 | 1.1E-05 | 0.042 |
| ENSMUSG00000052423 | B4galt3 | 14 | -0.8 | -4.5 | 1.1E-05 | 0.042 |
| ENSMUSG00000039219 | Arid4b | 42 | -0.7 | -4.5 | 1.1E-05 | 0.042 |
| ENSMUSG00000074247 | Dda1 | 12 | 0.7 | 4.6 | 1.1E-05 | 0.042 |
| ENSMUSG00000035027 | Map2k2 | 9 | -0.9 | -4.5 | 1.2E-05 | 0.045 |
| ENSMUSG00000033365 | Ipo13 | 8 | 0.4 | 4.4 | 1.3E-05 | 0.047 |
| ENSMUSG00000006024 | Napa | 16 | 0.3 | 4.5 | 1.4E-05 | 0.048 |
| ENSMUSG00000019189 | Rnf145 | 12 | -0.6 | -4.5 | 1.4E-05 | 0.048 |
| ENSMUSG00000005378 | Wbscr22 | 30 | 0.9 | 4.4 | 1.4E-05 | 0.048 |
| ENSMUSG00000020978 | Klhdc2 | 2 | 0.8 | 4.5 | 1.4E-05 | 0.048 |
| ENSMUSG00000023952 | Gtpbp2 | 36 | 0.6 | 4.4 | 1.4E-05 | 0.048 |
| ENSMUSG00000040268 | Plekha1 | 15 | -1.1 | -4.4 | 1.4E-05 | 0.048 |
| ENSMUSG00000042726 | Trafd1 | 10 | -0.4 | -4.4 | 1.5E-05 | 0.048 |
| ENSMUSG00000038406 | Scaf1 | 10 | 0.7 | 4.4 | 1.5E-05 | 0.050 |
| ENSMUSG00000063160 |  |  |  |  |  |  |
| ENSMUSG00000003762 | Numbl/Adck4 | 34 | 0.7 | 4.4 | 1.6E-05 | 0.050 |
| ENSMUSG00000022099 | Dmtn | 6 | 0.5 | 4.4 | 1.6E-05 | 0.052 |
| ENSMUSG00000008153 | Clstn3 | 5 | 1.3 | 4.4 | 1.7E-05 | 0.053 |


| ENSMUSG00000050875 | A730017C20Rik | 12 | -0.7 | -4.5 | 1.7E-05 | 0.053 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000001018 | Snapin | 7 | -0.5 | -4.5 | 1.8E-05 | 0.054 |
| ENSMUSG00000007850 | Hnrnph1 | 46 | 0.9 | 4.3 | 1.9E-05 | 0.056 |
| ENSMUSG00000021327 | Zkscan3 | 9 | 0.3 | 4.4 | 1.9E-05 | 0.056 |
| $\begin{aligned} & \text { ENSMUSG00000084708 } \\ & \text { ENSMUSG00000065862 } \\ & \text { ENSMUSG00000059796 } \end{aligned}$ | $\begin{gathered} \text { Gm22988/Gm2402 } \\ \text { 9/Eif4a1 } \end{gathered}$ | 38 | 0.5 | 4.3 | 1.8E-05 | 0.056 |
| ENSMUSG00000029050 | Ski | 20 | 0.5 | 4.4 | 2.0E-05 | 0.059 |
| ENSMUSG00000024576 | Csnk1a1 | 25 | 0.4 | 4.3 | 2.1E-05 | 0.062 |
| ENSMUSG00000003269 | Cyth2 | 23 | 0.4 | 4.3 | 2.4E-05 | 0.065 |
| ENSMUSG00000020612 | Prkar1a | 11 | 0.4 | 4.4 | 2.3E-05 | 0.065 |
| ENSMUSG00000020882 | Cacnb1 | 3 | 0.6 | 4.3 | 2.3E-05 | 0.065 |
| ENSMUSG00000024807 | Syvn1 | 34 | 0.6 | 4.3 | 2.4E-05 | 0.065 |
| ENSMUSG00000025575 | Cant1 | 5 | 0.8 | 4.4 | 2.4E-05 | 0.065 |
| ENSMUSG00000040447 | Spns2 | 7 | 0.5 | 4.3 | 2.4E-05 | 0.065 |
| ENSMUSG00000052387 | Trpm3 | 15 | 2.6 | 4.3 | 2.4E-05 | 0.065 |
| ENSMUSG00000031367 | Ap1s2 | 9 | 0.5 | 4.4 | 2.5E-05 | 0.067 |
| ENSMUSG00000040029 | Ipo8 | 20 | 1.5 | 4.3 | 2.5E-05 | 0.067 |
| ENSMUSG00000057522 | Spop | 19 | -1.2 | -4.3 | 2.6E-05 | 0.068 |
| ENSMUSG00000038696 | Mapkap1 | 7 | 0.5 | 4.3 | 2.7E-05 | 0.071 |
| ENSMUSG00000002280 | Narfl | 18 | -0.6 | -4.3 | 2.8E-05 | 0.072 |
| ENSMUSG00000057421 | Las11 | 14 | -0.8 | -4.3 | 3.0E-05 | 0.077 |
| ENSMUSG00000038611 | Phrf1 | 21 | 1.8 | 4.2 | 3.2E-05 | 0.079 |
| ENSMUSG00000034739 <br> ENSMUSG00000079592 | Mfrp/C1qtnf5 | 10 | 4.5 | 4.2 | 3.3E-05 | 0.083 |
| ENSMUSG00000031167 | Rbm3 | 3 | 0.8 | 4.2 | 3.5E-05 | 0.085 |
| ENSMUSG00000005417 | Mprip | 15 | 0.5 | 4.2 | 3.5E-05 | 0.086 |
| ENSMUSG00000020894 | Vamp2 | 13 | 0.5 | 4.3 | 3.6E-05 | 0.088 |
| ENSMUSG00000060261 | Gtf2i | 47 | -1.6 | -4.2 | 3.7E-05 | 0.088 |
| ENSMUSG00000084708 ENSMUSG00000065862 ENSMUSG00000059796 | Gm22988/Gm2402 9/Eif4a1 | 37 | 0.5 | 4.2 | 3.7E-05 | 0.088 |
| ENSMUSG00000025571 | Tnrc6c | 16 | -0.5 | -4.2 | 3.8E-05 | 0.090 |
| ENSMUSG00000052593 | Adam17 | 7 | 2.4 | 4.2 | 3.9E-05 | 0.092 |
| ENSMUSG00000020918 | Kat2a | 8 | 0.5 | 4.2 | 4.0E-05 | 0.094 |
| ENSMUSG00000025860 | Xiap | 24 | 0.2 | 4.2 | 4.1E-05 | 0.095 |
| ENSMUSG00000034675 | Dbn1 | 11 | 0.3 | 4.2 | 4.4E-05 | 0.099 |
| ENSMUSG00000071646 | Mta2 | 2 | 1.5 | 4.2 | 4.4E-05 | 0.099 |

# CHAPTER 3: Object-Location Training Elicits An Overlapping But Temporally Distinct Transcriptional Profile From Contextual Fear Conditioning 


#### Abstract

Hippocampus-dependent learning is known to induce changes in gene expression, but information on gene expression differences between different learning paradigms that require the hippocampus is limited. The bulk of studies investigating RNA expression after learning use the contextual fear conditioning task, which couples a novel environment with a footshock. Although contextual fear conditioning has been useful in discovering gene targets, gene expression after spatial memory tasks has received less attention. In this study, we used the object-location memory task and studied gene expression at two time points after learning in a high-throughput manner using a microfluidic qPCR approach. We found that expression of the classic immediate-early genes changes after object-location training in a fashion similar to that observed after contextual fear conditioning. However, the temporal dynamics of gene expression are different between the two tasks, with object-location memory producing gene expression changes that last at least 2 hours. Our findings indicate that different training paradigms may give rise to distinct temporal dynamics of gene expression after learning.


## Introduction

Long-term memory is critical to our lives, yet the molecular mechanisms that create and stabilize memories are still poorly understood. The hippocampus, which encodes contextual information, has been heavily studied in an effort to better understand these mechanisms. Transcription is required to convert labile short-term memories into stable long-term memories during the period of memory consolidation [11, 108]. The expression of many genes is regulated within the first hour after learning in the hippocampus [61, 70-72, 138, 139]. Epigenetic mechanisms, such as histone acetylation, can modulate this transcription to enhance or dampen long-term memory formation $[23,24,26,27,51,55,58,63]$.

Most research into transcriptional regulation in the hippocampus has used contextual fear conditioning as the paradigm to test learning and memory [70, 71, 111, 113]. This is primarily because contextual fear conditioning produces a robust memory that has a well-defined time of acquisition due to the requirement of only a single training session [2]. Although this task has proven useful for dissecting the phases of memory and mapping the transcriptional landscape after learning, it also introduces a footshock that can be stressful to the animal. It is therefore important to study gene expression in other memory tasks that are more similar to the learning events that occur in daily life. Spatial learning requires the hippocampus and can be measured using the Morris water maze, Barnes maze, or object-location memory (OLM) tasks that do not require a footshock. These spatial tasks are also known to regulate transcription in the hippocampus, including many of the same genes and processes required for contextual fear memory $[56,76,77,110,112,140-144]$. There is evidence that contextual and spatial learning in the hippocampus can utilize different molecular pathways [130], so gene expression may also differ after these two tasks. Like contextual fear memory,

OLM is a hippocampus-dependent task [43]. However, the targets and temporal resolution of the gene expression changes after OLM have not been thoroughly studied. The goal of this study was to investigate the transcriptional profile that occurs within the first transcriptional wave after OLM learning $[16,108]$ and compare this transcriptional profile to that of contextual fear conditioning. Gene expression changes within this window after fear conditioning are typically highest 30 minutes after training and return to baseline by 2 hours [61, 70, 109]. Using a Fluidigm HD microfluidic high-throughput qPCR system, we examined expression of 96 different candidate genes at both 30 minutes and 2 hours after OLM training in a single experiment. We found that the most commonly studied genes after fear conditioning show a similar profile after OLM. However, OLM produces long-lasting expression changes in a number of genes that are not observed after fear conditioning.

## Materials and Methods

## Subjects

Forty-two C57BL/6J mice were maintained under standard conditions with food and water available ad libitum. Adult male mice 3 months of age were kept on a 12-hr light/12-hr dark cycle with lights on at 7AM. All behavioral and biochemical experiments were performed during the light cycle with training starting at approximately 7AM (ZTO). All procedures were approved by the University of Pennsylvania Institutional Animal Care and Use Committee.

## Behavior

Object-location memory (OLM) was carried out as previously described [43, 56]. Briefly, naïve three month old male C57BI/6J mice were singly housed for a week and handled for $2 \mathrm{~min} / \mathrm{day}$ for five consecutive days prior to tissue collection. One animal per behavioral group was trained and dissected each day for 10 total days to allow all animals to be dissected at the same circadian time. Exploration was normal in all mice used in this experiment (data not shown). One animal per training session was tested in a 24 hr retrieval test the following day to ensure the training proceeded correctly. Half of the handled animals received OLM training, and half of the animals were left undisturbed on training day and were sacrificed at the same circadian time points as trained animals. On the day of training, OLM mice were given a single block of four 6 min trials with an inter-trial interval of 3 min . The animals were habituated to an empty arena with a black and white striped spatial cue on one wall in the first trial, followed by three trials of object exposure. Each mouse was exposed to three distinct objects: a rectangular metal tower, a glass bottle, and a white plastic cylinder that were arranged in a V-shaped spatial pattern in the arena. Objects were positioned in the arena with at least two inches of spacing around each object to allow free exploration of all objects. During the intertraining interval (ITI), animals were gently removed from the arenas, and the arenas and objects were cleaned with $70 \%$ ethanol. Objects were not moved during the ITI. Immediately following the final trial, animals were gently placed in their home cage, and returned to the colony room until tissue collection.

Fear conditioning was performed as previously described [55, 61] with handling for 3 days prior to conditioning. Briefly, the conditioning protocol entailed a single 2-sec, 1.5 mA footshock terminating at 2.5 minutes after placement of the mouse in the novel
chamber. Mice were left in the chamber for an additional 30 seconds and then returned to their homecage.

## RNA isolation

Hippocampi were dissected 30 minutes and 2 hours after the last training session into RNAlater (Qiagen, Valencia, CA) and frozen on dry ice. Tissue was homogenized using a TissueLyser system and RNA was extracted using the miRNeasy kit (Qiagen) according to the manufacturer's instructions.
cDNA synthesis and high-throughput qPCR
RNA concentration was determined using a NanoDrop spectrophotometer (Thermo Scientific, Waltham, MA) and $1 \mu \mathrm{~g}$ RNA was used in each RETROscript (Ambion, Austin, TX) cDNA synthesis reaction with random decamers, 10 x RT Buffer and no heat denaturation according to the manufacturer's protocol. Concentrated cDNA was used in a specific target reaction following the manufacturer's recommendations (Fluidigm Corp. South San Francisco, CA). Briefly, Taqman assays for all 96 probes were pooled to a concentration of $0.2 \mathrm{X}(1: 100)$ and 1.25 ul of the pooled assay mix was combined with 2.5ul 2X Taqman Preamp Master Mix (Life technologies, Carlsbad, CA) and $1.25 \mu \mathrm{l}$ cDNA. Taqman probe IDs can be found in Table S1. The preamplification reaction was cycled using the following protocol in a 7500 Fast Real-Time PCR system: 10 min at 95C, then 14 cycles of 95 C for 15 s followed by 60C for 4 min . Preamplified samples were diluted 1:5 using 1X TE. Samples were then delivered to the Molecular Profiling Core at the University of Pennsylvania, where they were run on a 96.96 Dynamic Array IFC on the Biomark HD machine (Fluidigm Corp).

For validation and the 120 minute fear conditioning experiment, cDNA reactions were diluted to $2 \mathrm{ng} / \mathrm{ul}$ in water, and real-time RT-PCR reactions were prepared in 384-well optical reaction plates with optical adhesive covers (Life technologies). Each reaction was composed of $2.25 \mu \mathrm{l}$ cDNA ( $2 \mathrm{ng} / \mathrm{ul}$ ), $2.5 \mu \mathrm{l} 2 \mathrm{x}$ Taqman Fast Universal Master Mix (Life Technologies), and $0.25 \mu \mathrm{l}$ of Taqman probe. Reactions were performed in triplicate on the Viia7 Real-Time PCR system (Life Technologies, Carlsbad, CA).

## Data analysis

High-throughput qPCR was analyzed using the Fluidigm Real Time PCR Analysis program and Microsoft Excel. Genes with at least one sample having an average $\mathrm{Ct} \geq 20$ were discarded as being non-expressed or failed reactions. This included Dnmt3b, Erbb2, Esrrg, Fosb, Hdac1, Hdac4, Jun, Nr6a1, Pparg, and Trdmt1, which brought the total number of genes tested to 86 . Relative quantification of gene expression between groups was performed using the $\Delta \Delta C t$ method as described previously [55]. The difference between each Ct and the average Ct for that gene was subtracted from the average of three housekeeper genes treated in the same manner. A p-value of 0.01 was used for significance to control for the number of $t$-tests performed. This $p$-value cutoff was chosen because we selected genes for analysis that we expected to change, and thus Bonferroni correction is too strict. This $1 \%$ chance of a type I error corresponds to one false positive per 100 t-tests. Because 86 t-tests were performed, this p-value would suggest less than 1 false positive in the data, limiting the amount of type I errors introduced by multiple testing.

## Results

## Immediate Early Genes Are Regulated 30 minutes after OLM training

We chose sixteen representative genes that have been studied 30 minutes after fear conditioning to examine expression profiles 30 minutes after OLM training. The genes were chosen for well-studied expression changes (Arc, Bdnf4, Egr1, Fos, Homer1), genes our lab has previously studied (Fos/2 [61], Gadd45 family [73]), or from microarray data (Btg2, Cpeb3, Histh2hab, Sik1, Sox18, Tob1, Tob2 [109]). cDNA samples underwent specific target amplification and were run on a 96.96 Fluidigm Biomark HD plate in triplicate ( 96 genes, 32 samples). The full list of Taqman assays is available in Table 3.1. Ten genes were excluded due to too low expression or a failed reaction, bringing the total number of genes tested to 86 (See Methods for genes). In all cases, immediate early gene (IEG) expression after OLM mirrored expression after fear conditioning (Figure 3.1). Previously studied genes including Arc, Bdnf4, Egr1, Fos and Homer1 were upregulated as anticipated (Arc 272\% p=4.7×10-8; Bdnf4 53\% p=3.3×10 ${ }^{-7}$; Egr1 $225 \% \mathrm{p}=4.4 \times 10^{-8}$; Fos $410 \% \mathrm{p}=1.2 \times 10^{-10}$; Homer1 $31 \% \mathrm{p}=1.4 \times 10^{-4}$ ) [70, 121, 139, 145]. The probe against Homer1 recognizes both Homer1a and Homer1c, but research from our lab and others suggests that this effect is primarily due to Homer1a [121]. Further investigation is required to investigate specific Homer1 isoforms regulated by OLM. Genes that our lab discovered to be regulated after contextual fear conditioning using microarrays [109], including Btg2 ( $27 \% \mathrm{p}=8.9 \times 10^{-6}$ ), Hist2h2ab ( $-26 \% \mathrm{p}=8.8 \times 10^{-4}$ ), Sik1 $\left(70 \% \mathrm{p}=1.3 \times 10^{-5}\right)$, Sox18 $(-21 \% \mathrm{p}=0.002)$, and $\operatorname{Tob} 2(30 \% \mathrm{p}=0.004)$ showed similar changes after OLM. The genes Gadd45b and Gadd $45 g$ showed increased expression ( $32 \% \mathrm{p}=0.004,32 \% \mathrm{p}=0.001$ ) while Gadd45a did not ( $\mathrm{p}=0.20$ ), as has been reported previously by our lab and others [73, 146]. This observation suggests that the most
commonly studied genes after contextual fear conditioning are similarly regulated after spatial behavioral tasks such as object-location memory.

## Nuclear Hormone Receptors Display a Limited Response to OLM

A subset of nuclear hormone receptors are known to be regulated 30 minutes after fear conditioning, including the Nr4a family of orphan nuclear receptors [61]. We tested all 37 nuclear hormone receptors that are expressed in the hippocampus for changes after OLM training (Figure 3.2). The Nr4a family of nuclear receptors (Nr4a1, Nr4a2, Nr4a3), which are known to be necessary for long-term fear memory [58, 61, 144], all displayed increased expression at 30 minutes after OLM. Rev-ErbA (NR1D1), COUP-TFII (NR2F2), and retinoid X receptor gamma ( Rxrg ) all showed decreased expression at 30 minutes. No other nuclear receptors were observed to respond to spatial learning at this time point or at 120 minutes after OLM (data not shown). This contrasts with the large number of nuclear receptors that our lab observed to change after fear conditioning training in our previous study [61], which included increased expression of 13 nuclear receptor genes between 30 and 120 minutes after training. These results may indicate transcriptional regulation of this class of genes depends on the training paradigm.

## Regulators of Transcription Show Limited Changes in Response to OLM

Histone acetylation is known to be a crucial regulator of transcription during memory consolidation $[23,24,26-28,51,55,58,143]$. To test whether expression levels of histone acetylation modifying enzymes are regulated by OLM, we tested all histone deacetylases (HDACs, Figure 3.3A) and 16 histone acetyltransferases (HATs, Figure 3.3B) representing each class of enzyme, including the HATs CBP and p300 that have been shown to be essential for memory formation. The probes against Hdac1 and 79

Hdac4 did not amplify and were discarded. None of the HATs tested showed a gene expression change, in contrast to previous reports showing changes in expression of CBP, p300 and PCAF after the Morris Water Maze [77]. However, Hdac7 displayed reduced expression after OLM. HDAC7 is a class Ila HDAC that has not been previously linked to memory formation. This may suggest a novel role for HDAC7 in hippocampusdependent memory formation. In addition to the regulators of histone acetylation, we chose ten genes that are known to regulate transcription in other ways. None of these genes showed any changes in transcription at 30 minutes after OLM training (Figure 3.4).

## OLM Induces Longer Lasting Gene Expression Changes than Fear Conditioning

In addition to the 30 minute timepoint that has shown such robust changes after fear conditioning, we also tested hippocampal samples taken 2 hours after OLM training to investigate the persistence of these transcriptional changes. Interestingly, a number of genes that are upregulated at 30 minutes remain elevated 2 hours after OLM training. This includes highly induced genes that appear to be slowly returning to baseline, such as Egr1 and Fos, but also genes that maintain a similar level of induction as observed at 30 minutes such as Bdnf4, Fos/2, Homer1, Nr4a2 and Nr4a3 (Figure 3.5A). Sin3a was not changed at 30 minutes, but shows a selective change at 2 hours. The gene expression profiles at 30 minutes and 2 hours for Arc, Egr1, Fos, Nr4a1, and Nr4a2 were confirmed by standard 384 -well qPCR (data not shown). To test whether these same genes show transcriptional changes after fear conditioning, we prepared cDNA from samples that were collected 2 hours after fear conditioning. None of the genes determined to change 2 hours after OLM showed a significant change 2 hours after fear
conditioning (Figure 3.5B), indicating a long-lasting gene expression response specific to OLM.

## Discussion

In this study, we investigated the transcriptional changes that occur in response to OLM training using powerful high-throughput qPCR technology and compared these changes to fear conditioning. In a single run, we were able to study 96 different genes in 2 different time points after OLM training with $n=8$ mice per group using microfluidic highthroughput qPCR. This type of throughput, flexibility, and consistency is not possible with any other qPCR technology. In addition to requiring more pipetting steps, standard qPCR would have required the same housekeepers to be run on each individual plate and limited the number of targets that could be tested. Using a high-throughput approach allowed us to reliably determine that gene expression changes after OLM last longer than similar expression changes after contextual fear.

Our study discovered that commonly studied IEGs, such as Fos and Arc, show similar expression differences after fear conditioning and after OLM, indicating overlap between contextual and spatial learning. In a previous study from our lab [61], we found that a number of nuclear receptors exhibit increased expression after contextual fear conditioning. Our current findings suggest a more limited regulation of this class of genes after OLM. It is unclear whether the wider regulation after fear conditioning is in response to the footshock or whether the timecourse of expression after OLM is different. As seen after fear conditioning, all 3 members of the Nr4a family of orphan nuclear receptors were upregulated after OLM. However, while Nr4a1 returned to baseline by 2 hours, Nr4a2 and Nr4a3 did not, suggesting that different processes may
regulate Nr4a1 than the other two family members. Future studies will aim to determine how expression increases of Nr 4 a 2 and Nr 4 a 3 are maintained after OLM training.

It is interesting to note that Hdac7 and Sin3a are regulated by OLM while HATs are not. This may suggest that relieving the negative repression of histone acetylation is a crucial step for long-term memory formation. Although class I HDACs have been heavily implicated in learning and memory [56, 58, 63, 101], class Ila HDACs have received less attention. A study by Agis-Balboa et al. demonstrated that loss of the class lla member HDAC5 impairs spatial memory [104], but those experiments used a complete knockout mouse line that has the potential for developmental or extrahippocampal effects. Our study is the first to observe changes in Hdac7 in response to learning in the hippocampus.

The most intriguing finding of this study was the long-lasting regulation of gene expression 2 hours after OLM, something that is not seen after fear conditioning. It might be expected that the fear of a footshock would produce a stronger transcriptional response in the hippocampus than would the spatial rearrangement of objects. There are a number of potential causes for this disparity, although the most likely explanation is that the multiple training sessions required for OLM induce a stronger response than the single shock training used by our lab for fear conditioning. It would be interesting to test whether a multiple shock fear conditioning protocol induces longer lasting gene expression changes. Also, there could be an association between the novel context and the novel objects formed during OLM training that is not present in fear conditioning. Testing mice in the context only, introducing novel objects, or altering the number of training trials could determine whether these changes are sufficient to elicit gene expression changes. Further, different molecular mechanisms may regulate contextual
and spatial learning [130]. Future studies can test for changes at the protein level, although mRNA and protein levels generally agree after learning [15, 147]. Additional investigation into later time points after OLM training will be required to see if gene expression changes that occur well after fear conditioning [145] also exist after OLM. It is interesting that not all genes with increased expression at the 30 minute timepoint remain elevated for 2 hours after OLM. Future studies will determine whether specific epigenetic modifications regulate this longer term maintenance of gene expression at particular genes.

## Figure Legends

Figure 3.1. Classic IEGs Show Expected Expression Changes after OLM Training. 16 genes that are known to be induced 30 minutes after contextual fear conditioning were studied 30 minutes after OLM training. Each gene tested displayed the expression change that would be expected after contextual fear conditioning, indicating these genes may represent a common transcriptional response to learning. All error bars denote s.e.m. and *indicates $p<0.01$.

Figure 3.2. Limited Expression Changes of Nuclear Receptors after OLM training. Because of the known involvement of the Nr4a nuclear receptor family in memory, we tested expression of all nuclear receptors expressed in the hippocampus 30 minutes after OLM training. The Nr4a family displayed increased expression after OLM, while NR1D1, NR2F2, and RXRg had reduced expression. All error bars denote s.e.m. and * indicates $\mathrm{p}<0.01$.

Figure 3.3. Modifiers of Histone Acetylation Display Limited Regulation after OLM Training. Histone modifying enzymes were tested for expression changes 30 minutes after OLM training. (A) Hdac7, a class Ila HDAC, was the only family member found to change expression after OLM. (B) No HATs were observed to change expression after OLM. All error bars denote s.e.m. and * indicates $p<0.01$.

Figure 3.4. No Changes in Other Transcriptional Regulators after OLM Training. Other genes that can regulate gene expression, including DNMTs, were tested 30
minutes after OLM training. No differences in any gene were observed. All error bars denote s.e.m. and *indicates $p<0.01$.

Figure 3.5. OLM Training Induces Long-Lasting Changes in Gene Expression Not Seen after Fear Conditioning. (A) Every gene was also tested 2 hours after OLM training to observe the maintenance of transcription. Genes shown in this figure are those that were changed at 2 hours after OLM, all other genes were unchanged. Sin3a was the only gene uniquely regulated at the 2 hour time point. (B) These same genes do not show gene expression changes 2 hours after contextual fear conditioning. All error bars denote s.e.m. and *indicates $p<0.01$.

Table 3.1. Taqman Assays Used In This Experiment.

## Contributions

This chapter was written by Shane G. Poplawski with suggestions by Ted Abel, Hannah Schoch and Karl Peter Giese. Experiments were planned by Shane Poplawski, Hannah Schoch and Mathieu Wimmer. Experiments were carried out by Shane Poplawski, Hannah Schoch, and Joshua Hawk. Behavioral scoring was performed by Jennifer Walsh. We thank Morgan Bridi, Giulia Porcari, and Robbert Havekes for constructive discussions and critical reading of the chapter.

This chapter was published in: Poplawski SG*, Schoch H*, Wimmer M, Hawk JD, Walsh JL, Giese KP, Abel T. "Object-Location Training Elicits an Overlapping, but Temporally Distinct Transcriptional Profile from Contextual Fear Conditioning" Neurobiology of Learning and Memory, 2014. 116C: p 90-95.

Figures
Figure 3.1


Figure 3.2


Figure 3.3


Figure 3.4


Figure 3.5


## Tables

Table 3.1

| Gene | Taqman ID |
| :---: | :---: |
| AR | Mm00442688_m1 |
| Arc | Mm01204954_g1 |
| Bdnf4 | Mm00432069_m1 |
| Btg2 | Mm00476162_m1 |
| Cdk12 | Mm00660704_m1 |
| Cdk5 | Mm01164910_m1 |
| Cdyl | Mm00515473_g1 |
| Clock | Mm00455950_m1 |
| COUP-Tfa (NR2f1) | Mm01354342_m1 |
| COUP-TFb (NR2f2) | Mm00772789_m1 |
| COUP-Tfy (NR2f6) | Mm01340321_m1 |
| Cpeb3 | Mm01204299_m1 |
| Creb1 | Mm01254160_m1 |
| Crebbp | Mm01342452_m1 |
| Dnmt1 | Mm01151062_g1 |
| Dnmt2 (Trdmt1) | Mm00438511_m1 |
| Dnmt3a | Mm00432881_m1 |
| Dnmt3b | Mm01240113_m1 |
| Egr1/Zif268 | Mm00656724_m1 |
| Elp3 | Mm00804536_m1 |
| Ep300 | Mm00625535_m1 |
| ERa (NR3a1, ESR1) | Mm00433149_m1 |
| Erbb2 | Mm01306793_g1 |


| Esrra (ERRa) | Mm00433143_m1 |
| :---: | :---: |
| Esrrb (ERRb) | Mm00442411_m1 |
| Esrrg (ERRg) | Mm00516267_m1 |
| Fos | Mm00487425_m1 |
| FosB | Mm00500401_m1 |
| Fosl2 | Mm00484442_m1 |
| Gadd45a | Mm00432802_m1 |
| Gadd45b | Mm00435123_m1 |
| Gadd45g | Mm00442225_m1 |
| Gapdh | Mm99999915_g1 |
| GCNF (NR6a1) | Mm00599848_m1 |
| GR, Nr3c1 | Mm00433832_m1 |
| Gtf3c4 (TFIIIC90) | Mm00557022_m1 |
| Hat1 | Mm00509140_m1 |
| Hdac1 | Mm01351187_m1 |
| Hdac10 | Mm01308119_g1 |
| Hdac11 | Mm00523422_m1 |
| Hdac2 | Mm00515108_m1 |
| Hdac3 | Mm00515916_m1 |
| Hdac4 | Mm01304741_m1 |
| Hdac5 | Mm00515917_m1 |
| Hdac6 | Mm00515945_m1 |
| Hdac7 | Mm00469520_m1 |
| Hdac8 | Mm01224980_m1 |
| Hdac9 | Mm00458454_m1 |
| Hist2h2ab | Mm01613463_s1 |


| Homer1 | Mm00516275_m1 |
| :---: | :---: |
| Hprt | Mm01545399_m1 |
| Jun | Mm00495062_s1 |
| Kat2a (GCN5L2) | Mm00517402_m1 |
| Kat2B (PCAF) | Mm00451387_m1 |
| Kat5 (TIP60) | Mm01231512_m1 |
| LXRb (NR1h2) | Mm00437265_g1 |
| Mgea5 (NCOAT) | Mm00452409_m1 |
| MR (NR3C2) | Mm01241596_m1 |
| Myst1 (MOF) | Mm00458911_m1 |
| Myst2 (HBO1/HBOA) | Mm00624391_m1 |
| Myst3 (MOZ) | Mm01211941_m1 |
| Myst4 (MOZ2) | Mm00450564_m1 |
| Ncoa1 (SRC1) | Mm00447958_m1 |
| Nr1h2 (LXRb) | Mm00437265_g1 |
| Nr1h3 (LXRa) | Mm00443451_m1 |
| Nr4a1 | Mm00439358_m1 |
| Nr4a2 | Mm00443056_m1 |
| Nr4a3 | Mm00450074_m1 |
| PPARa | Mm00440939_m1 |
| PPARd | Mm00803184_m1 |
| PPARg | Mm01184322_m1 |
| PR (NR3C3, PGR) | Mm00435628_m1 |
| RARa | Mm01296312_m1 |
| RARb | Mm01319677_m1 |
| RARg | Mm00441091_m1 |


| Rb1 | Mm00485586_m1 |
| :---: | :---: |
| Rbbp7 | Mm01702744_mH |
| REV-ERBa (Nr1d1) | Mm00520708_m1 |
| REV-ERBb (Nr1d2) | Mm01310356_g1 |
| RORa | Mm01173766_m1 |
| RORb | Mm01204855_m1 |
| RXRa | Mm01332431_m1 |
| RXRb | Mm00441193_m1 |
| RXRg | Mm00436410_m1 |
| Sik1 | Mm00440317_m1 |
| Sin3a | Mm00488256_m1 |
| Sin3b | Mm00550123_m1 |
| Sox18 | Mm00656049_gH |
| TLX (NR2e1) | Mm00455855_m1 |
| Tob1 | Mm01204299_m1 |
| Tob2 | Mm00451524_s1 |
| TR2 (NR2C1) | Mm00449123_m1 |
| TR4 (NR2c2) | Mm01182440_m1 |
| TRa (NR1a1, Thra) | Mm00579691_m1 |
| TRb (NR1a2, Thrb) | Mm00437044_m1 |
| Tuba4a | Mm00849767_s1 |

## CHAPTER 4: Chromatin Accessibility Is Increased After Learning


#### Abstract

Transcription is a tightly regulated process that can be controlled by changes to the epigenome, including histone modifications and chromatin accessibility. Although it is well established that transcription during memory consolidation is crucial for long-term memory, little is known about how this transcription is regulated. Histone acetylation is known to increase in bulk extracts after contextual fear conditioning and pharmacologically increasing histone acetylation enhances memory formation. Increasing histone acetylation would be expected to "loosen" the interaction between DNA and histones and increase accessibility. Using high-throughput sequencing methods, we investigated the genome-wide changes in histone acetylation and chromatin accessibility that occur after learning. We discovered a genome-wide increase in chromatin accessibility after learning that has not been previously reported. Surprisingly, this increase in accessibility was not accompanied by a corresponding increase in H3K9/14 acetylation or change in nucleosome positioning. We propose that a combination of histone and DNA modifications act in concert to regulate chromatin accessibility after learning.


## Introduction

Despite 50 years of evidence that transcription and protein synthesis are required for long-term memory formation [11, 148], little is understood about the regulation of this gene expression. CREB and CREB-binding protein (CBP) are transcriptional regulators that are known to be crucial for this process [23-28, 149]. Besides being a CREB coactivator [150], CBP is also a histone acetyltransferase (HAT) that acetylates the N terminal tails of histone proteins [24]. This has led to numerous studies linking histone acetylation to long-term memory, including the finding that memory can be enhanced by blocking histone deacetylase (HDAC) proteins that remove acetyl groups [51, 55, 143]. We therefore hypothesized that histone acetylation is the critical epigenetic modification that regulates the gene transcription necessary for memory.

One way in which histone acetylation is thought to increase transcription is by reducing the positive charge of lysine residues, thereby weakening the interaction between histone proteins and the negatively charged DNA [35]. This weakening would be expected to increase the accessibility of chromatin to transcription factors and basal transcriptional machinery, allowing gene expression to occur [36]. Histone acetylation works in concert with a number of other histone and DNA modifications to form a "code" that can regulate gene expression [30]. Histone acetylation can also be "read" by proteins containing a bromodomain, including nucleosome remodeler Brg1 [151], further increasing the accessibility of chromatin. Because of the known association with Brg1, histone acetylation can lead to nucleosome repositioning. Nucleosome repositioning can include shifting or displacement of nucleosomes that alter chromatin accessibility [152].

Therefore, the increase in histone acetylation after learning [51,53] would be expected to result in increased chromatin accessibility to facilitate gene expression.

In this study, we used high-throughput sequencing techniques to study genomewide histone acetylation after learning using ChIP-seq. Specifically, we investigated H3K9/14ac, a mark that our lab has previously seen to increase at the Nr4a2 promoter after contextual fear conditioning [55]. Increases in H3K9ac and H3K14ac after learning have also been noted previously in bulk histone extracts [51, 53]. We also used Sonoseq [82] to study chromatin accessibility and MNase-seq [153] to study nucleosome positioning, two potential downstream effects of histone modifications. We found that H3K9/14ac shows a small increase in response to fear conditioning, but that this change is not large enough to be statistically significant at any individual gene. However, chromatin accessibility shows a large increase in response to learning that is not due to nucleosome repositioning. The regions showing increases after learning are enriched within gene bodies and show overlap with genes that are alternatively spliced after learning and those that regulate autism, a disorder associated with cognitive deficits. Publicly available H3K4me3 and H3K27me3 data sets also show high overlap with differential Sono-seq signal. This may suggest that other histone modifications may play a larger role in transcriptional regulation than $\mathrm{H} 3 \mathrm{~K} 9 / 14 \mathrm{ac}$, or that small changes in a number of modifications may cause a large change in accessibility.

## Materials and Methods

## Subjects

C57BL/6J mice were maintained under standard conditions with food and water available ad libitum. Adult male mice 3 months of age were kept on a $12-\mathrm{hr}$ light/12-hr
dark cycle with lights on at 7AM. All behavioral and biochemical experiments were performed during the light cycle with training starting at approximately 10AM (ZT3). All procedures were approved by the University of Pennsylvania Institutional Animal Care and Use Committee.

## Fear conditioning

Fear conditioning was performed as previously described [55, 61]. Briefly, mice were handled for 3 days prior to training, placed in a novel context within a soundproof chamber and allowed to explore for 148 seconds. A 2 second 1.5 mA footshock was delivered and mice were kept in the novel context for an additional 30 seconds. Mice were removed to their homecage after training. One mouse was trained and dissected per day to allow for dissection to occur at the same circadian time every day.

## Histone acetylation ChIP-seq pilot study

ChIP was performed following the Covaris truChIP Tissue Chromatin Shearing Kit (Covaris Woburn, MA) with minor modifications. Hippocampi were rapidly dissected 30 minutes after fear conditioning, finely chopped, and placed in $500 \mu \mathrm{l} 1 \%$ formaldehyde for 5 minutes. $28 \mu \mathrm{l}$ Covaris quenching buffer was added and samples were rocked for an additional 5 minutes. After two washes in PBS containing protease inhibitor, crosslinked tissue was frozen on dry ice. Each sample was homogenized in 1 ml Covaris lysis buffer containing protease inhibitors, washed in PBS, and resuspended in $280 \mu \mathrm{l}$ Covaris SDS buffer. Samples were split into two MICROtubes, sonicated in a Covaris S220 ultrasonicator, combined and precleared with $50 \mu \mathrm{l} 50 \%$ protein G slurry (Life Technologies Grand Island, NY). The supernatant was then split into immunoprecipitations for H3K9/14ac (Millipore \#06-599), H3K18ac (Cell Signaling 99
\#9675), or H3K27ac (Abcam \#ab4729) (2 $\mu \mathrm{g}$ antibody per IP) or set aside as an input sample. After rocking overnight, $80 \mu \mathrm{l} 50 \%$ protein G slurry was added to each IP and samples were rocked for 2 hours at $4^{\circ} \mathrm{C}$. Beads were washed for 5 min at RT with the following chilled buffers: Low Salt (0.1\% SDS, 1\% TX-100, 2 mM EDTA, 20 mM Tris- HCl pH8, 150 mM NaCl ) High Salt ( $0.1 \%$ SDS, $1 \%$ TX-100, 2 mM EDTA, 20 mM Tris-HCl pH8, 500 mM NaCl ), LiCl (1\% NP-40, 1\% deoxycholate, 1 mM EDTA, 10 mM Tris-HCl pH8, 250 mM LiCl), 1x TE, 1x TE. Samples were then eluted twice in $100 \mu \mathrm{l}$ ChIP Elution Buffer ( $1 \% \mathrm{SDS}, 0.1 \mathrm{M} \mathrm{NaHCO} 3$ ), $8 \mu \mathrm{MM} \mathrm{NaCl}$ was added, and samples were incubated at $65^{\circ} \mathrm{C}$ overnight. The next day, $1 \mu \mathrm{l}$ Proteinase K (Roche Diagnostics Indianapolis, IN) was added and samples were incubated an additional hour at $65^{\circ} \mathrm{C}$, then $1 \mu \mathrm{RNaseA}$ was added and samples were incubated for 30 min at $37^{\circ} \mathrm{C}$. Samples were purified using a standard phenol:chloroform extraction and ethanol precipitation.

## Sono-seq and H3K9/14ac ChIP-seq DNA preparation

ChIP was performed as previously described [55], and Sono-seq analysis was performed using the input samples of this experiment. Thirty minutes after fear conditioning, mice were sacrificed and the hippocampus was rapidly dissected. The hippocampus was finely chopped and placed into $500 \mu \mathrm{l} 2 \%$ PFA for 10 minutes to crosslink tissue. Crosslinking was stopped by the addition of $100 \mu \mathrm{l} 1 \mathrm{M}$ glycine, crosslinked tissue was washed $3 x$ in ice cold PBS containing protease inhibitor cocktail (Sigma-Aldrich St. Louis, MO), and crosslinked tissue was frozen at $-80^{\circ} \mathrm{C}$. Chromatin was prepared by dounce homogenizing the tissue in 1 ml ChIP cell lysis buffer ( 10 mM Tris $\mathrm{HCl} \mathrm{pH} 8.1,10 \mathrm{mM} \mathrm{NaCl}, 3 \mathrm{mM} \mathrm{MgCl} 2,0.5 \% \mathrm{NP}-40$ ), centrifuging at 5500 g for 5 minutes at $4^{\circ} \mathrm{C}$, removing the supernatant, and resuspending the pellet in $300 \mu \mathrm{ChIP}$ nuclear lysis buffer ( 50 mM Tris pH 8.1, 5 mM EDTA, 1\% SDS).

Chromatin was sonicated using a Bioruptor sonicator with the following conditions: 30 sec on, 1 min off for 15 minutes for two bouts with ice addition between bouts. Chromatin was centrifuged for 5 minutes at 14000 g at $4^{\circ} \mathrm{C}$ and the supernatant was removed. Chromatin size was tested by removing an aliquot, reversing crosslinks, cleaning DNA using the Qiagen MinElute PCR purification kit (Qiagen Valencia, CA), quantification by NanoDrop (Thermo Scientific Whatham, MA), and running the DNA on a $1 \%$ agarose gel. Sono-seq DNA was prepared by removing $1 \mu \mathrm{~g}$ sonicated chromatin. H3K9/14ac ChIP-seq DNA was prepared by removing $10 \mu \mathrm{~g}$ chromatin, adding $7.5 \mu \mathrm{~g}$ of H3K9/14ac antibody (Millipore \#06-599), and ChIP dilution buffer (16.7 mM Tris-Hcl pH 8.1, $1.1 \%$ TX-100, $0.01 \%$ SDS, $167 \mathrm{mM} \mathrm{NaCl}, 1.2 \mathrm{mM}$ EDTA) to 1 mL . IPs were rocked at $4^{\circ} \mathrm{C}$ overnight. Protein G incubation and washes were performed as described for the histone acetylation pilot experiment. The next morning, $4 \mu \mathrm{l} 0.5 \mathrm{M}$ EDTA, $8 \mu \mathrm{l} 1 \mathrm{M}$ TrisHCl pH 7.5 , and $1 \mu \mathrm{l}$ Proteinase K (Roche Diagnostics) were added and samples were incubated an additional hour at $65^{\circ} \mathrm{C}$. Samples were purified using the Qiagen MinElute PCR purification kit.

## MNase-seq DNA preparation

MNase-seq [153] was performed by dissecting mice 30 minutes after contextual fear conditioning, rapidly dissecting the hippocampus, and immediately dounce homogenizing in $500 \mu \mathrm{l}$ Douncing buffer ( 10 mM Tris, 4 mM MgCl 2 , 1 mM CaCl 2 adjust pH of solution to 7.5 ) containing protease inhibitor cocktail. $2 \mu \mathrm{l}$ micrococcal nuclease (NEB Ipswich, MA) was added to each sample and they were incubated at $37^{\circ} \mathrm{C}$ for 16 min. $125 \mu \mathrm{l} 0.5 \mathrm{M}$ EDTA was added to stop the reaction, and $375 \mu \mathrm{l}$ water was added to bring the volume to 1 ml . Samples were rocked at $4^{\circ} \mathrm{C}$ for 1 hour, precleared with $50 \mu \mathrm{l}$ 101
protein $G$ agarose beads for 30 minutes, and the supernatant was removed. An aliquot from each sample was proteinase $K$ and RNase $A$ (NEB) treated, quantified by NanoDrop, and run on a $2 \%$ agarose gel to ensure mononucleosomes were selectively isolated.

## Library Preparation

Library preparation for ChIP-seq pilot study was performed using the KAPA library preparation kit (KAPA Biosystems Boston, MA) according to the manufacturer's instructions with $3 \mu \mathrm{M}$ adapters to account for low concentration samples. Size selection was performed using AMPure XP beads (Beckman Coulter Brea, CA) by adding $40 \mu \mathrm{l}$ beads to $50 \mu \mathrm{l}$ of sample, discarding the beads, adding $9.5 \mu \mathrm{~L}$ beads to the supernatant and eluting. Ideal PCR cycles were determined for each set of samples and 18 cycles was used for $\mathrm{H} 3 \mathrm{~K} 9 / 14 \mathrm{ac}, 15$ cycles for H3K18ac, and 13 cycles for H3K27ac.

Library preparation for Sono-seq and H3K9/14ac ChIP-seq samples was performed using a combination of the ChIP-seq DNA sample prep kit and the Multiplexing Sample Preparation Oligonucleotide Kit (Illumina San Diego, CA). End repair and adenylation were performed as described in the ChIP protocol, while adapter ligation was performed as described in the Multiplexing protocol with the modification that adapters were diluted 1:10 for H3K9/14ac samples to account for reduced input. A gel slice 300bp +/- 25bp was cut from the gel for size selection, the Qiagen Gel Extraction kit was used to purify DNA and 23 PCR cycles were used following the PCR reaction setup described in the Multiplexing kit. The final product was run on a gel and purified by the Qiagen Gel Extraction kit again.

Library preparation for MNase-seq libraries was performed using the KAPA library preparation kit. $2 \mu \mathrm{~g}$ of each sample was used according to the manufacturer's instructions. The mononucleosome band was gel extracted and 7 cycles of PCR were determined sufficient to obtain a suitable library concentration.

## Sequencing

Libraries were quantified using the KAPA Library Quantification kit, normalized to 10 nM , and submitted to the PGFI sequencing core (Sono-seq and H3K9/14ac) or NextGeneration Sequencing Core (ChIP-seq pilot study, MNase-seq) at the University of Pennsylvania for 100bp single-end sequencing on an Illumina HiSeq2000. The histone acetylation pilot experiment produced an average of 57 million input reads, 57 million H3K9/14ac reads, 57 million H3K18ac reads, and 56 million H3K27ac reads. Sono-seq samples were run in their own lane producing an average of 148 million reads per sample. H3K9/14ac ChIP-seq samples were multiplexed 3 per lane producing an average of 45 million reads per sample. MNase-seq samples were multiplexed 2 per lane producing an average of 99 million reads per sample.

## Analysis

The histone acetylation pilot experiment was analyzed using uniquely aligned reads +/1 kb around each promoter. After upper quantile normalization [125], an arbitrary 2-fold cutoff was applied and the resulting differences were reported.

Sono-seq and H3K9/14ac samples were analyzed using a custom algorithm. Aligned reads cover $\sim 25 \%$ of the genome, showing that only $1 / 4$ of the chromatin is accessible in the hippocampus. Aligned reads with alignment score $<40$ as well as isolated duplicate 103
reads were removed for subsequent analysis. No published algorithm for the analysis of epigenetic HTS data exists that will incorporate biological replicates and detect statistically significant differences between treatments, such as learning and controls. We developed a novel approach to determine differences in Sono-seq peaks between learning and control mice in collaboration with Dr. Nancy Zhang (Wharton School of the University of Pennsylvania), which we named DEScan. Figure 4.1 depicts the series of computational steps taken to obtain differential peaks between learning and controls using DEScan. First, peaks were called separately on each sample using a variable window scan with a Poisson model and the surrounding 10kb as background. Second, peaks from all samples were aligned and only peaks present in at least $30 \%-50 \%$ of the samples were considered for further analysis. Third, aligned peaks were sampled to determine the dispersion parameter for a negative binomial model and subsequently a variable window scan was used within the aligned peaks to test for differential enrichment in FC vs HC using EdgeR [125] and the previously determined dispersion parameter. P-values were transformed to $z$-scores to ensure normality and only regions with a $z$-score $>1.9$ were reported ( 2 or more standard deviations from the average).

MNase-seq samples were analyzed using HTSeq [124] to produce a TSS plot, which was compared to the Sono-seq TSS plot.

## Results

Histone acetylation is a critical component regulating the transcription necessary for long-term memory formation $[23,24,27,51,55,58,63,143]$. Therefore, we used ChIPseq to test genome-wide histone acetylation occupancy throughout the genome 30
minutes after fear conditioning. We chose 3 common acetylation marks: H3K9/14ac and H3K18ac and H3K27ac. H3K9/14ac has been previously observed to increase at Nr4a genes at this timepoint by our lab [55] and both H3K9ac and H3K14ac have been demonstrated to change in bulk histone extracts after learning [51, 53]. H3K18ac and H3K27ac are two marks that have been shown to be dependent on CBP/p300 [54]. Although these marks are found primarily at promoters of expressed genes, H3K27ac is also found at active enhancers [154].

Our pilot ChIP-seq study ( $\mathrm{n}=1$ ) with these marks found that H3K9/14ac showed the largest number of promoters with $>2$ fold enrichment in either homecage or fear conditioned animals (Figure 4.2). Therefore, we followed up H3K9/14ac with ChIP-seq of $n=9$ samples to determine whether these changes at promoters were statistically significant. Genome-wide occupancy of H3K9/14ac at transcription start sites (TSSs) throughout the genome showed only a modest increase after fear conditioning (Figure 4.3). 38,211 regions in the genome were found to contain $\mathrm{H} 3 \mathrm{~K} 9 / 14 \mathrm{ac}$ peaks with an average length of 2.3 kb . The small increase seen when averaging over all TSSs was not observed to be statistically significant at any individual gene, indicating that the changes in H3K9/14ac after learning are too small to be studied using ChIP-seq on whole hippocampus.

H3K9/14ac is not the only histone acetylation mark that could be regulating the gene expression changes that occur after learning [51, 53]. Histone methylation, phosphorylation, ubiquitination and other marks work in concert to regulate transcriptional status [30, 155]. Indeed, histone phosphorylation [52] and methylation [156] have also been implicated in memory formation. However, studying every possible modification occurring during memory consolidation would be too costly and timeconsuming. Therefore, we focused our attention on the common target of these histone
modifications, which is increased chromatin accessibility. To study chromatin accessibility, we used Sono-seq [82]. Sono-seq combines weak sonication with low basepair size selection. The DNA that accumulates fastest at the low basepair size is the chromatin that was easiest to sonicate, which would be expected to be the most accessible chromatin. In our experiment, weak sonication of our ChIP-seq samples produced Sono-seq signal in our "input" samples. Although this is a potential confounding factor in the H3K9/14ac data, we found that H3K9/14ac signal does not correlate better with its paired input data as it does with input data from another sample (data not shown), suggesting the immunoprecipitation overcomes any bias introduced by small peaks in the input.

Sono-seq signal was found to correlate with DNase hypersensitivity and RNA polymerase II binding, two hallmarks of accessible chromatin, but not have a perfect overlap with either as previously reported [82]. Using Sono-seq, we saw a striking increase in accessibility at transcription start sites throughout the genome (Figure 4.4). Of the 36,186 regions in the genome that show Sono-seq peaks (800bp average length), 3064 regions ( $8.5 \%$ ) were determined to be significantly regulated ( $p$-value $<0.05$ ) after fear conditioning with an average length of 560 bp (Table 4.1). To determine whether these differential regions represent an important class of genes, we investigated where the signal was occurring. Differentially enriched regions were more likely to occur within gene bodies than at TSSs (Figure 4.5). Differences in the gene body suggest that mRNA elongation or splicing may be a target of chromatin accessibility. Promoter regions (upstream and overlapStart) also show an enrichment in differential regions (26\% of all signal, $35 \%$ of differential signal).

This increase in chromatin accessibility may regulate the gene expression necessary for long-term memory formation. Therefore, we first investigated whether 106
genes enriched in this data set correlated with genes identified in our previous RNA-seq study (Chapter 2). Although these regions do not show significant enrichment of genes regulated by contextual fear, these differential regions do exhibit significant enrichment for genes that showed differential alternative splicing in our previous study (Table 4.2). This finding agrees with the previous result that the differential Sono-seq peaks were enriched within gene bodies. Enrichment for alternatively spliced genes suggests that the Sono-seq changes that occur within gene bodies may be regulating splicing, but total levels of gene expression may or may not be affected.

Because these differential regions include many more genes in addition to those showing gene expression changes after learning, we also investigated whether they exhibit overlap with other gene sets related to cognitive functioning. We discovered that genes associated with autism (from the SFARI database) showed enrichment in the differential Sono-seq gene list (Table 4.2). Autism is a disorder with known cognitive dysfunction and often shows comorbidity with intellectual disability [157]. Although many of these genes don't show gene expression changes at 30 minutes after fear conditioning, increasing accessibility at these genes may still be affecting cognitive processing through regulation at other times. Genes associated with a brain disorder without cognitive dysfunction, epilepsy (from GenEpi), were not observed to have any enrichment in the differential Sono-seq signal.

We next sought to determine how this increased chromatin accessibility was being regulated. The increase in chromatin accessibility after fear conditioning could be due to many factors, but the most obvious explanation is that it represents a change in nucleosome positioning. To test this, we performed MNase-seq [153]. Micrococcal nuclease (MNase) is an enzyme that digests DNA. When incubated under optimized conditions, MNase can be used to preferentially digest linker chromatin between
nucleosomes, leaving behind only mononucleosome-bound DNA. The mononucleosome-bound DNA can then be sequenced to map genome-wide nucleosome occupancy. However, we found no difference in MNase-seq after learning (Figure 4.6). Therefore, nucleosome repositioning is not the major driver in changes in chromatin accessibility after learning.

Another possibility is that other histone modifications besides H3K9/14ac are regulating this accessibility. The histone code hypothesis [30] states that histone modifications will act in a combinatorial manner to form a code that can be read to cause downstream effects. Thus, a small change in a number of histone modifications, such as H3K9/14ac, could have large consequences for downstream effects such as chromatin accessibility. To test whether differential Sono-seq regions overlap with other histone modifications, we decided to look at other marks computationally. We compared publicly available ENCODE ChIP-seq data to our differential Sono-seq regions (Table 4.3). As anticipated, we found good overlap between differential Sono-seq regions and our H3K9/14ac ChIP-seq peaks (5.1\% of H3K9/14ac peaks overlap with differential Sonoseq peaks for a total of $64 \%$ of differential Sono-seq peaks). We also found overlap with both H3K4me3, an activating mark, and H3K27me3, a repressive mark, in the ENCODE data. Interestingly, these marks can be found in combination at "bivalent domains" regions that are repressed but poised for activation during development [158]. This may suggest a role for memory in regulating chromatin accessibility at poised genes. Fear conditioning may cause changes in a multitude of histone and DNA modifications that work cooperatively to regulate chromatin accessibility throughout the genome.

## Discussion

Although the regulation of gene expression after learning is not well understood, epigenetic mechanisms are beginning to be appreciated as an important player in the process. In this study, we sought to investigate the genes regulated by histone acetylation and chromatin accessibility. Our data indicate that the changes in H3K9/14ac are too small to be detected significantly when using whole hippocampal tissue. To our surprise, chromatin accessibility, which would be the expected effect of histone acetylation changes, was significantly changed after learning. The differential regions were enriched for gene bodies and promoter regions. Genes that show differential alternative splicing and those known to be implicated in autism spectrum disorders were enriched in differential Sono-seq regions, implicating the importance of this regulation.

Our data suggest that the increase in $\mathrm{H} 3 \mathrm{~K} 9 / 14 \mathrm{ac}$ at promoters is small and not significant when applied on a gene-by-gene basis. This could be due to the cellular dilution of our sample. The brain is a very complex organ and the hippocampus contains excitatory neurons, inhibitory neurons and glia. Although neurons account for roughly $50 \%$ of cells in the brain [159], these include both excitatory and inhibitory cells. Only 18$35 \%$ of excitatory neurons are activated by any particular memory trace in the hippocampus [17]. Therefore, changes in this subset of cells will be highly diluted by signal from surrounding cells, making changes look small and insignificant. We therefore suggest that sorting may be necessary to study epigenetic changes in the brain, especially when dealing with small responses such as that to contextual learning. Despite these limitations, we were able to detect a significant difference in chromatin accessibility by using Sono-seq after learning.

There are a number of explanations for the increase chromatin accessibility without a corresponding increase in H3K9/14ac, including nucleosome repositioning,
other epigenetic marks, transcription factor occupancy, or histone variant incorporation. We used MNase-seq to rule out nucleosome repositioning as the mechanism regulating this increase in chromatin accessibility. We believe the most likely explanation for the large increase in chromatin accessibility is that a number of histone and DNA modifications are working in concert to promote opening [30]. In this case, the change of any one modification (such as H3K9/14ac) may be small throughout the genome, but the effect of the combinatorial combinations of marks could be large. Based on computational comparisons of our data with publicly available ChIP-seq data, H3K4me3 and H3K27me3 appear to have a role in this process. Future studies will be needed to demonstrate overlap between regions of chromatin accessibility and histone modifications occurring at these sites after learning. However, we cannot rule out other possibilities such as transcription factor binding as the cause of this increase in accessibility.

Although the function of an increase in chromatin accessibility at such a large number of genes is uncertain, it is feasible that chromatin accessibility is acting as a gate to transcriptional regulation. Contextual conditioning may open this gate, and allow transcription or epigenetic factors to bind to their respective targets to activate or poise transcription. An alternative hypothesis would be that there are a larger number of gene expression changes than we currently appreciate, which may be more easily observed after cell sorting. Finally, it is also possible that these changes occurring within cell bodies are affecting processing of RNA such as splicing. Because we know that genes showing differential exon usage (Chapter 2) are enriched in differential Sono-seq regions, this is an exciting avenue for future research.

## Figure Legends

Figure 4.1. DEScan flowthrough. Our custom computational approach to detect differential peaks between treatment and controls using replicates is shown.

Figure 4.2. H3K9/14ac shows the most promoters with $\mathbf{> 2}$ fold differences. H3K9/14ac displayed 485 promoters with higher acetylation after fear conditioning and 265 with lower acetylation after fear conditioning. H3K18ac had 128 and 175 promoters respectively. H3K27ac showed fewer changes at promoters (4 and 9) in part because H3K27ac is found at both promoter and enhancer regions.

Figure 4.3. H3K9/14ac signal is slightly increased around the TSS after learning. Normalized read counts of H3K9/14ac data are plotted +/- 2 kb of the TSS. The dip is seen directly over the TSS because the nucleosome at that position is ejected during transcription. The read counts in the 3' peak appear to be slightly increased after fear conditioning.

Figure 4.4. Sono-seq signal has a dramatic increase around the TSS after learning. Normalized read counts of Sono-seq data are plotted $+/-2 \mathrm{~kb}$ of the TSS. There is a large increase surrounding the TSS in the fear conditioned group.

Figure 4.5. Differences in Sono-seq signal are primarily located within gene bodies. Pie chart shows the distribution of A) total Sono-seq regions and B) differential Sono-seq regions with regards to gene location. Intergenic (orange) represents regions
with no genes within 5 kb . Differential regions are more likely to occur near genes and specifically within gene bodies (green).

Figure 4.6. No difference in nucleosome positioning around the TSS after learning. Normalized read counts of MNase-seq data are plotted $+/-2 \mathrm{~kb}$ of the TSS. There is a no difference seen at any point around the TSS. The large dip just before the TSS and a well-positioned nucleosome just after the TSS are typical of this type of data.

Table 4.1. List of significantly regulated Sono-seq peaks after learning. Differential Sono-seq peaks with a Z-score greater than 1.9 ( $p$-value $<0.05$ ) are shown. Start and End are the respective genomic start at end sites of each peak. $Z$ denotes the $Z$-score. Distance measures the distance to the nearest TSS. Feature displays what this peak was called for the chart in Figure 4.6.

## Table 4.2. Comparison of differential Sono-seq regions to gene lists that regulate

 memory. The differential Sono-seq peaks were compared to other gene lists in a $2 \times 2$ contingency table using Chi-square tests with Yates correction. Significant enrichment is shown in red. Number of expressed genes is how many genes from that gene list are expressed and Expressed genes near differential Sono-seq region show how many of those expressed genes were near a differential Sono-seq peak. Overlap represents the number of expressed genes near differential Sono-seq region/number of differential Sono-seq regions (1694). Expected overlap represents the number of expressed genes in that list divided by the number of expressed genes in the hippocampus (18008). pvalues were found to be 0.40 (RNA-seq), 0.02 (diffSplice), 0.01 (autism), and 0.43 (GenEpi).Table 4.3. Comparison of publicly available ChIP-seq data to our differential Sonoseq regions. Differential regions compose $8.5 \%$ of all Sono-seq regions. Our histone acetylation peaks show $5.1 \%$ overlap, which cover $64 \%$ of differential Sono-seq peaks. As previously shown, Sono-seq signal has overlap with both RNA polymerase II binding and DNase hypersensivity, but not perfect overlap with either. H3K4me3 (7.8\% and $8.7 \%, 48 \%$ and $56 \%$ of differential peaks) and H3K27me3 (8.7\%, 34\% of differential peaks) appear to have large overlap with Sono-seq differential signals (red), implicating these marks in regulating chromatin accessibility after learning.

## Contributions

This chapter was written by Shane G. Poplawski with suggestions by Ted Abel. Lucia Peixoto co-wrote the Sono-seq and H3K9/14ac methods section, helped to design figures, and was instrumental in the computational work performed in this chapter. Experiments were planned and carried out by Lucia Peixoto, Shane Poplawski, and Mathieu Wimmer. Jieun Jeong (UPENN) assisted with analysis of the ChIP-seq pilot study and Nancy Zhang (Wharton) provided tremendous help with the analysis of the H3K9/14ac, Sono-seq and MNase-seq samples. Members of both the PGFI sequencing core as well as the NGSC core at UPENN provided assistance with library preparation, pooling and sequencing. We thank Giulia Porcari for constructive discussions and critical reading of the chapter.

Figure 4.1

High-throughput sequencing reads mapped to the mouse genome


## Stage I: Peak calling for multiple biological replicates

- Peak calling on all samples individually using Poisson models
- Align peaks to form common regions:
- For each sample, join peaks within 200 bp of each other
- Merge peaks across samples to form regions.
- Discard regions present in < $30 \%$ of the samples to assure reproducibility


## Stage II: Differential signal detection

- Sample a subset of regions to fit a common genomewide dispersion function for Negative Binomial model.
- If region >150bp, perform variable window scan (150 width every 50 bp)
- Test for differential signal between groups using a negative Binomial model and the dispersion parameter estimated above
- Transform p-values to z -scores

Figure 4.2


Figure 4.3


Figure 4.4


Figure 4.5

A Distribution of Sono-seq signal

$3 \%$

B Distribution of Sono-seq FCvsHC differences


Figure 4.6


Tables
Table 4.1

| GenelD | Start | End | Z | Distance | Feature |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000038545 | 46650358 | 46650808 | 2.38 | 21 | inside |
| ENSMUSG00000091175 | 91126296 | 91127646 | 2.35 | -35491 | intergenic |
| ENSMUSG00000058975 | 46397243 | 46397993 | 2.35 | 746 | inside |
| ENSMUSG00000021061 | 76710090 | 76710690 | 2.32 | 457 | overlap |
| ENSMUSG00000092763 | 99656981 | 99657381 | 2.32 | -761 | upstream |
| ENSMUSG00000043372 | 103132165 | 103132565 | 2.31 | -264 | overlap |
| ENSMUSG00000027618 | 156144135 | 156144535 | 2.31 | 51 | overlap |
| ENSMUSG00000058537 | 58224812 | 58225312 | 2.30 | 894 | inside |
| ENSMUSG00000041650 | 122525555 | 122526055 | 2.30 | -8769 | intergenic |
| ENSMUSG00000022419 | 55112141 | 55112641 | 2.30 | -176 | overlap |
| ENSMUSG00000021549 | 85289235 | 85289685 | 2.29 | -206 | upstream |
| ENSMUSG00000062087 | 70479907 | 70480307 | 2.29 | -97522 | intergenic |
| ENSMUSG00000079657 | 24533112 | 24533762 | 2.29 | 1098 | inside |
| ENSMUSG00000095972 | 67133377 | 67133927 | 2.29 | 41614 | intergenic |
| ENSMUSG00000018661 | 113649274 | 113649674 | 2.28 | 105 | inside |
| ENSMUSG00000096863 | 52240748 | 52242098 | 2.28 | -86141 | intergenic |
| ENSMUSG00000087993 | 80834863 | 80835613 | 2.27 | 6066 | intergenic |
| ENSMUSG00000035486 | 80356556 | 80357106 | 2.27 | 97 | inside |
| ENSMUSG00000021278 | 111275016 | 111275416 | 2.27 | 3905 | inside |
| ENSMUSG00000088128 | 76177521 | 76178071 | 2.27 | 36684 | intergenic |
| ENSMUSG00000004892 | 87999317 | 88000017 | 2.27 | 1039 | inside |
| ENSMUSG00000038156 | 113766062 | 113766712 | 2.27 | 64 | inside |
| ENSMUSG00000078670 | 73739960 | 73740410 | 2.27 | -347 | overlap |
| ENSMUSG00000055053 | 81430265 | 81430815 | 2.26 | 740 | inside |
| ENSMUSG00000098341 | 55471155 | 55471755 | 2.26 | 6621 | intergenic |
| ENSMUSG00000048915 | 62881908 | 62882658 | 2.26 | -591 | upstream |
| ENSMUSG00000096065 | 147595873 | 147596723 | 2.26 | 29012 | intergenic |
| ENSMUSG00000097960 | 120227930 | 120228480 | 2.26 | 300 | overlap |
| ENSMUSG00000041012 | 114843817 | 114844317 | 2.26 | 335 | overlap |
| ENSMUSG00000026207 | 75375444 | 75375944 | 2.25 | 147 | inside |
| ENSMUSG00000019897 | 105840552 | 105841252 | 2.25 | -515 | overlap |
| ENSMUSG00000037904 | 110978217 | 110978767 | 2.25 | 823 | inside |
| ENSMUSG00000035451 | 57542311 | 57543011 | 2.25 | 3810 | inside |
| ENSMUSG00000052727 | 99515235 | 99516485 | 2.25 | 1367 | inside |
| ENSMUSG00000033960 | 4635339 | 4635739 | 2.25 | 410 | inside |
| ENSMUSG00000023391 | 71546078 | 71546628 | 2.25 | 676 | inside |
| ENSMUSG00000083780 | 105798487 | 105799137 | 2.25 | 8618 | downstream |
| ENSMUSG00000099130 | 111276966 | 111277465 | 2.25 | -1872 | upstream |
| ENSMUSG00000046658 | 143254598 | 143255048 | 2.25 | 15424 | inside |
| ENSMUSG00000015957 | 98835066 | 98835566 | 2.25 | -46 | overlap |
| ENSMUSG00000039428 | 89404046 | 89404446 | 2.25 | 176 | overlap |
| ENSMUSG00000043794 | 25597000 | 25597550 | 2.25 | -12 | overlap |
| ENSMUSG00000025161 | 120959067 | 120959517 | 2.24 | 10587 | inside |


| ENSMUSG00000041730 | 32597844 | 32598244 | 2.24 | -2083 | upstream |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000050846 | 75941039 | 75941489 | 2.24 | 87 | inside |
| ENSMUSG00000073600 | 35654077 | 35655127 | 2.24 | 1110 | inside |
| ENSMUSG00000055044 | 40270954 | 40271354 | 2.24 | 888 | inside |
| ENSMUSG00000050592 | 32083919 | 32084519 | 2.24 | 938 | inside |
| ENSMUSG00000063600 | 29082090 | 29082790 | 2.24 | 67 | inside |
| ENSMUSG00000064254 | 24585394 | 24585894 | 2.24 | -2149 | upstream |
| ENSMUSG00000034799 | 71671159 | 71671659 | 2.24 | 598 | inside |
| ENSMUSG00000021273 | 63164339 | 63164839 | 2.23 | 13454 | inside |
| ENSMUSG00000039456 | 93831900 | 93832300 | 2.23 | -221 | overlap |
| ENSMUSG00000046138 | 29805113 | 29805563 | 2.23 | 896 | inside |
| ENSMUSG00000085438 | 119594235 | 119594785 | 2.23 | -62 | overlap |
| ENSMUSG00000027070 | 69585014 | 69585914 | 2.23 | 1051 | inside |
| ENSMUSG00000090116 | 99467139 | 99467539 | 2.23 | 39631 | inside |
| ENSMUSG00000031775 | 94695839 | 94696489 | 2.23 | 403 | overlap |
| ENSMUSG00000026608 | 189007610 | 189008010 | 2.22 | 230 | overlap |
| ENSMUSG00000059406 | 80888106 | 80888656 | 2.22 | 8386 | inside |
| ENSMUSG00000020152 | 20112478 | 20112978 | 2.22 | 435 | overlap |
| ENSMUSG00000051864 | 86620266 | 86620716 | 2.22 | 405807 | intergenic |
| ENSMUSG00000034730 | 74517866 | 74518516 | 2.22 | 1670 | inside |
| ENSMUSG00000016624 | 84854221 | 84855071 | 2.22 | 1828 | inside |
| ENSMUSG00000071033 | 91088370 | 91088920 | 2.22 | -123 | overlap |
| ENSMUSG00000045817 | 84184777 | 84185877 | 2.22 | 3170 | inside |
| ENSMUSG00000054057 | 18039358 | 18039908 | 2.22 | -1621 | upstream |
| ENSMUSG00000086657 | 14073608 | 14074658 | 2.22 | 326 | overlap |
| ENSMUSG00000034185 | 115897537 | 115898587 | 2.22 | 6871 | inside |
| ENSMUSG00000099149 | 19529593 | 19530393 | 2.22 | -25749 | intergenic |
| ENSMUSG00000015112 | 6216654 | 6217154 | 2.22 | 464 | overlap |
| ENSMUSG00000007946 | 101818499 | 101818949 | 2.22 | 186 | inside |
| ENSMUSG00000086604 | 5056480 | 5057030 | 2.22 | 631 | inside |
| ENSMUSG00000040725 | 25754494 | 25754894 | 2.22 | 236 | overlap |
| ENSMUSG00000032518 | 120127867 | 120128267 | 2.22 | 101 | inside |
| ENSMUSG00000056724 | 110653331 | 110653881 | 2.22 | 830 | inside |
| ENSMUSG00000056031 | 85843978 | 85844378 | 2.22 | -693 | upstream |
| ENSMUSG00000041040 | 59912450 | 59913050 | 2.21 | -556 | overlap |
| ENSMUSG00000041889 | 135374906 | 135375356 | 2.21 | 331 | overlap |
| ENSMUSG00000039239 | 186705304 | 186705854 | 2.21 | 688 | inside |
| ENSMUSG00000020178 | 75316584 | 75317184 | 2.21 | -359 | overlap |
| ENSMUSG00000035873 | 108331873 | 108332423 | 2.21 | -316 | overlap |
| ENSMUSG00000033697 | 76817021 | 76817921 | 2.21 | 1149 | inside |
| ENSMUSG00000033581 | 22161341 | 22161791 | 2.21 | 1958 | inside |
| ENSMUSG00000040852 | 84511909 | 84512309 | 2.21 | 14 | inside |
| ENSMUSG00000049112 | 112488739 | 112489339 | 2.21 | 1069 | inside |
| ENSMUSG00000046295 | 71406140 | 71406590 | 2.21 | 130 | inside |
| ENSMUSG00000097928 | 32009200 | 32010000 | 2.21 | 696 | inside |
| ENSMUSG00000097120 | 84201042 | 84201492 | 2.21 | -3375 | upstream |
| ENSMUSG00000019066 | 21918091 | 21918491 | 2.21 | 101 | overlap |
| ENSMUSG00000037419 | 14380891 | 14381291 | 2.21 | 351 | overlap |
| ENSMUSG00000026494 | 178527813 | 178528463 | 2.20 | -1312 | upstream |
| ENSMUSG00000026238 | 86525614 | 86526564 | 2.20 | -1122 | upstream |
| ENSMUSG00000090142 | 20432188 | 20433188 | 2.20 | 12372 | intergenic |


| ENSMUSG00000019817 | 13060623 | 13061173 | 2.20 | -30068 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000087036 | 118568665 | 118569165 | 2.20 | -89207 | intergenic |
| ENSMUSG00000018537 | 97699765 | 97700215 | 2.20 | 732 | inside |
| ENSMUSG00000020867 | 94491168 | 94491568 | 2.20 | -4989 | upstream |
| ENSMUSG00000044052 | 101173515 | 101174015 | 2.20 | 1928 | inside |
| ENSMUSG00000053263 | 3331826 | 3332226 | 2.20 | -648 | upstream |
| ENSMUSG00000097917 | 99627535 | 99627935 | 2.20 | 63694 | intergenic |
| ENSMUSG00000034168 | 86881877 | 86883077 | 2.20 | 2937 | inside |
| ENSMUSG00000062691 | 78916567 | 78917017 | 2.20 | 68 | inside |
| ENSMUSG00000052026 | 61013638 | 61014688 | 2.20 | 561 | overlap |
| ENSMUSG00000062661 | 31246001 | 31246401 | 2.20 | 178 | inside |
| ENSMUSG00000027490 | 154569629 | 154570029 | 2.20 | 263 | overlap |
| ENSMUSG00000027994 | 129969399 | 129969849 | 2.20 | 807 | inside |
| ENSMUSG00000072872 | 100125297 | 100125697 | 2.20 | 162144 | intergenic |
| ENSMUSG00000035314 | 99381366 | 99381866 | 2.20 | -183 | overlap |
| ENSMUSG00000030544 | 79792560 | 79793110 | 2.20 | 1228 | inside |
| ENSMUSG00000014837 | 105289122 | 105289622 | 2.20 | 406 | overlap |
| ENSMUSG00000097736 | 122568617 | 122569017 | 2.20 | -3880 | upstream |
| ENSMUSG00000031993 | 30426650 | 30427050 | 2.20 | -679 | upstream |
| ENSMUSG00000087514 | 69106814 | 69107214 | 2.19 | -74 | upstream |
| ENSMUSG00000084332 | 69376959 | 69378309 | 2.19 | -245660 | intergenic |
| ENSMUSG00000062075 | 80917613 | 80918113 | 2.19 | 632 | inside |
| ENSMUSG00000017776 | 75679327 | 75679777 | 2.19 | 68 | inside |
| ENSMUSG00000063564 | 51289580 | 51290030 | 2.19 | -340 | overlap |
| ENSMUSG00000061111 | 120549636 | 120550036 | 2.19 | 91 | overlap |
| ENSMUSG00000045201 | 15509735 | 15510185 | 2.19 | -70748 | intergenic |
| ENSMUSG00000047921 | 73060523 | 73060973 | 2.19 | 681 | inside |
| ENSMUSG00000058392 | 32036472 | 32036872 | 2.19 | 372 | inside |
| ENSMUSG00000044646 | 75819509 | 75819909 | 2.19 | -669 | upstream |
| ENSMUSG00000024483 | 36560377 | 36560877 | 2.19 | 390 | inside |
| ENSMUSG00000097593 | 80322124 | 80322624 | 2.19 | -1211 | upstream |
| ENSMUSG00000024423 | 12973578 | 12974078 | 2.19 | 1326 | inside |
| ENSMUSG00000038555 | 34840309 | 34840709 | 2.19 | -280 | overlap |
| ENSMUSG00000025213 | 45076425 | 45076925 | 2.19 | 1184 | inside |
| ENSMUSG00000035372 | 3708265 | 3708815 | 2.19 | -68 | overlap |
| ENSMUSG00000068290 | 130664223 | 130664623 | 2.19 | 436 | inside |
| ENSMUSG00000041912 | 94413090 | 94413490 | 2.19 | -206 | overlap |
| ENSMUSG00000085909 | 12402901 | 12403301 | 2.19 | 131502 | intergenic |
| ENSMUSG00000028610 | 107683763 | 107684213 | 2.19 | 467 | inside |
| ENSMUSG00000053839 | 31093184 | 31093634 | 2.19 | -1800 | upstream |
| ENSMUSG00000037541 | 143978027 | 143979327 | 2.19 | -23901 | intergenic |
| ENSMUSG00000071064 | 79027931 | 79029281 | 2.19 | -506 | overlap |
| ENSMUSG00000047897 | 87015500 | 87016100 | 2.19 | -37 | overlap |
| ENSMUSG00000097642 | 22532850 | 22533600 | 2.18 | -207914 | intergenic |
| ENSMUSG00000048402 | 119053261 | 119054061 | 2.18 | 358 | overlap |
| ENSMUSG00000020020 | 93834084 | 93834484 | 2.18 | 2529 | inside |
| ENSMUSG00000020325 | 79779656 | 79780156 | 2.18 | 2384 | inside |
| ENSMUSG00000020346 | 49250136 | 49250586 | 2.18 | 5945 | inside |
| ENSMUSG00000070407 | 63924927 | 63925527 | 2.18 | -2637 | upstream |
| ENSMUSG00000037275 | 58167936 | 58168336 | 2.18 | 603 | inside |
| ENSMUSG00000046605 | 121673194 | 121673644 | 2.18 | -41 | upstream |


| ENSMUSG00000035910 | 25055687 | 25056087 | 2.18 | -317 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000023938 | 45506657 | 45507057 | 2.18 | -184 | overlap |
| ENSMUSG00000096847 | 45545258 | 45545658 | 2.18 | 4419 | inside |
| ENSMUSG00000097330 | 37762959 | 37763659 | 2.18 | 5283 | inside |
| ENSMUSG00000083282 | 4855116 | 4855516 | 2.18 | -13 | overlap |
| ENSMUSG00000037771 | 158610903 | 158611553 | 2.18 | 136 | inside |
| ENSMUSG00000037211 | 37639840 | 37640540 | 2.18 | -107 | overlap |
| ENSMUSG00000037661 | 30855845 | 30856245 | 2.18 | -105 | overlap |
| ENSMUSG00000077161 | 62832648 | 62833198 | 2.18 | 20220 | intergenic |
| ENSMUSG00000094832 | 22745761 | 22746461 | 2.18 | -29869 | intergenic |
| ENSMUSG00000037428 | 137032787 | 137033237 | 2.18 | 2492 | inside |
| ENSMUSG00000055204 | 90365326 | 90365826 | 2.18 | 859 | inside |
| ENSMUSG00000044341 | 119744332 | 119744782 | 2.18 | 4406 | downstream |
| ENSMUSG00000030760 | 98309116 | 98309666 | 2.18 | 12092 | inside |
| ENSMUSG00000032504 | 113708067 | 113708517 | 2.18 | 192 | overlap |
| ENSMUSG00000088730 | 15634811 | 15635461 | 2.17 | 650141 | intergenic |
| ENSMUSG00000018334 | 79146377 | 79146777 | 2.17 | 30 | overlap |
| ENSMUSG00000020340 | 46311636 | 46312036 | 2.17 | 1223 | inside |
| ENSMUSG00000044847 | 45944727 | 45945327 | 2.17 | 208 | overlap |
| ENSMUSG00000063129 | 55538457 | 55538907 | 2.17 | -26782 | intergenic |
| ENSMUSG00000021670 | 96670585 | 96671135 | 2.17 | 351 | overlap |
| ENSMUSG00000036606 | 89167239 | 89168589 | 2.17 | 3881 | inside |
| ENSMUSG00000047428 | 46297844 | 46298294 | 2.17 | 423 | inside |
| ENSMUSG00000001870 | 75005270 | 75005770 | 2.17 | -298 | overlap |
| ENSMUSG00000045817 | 84186427 | 84186877 | 2.17 | 1520 | inside |
| ENSMUSG00000092569 | 75342959 | 75343509 | 2.17 | 24129 | intergenic |
| ENSMUSG00000018822 | 42201626 | 42202026 | 2.17 | 626 | inside |
| ENSMUSG00000053896 | 68599286 | 68600386 | 2.17 | 16873 | inside |
| ENSMUSG00000068154 | 146222835 | 146223235 | 2.17 | 914 | inside |
| ENSMUSG00000026755 | 39008201 | 39008651 | 2.17 | 125 | inside |
| ENSMUSG00000050896 | 84886482 | 84886932 | 2.17 | 228 | overlap |
| ENSMUSG00000017817 | 163375349 | 163375749 | 2.17 | 22644 | inside |
| ENSMUSG00000008999 | 172940058 | 172940508 | 2.17 | 263 | overlap |
| ENSMUSG00000028262 | 144828325 | 144829625 | 2.17 | -8831 | intergenic |
| ENSMUSG00000082279 | 107322737 | 107323687 | 2.17 | 5762 | downstream |
| ENSMUSG00000029366 | 88765076 | 88765576 | 2.17 | 80 | inside |
| ENSMUSG00000030435 | 5061761 | 5062561 | 2.17 | -382 | overlap |
| ENSMUSG00000038497 | 13287130 | 13287580 | 2.17 | -883 | upstream |
| ENSMUSG00000079070 | 32884700 | 32885200 | 2.17 | 65326 | downstream |
| ENSMUSG00000033106 | 106210389 | 106210889 | 2.17 | 544 | inside |
| ENSMUSG00000019777 | 36974595 | 36975195 | 2.16 | 51 | inside |
| ENSMUSG00000034707 | 121365245 | 121365695 | 2.16 | 155 | inside |
| ENSMUSG00000085707 | 53463327 | 53463877 | 2.16 | 286 | overlap |
| ENSMUSG00000010086 | 61449227 | 61449627 | 2.16 | 4904 | inside |
| ENSMUSG00000021265 | 108835844 | 108836244 | 2.16 | 39 | overlap |
| ENSMUSG00000040867 | 109033584 | 109033984 | 2.16 | 34633 | inside |
| ENSMUSG00000091105 | 21783788 | 21784188 | 2.16 | -7081 | intergenic |
| ENSMUSG00000040640 | 27621727 | 27622377 | 2.16 | -715 | upstream |
| ENSMUSG00000054863 | 87664816 | 87665266 | 2.16 | 39586 | inside |
| ENSMUSG00000088128 | 76180771 | 76182071 | 2.16 | 33434 | intergenic |
| ENSMUSG00000006740 | 6241365 | 6241765 | 2.16 | 809 | inside |


| ENSMUSG00000024360 | 34931277 | 34931727 | 2.16 | 730 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000042834 | 33384477 | 33385177 | 2.16 | 79552 | intergenic |
| ENSMUSG00000040913 | 45659976 | 45660376 | 2.16 | 336 | overlap |
| ENSMUSG00000050174 | 37481153 | 37481553 | 2.16 | -60942 | intergenic |
| ENSMUSG00000035069 | 103313637 | 103314237 | 2.16 | -175 | overlap |
| ENSMUSG00000046079 | 105700288 | 105700688 | 2.16 | 319 | inside |
| ENSMUSG00000084950 | 87980339 | 87980839 | 2.16 | -544 | upstream |
| ENSMUSG00000092318 | 12010383 | 12010783 | 2.16 | 5317 | downstream |
| ENSMUSG00000006315 | 30729057 | 30729507 | 2.16 | 477 | inside |
| ENSMUSG00000031627 | 46740654 | 46741254 | 2.16 | 909 | inside |
| ENSMUSG00000031986 | 124898054 | 124898554 | 2.16 | 168 | inside |
| ENSMUSG00000034518 | 74994040 | 74994540 | 2.16 | 684 | inside |
| ENSMUSG00000086225 | 100643416 | 100644266 | 2.16 | -17451 | intergenic |
| ENSMUSG00000032440 | 116174667 | 116175067 | 2.16 | 696 | inside |
| ENSMUSG00000039349 | 184882810 | 184883310 | 2.15 | 408 | overlap |
| ENSMUSG00000088582 | 148301958 | 148302508 | 2.15 | 25150 | intergenic |
| ENSMUSG00000036478 | 96617194 | 96617644 | 2.15 | 193 | inside |
| ENSMUSG00000055670 | 72796377 | 72796777 | 2.15 | 151 | inside |
| ENSMUSG00000048616 | 89301568 | 89301968 | 2.15 | 991 | inside |
| ENSMUSG00000020831 | 70237638 | 70238038 | 2.15 | 276 | overlap |
| ENSMUSG00000045440 | 55598811 | 55599561 | 2.15 | -106 | overlap |
| ENSMUSG00000097801 | 80518826 | 80519326 | 2.15 | 90 | overlap |
| ENSMUSG00000046314 | 45074158 | 45074608 | 2.15 | 335 | overlap |
| ENSMUSG00000038175 | 45389047 | 45389747 | 2.15 | -695 | overlap |
| ENSMUSG00000084651 | 109903058 | 109903558 | 2.15 | -751 | upstream |
| ENSMUSG00000042622 | 79347421 | 79348171 | 2.15 | -120 | overlap |
| ENSMUSG00000022360 | 58134546 | 58134946 | 2.15 | 536 | inside |
| ENSMUSG00000022450 | 82354071 | 82354621 | 2.15 | 220 | overlap |
| ENSMUSG00000041205 | 20240641 | 20241041 | 2.15 | 717 | inside |
| ENSMUSG00000073411 | 35262722 | 35263172 | 2.15 | -8 | overlap |
| ENSMUSG00000024421 | 12333854 | 12334304 | 2.15 | -170 | overlap |
| ENSMUSG00000097430 | 37143459 | 37143909 | 2.15 | 35063 | intergenic |
| ENSMUSG00000069833 | 8988876 | 8989326 | 2.15 | -408 | overlap |
| ENSMUSG00000037902 | 129593423 | 129593923 | 2.15 | 588 | inside |
| ENSMUSG00000059540 | 181680271 | 181680771 | 2.15 | -39 | overlap |
| ENSMUSG00000065454 | 180893616 | 180894116 | 2.15 | -415 | overlap |
| ENSMUSG00000025314 | 90579764 | 90580364 | 2.15 | 883 | inside |
| ENSMUSG00000037197 | 11553444 | 11553844 | 2.15 | 50709 | intergenic |
| ENSMUSG00000087100 | 172007471 | 172008121 | 2.15 | -3476 | upstream |
| ENSMUSG00000037625 | 31149717 | 31150267 | 2.15 | -229 | overlap |
| ENSMUSG00000033721 | 109340750 | 109341300 | 2.15 | 97 | inside |
| ENSMUSG00000029003 | 148130033 | 148130783 | 2.15 | -351 | overlap |
| ENSMUSG00000052520 | 96650989 | 96651589 | 2.15 | 13165 | inside |
| ENSMUSG00000033365 | 117914567 | 117915167 | 2.15 | 432 | overlap |
| ENSMUSG00000015942 | 134184157 | 134184557 | 2.15 | 138 | inside |
| ENSMUSG00000035266 | 100798174 | 100798574 | 2.15 | 424 | inside |
| ENSMUSG00000043614 | 135077707 | 135078107 | 2.15 | 559 | inside |
| ENSMUSG00000030087 | 90462724 | 90463274 | 2.15 | 148 | inside |
| ENSMUSG00000003423 | 45154156 | 45154556 | 2.15 | -147 | overlap |
| ENSMUSG00000021217 | 36698154 | 36698554 | 2.15 | 36 | inside |
| ENSMUSG00000009545 | 143106808 | 143107358 | 2.15 | -446 | overlap |


| ENSMUSG00000041775 | 138846143 | 138846793 | 2.15 | 130 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000066180 | 126291859 | 126292409 | 2.15 | 8057 | intergenic |
| ENSMUSG00000093241 | 108956072 | 108956622 | 2.15 | 19212 | intergenic |
| ENSMUSG00000031995 | 31131700 | 31132150 | 2.15 | 153 | overlap |
| ENSMUSG00000074345 | 54068078 | 54068478 | 2.15 | 333 | overlap |
| ENSMUSG00000026563 | 166379202 | 166379602 | 2.14 | 105 | inside |
| ENSMUSG00000048960 | 10993107 | 10993707 | 2.14 | -358 | overlap |
| ENSMUSG00000026080 | 38897388 | 38897938 | 2.14 | 772 | inside |
| ENSMUSG00000019997 | 24595473 | 24596073 | 2.14 | 31 | inside |
| ENSMUSG00000020218 | 121033695 | 121034395 | 2.14 | -265 | overlap |
| ENSMUSG00000000282 | 74837627 | 74838577 | 2.14 | 6707 | inside |
| ENSMUSG00000040610 | 33202438 | 33203038 | 2.14 | 1150 | inside |
| ENSMUSG00000000804 | 85138630 | 85139930 | 2.14 | 1531 | inside |
| ENSMUSG00000001504 | 72631468 | 72632068 | 2.14 | 2648 | inside |
| ENSMUSG00000021991 | 29720727 | 29721727 | 2.14 | 1137 | inside |
| ENSMUSG00000036158 | 93519120 | 93519570 | 2.14 | 76771 | inside |
| ENSMUSG00000068284 | 44173560 | 44174010 | 2.14 | 314 | inside |
| ENSMUSG00000089774 | 92058700 | 92059100 | 2.14 | 378 | inside |
| ENSMUSG00000094726 | 82000307 | 82001007 | 2.14 | -542922 | intergenic |
| ENSMUSG00000034786 | 34587301 | 34587701 | 2.14 | -2505 | upstream |
| ENSMUSG00000043991 | 36287327 | 36288027 | 2.14 | 6230 | inside |
| ENSMUSG00000053441 | 58836727 | 58837677 | 2.14 | -37 | overlap |
| ENSMUSG00000089290 | 23803630 | 23804030 | 2.14 | 13741 | intergenic |
| ENSMUSG00000075227 | 6061665 | 6062065 | 2.14 | 473 | inside |
| ENSMUSG00000025171 | 41981219 | 41981719 | 2.14 | -544 | upstream |
| ENSMUSG00000097787 | 32388425 | 32389225 | 2.14 | -791 | overlap |
| ENSMUSG00000074793 | 131127090 | 131127540 | 2.14 | -190 | overlap |
| ENSMUSG00000026849 | 30967501 | 30968201 | 2.14 | 432 | overlap |
| ENSMUSG00000042272 | 77279969 | 77280369 | 2.14 | 623 | inside |
| ENSMUSG00000085388 | 19445944 | 19446744 | 2.14 | 730 | overlap |
| ENSMUSG00000008604 | 88553767 | 88554167 | 2.14 | 51 | inside |
| ENSMUSG00000013622 | 31048284 | 31049184 | 2.14 | -28 | overlap |
| ENSMUSG00000035187 | 101663738 | 101664288 | 2.14 | 1488 | inside |
| ENSMUSG00000068328 | 83054299 | 83054849 | 2.14 | -354 | overlap |
| ENSMUSG00000001761 | 29735443 | 29735893 | 2.14 | -251 | overlap |
| ENSMUSG00000042087 | 117907354 | 117907854 | 2.14 | 406 | overlap |
| ENSMUSG00000086414 | 105761512 | 105762712 | 2.14 | -809 | overlap |
| ENSMUSG00000098708 | 63757577 | 63758177 | 2.14 | -138956 | intergenic |
| ENSMUSG00000052301 | 126847659 | 126848809 | 2.14 | 243 | inside |
| ENSMUSG00000048583 | 142667608 | 142668108 | 2.14 | -792 | upstream |
| ENSMUSG00000056043 | 35584394 | 35584844 | 2.14 | 1188 | inside |
| ENSMUSG00000062944 | 128238249 | 128238649 | 2.14 | -218 | upstream |
| ENSMUSG00000070526 | 62463727 | 62464377 | 2.14 | 783 | inside |
| ENSMUSG00000055148 | 72318873 | 72319523 | 2.14 | -160 | overlap |
| ENSMUSG00000031715 | 80739142 | 80739642 | 2.14 | 355 | overlap |
| ENSMUSG00000033021 | 75422513 | 75423013 | 2.13 | -13417 | intergenic |
| ENSMUSG00000019467 | 127188754 | 127189154 | 2.13 | 1300 | inside |
| ENSMUSG00000097586 | 22272945 | 22273445 | 2.13 | 8791 | intergenic |
| ENSMUSG00000034427 | 115859118 | 115859568 | 2.13 | -2494 | upstream |
| ENSMUSG00000099101 | 78178677 | 78179077 | 2.13 | -150 | includeFeature |
| ENSMUSG00000043099 | 75165277 | 75166427 | 2.13 | 4242 | inside |


| ENSMUSG00000087842 | 113229316 | 113230066 | 2.13 | 15167 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000057469 | 16811092 | 16811542 | 2.13 | 127 | inside |
| ENSMUSG00000042286 | 31176549 | 31177249 | 2.13 | -7908 | intergenic |
| ENSMUSG00000087836 | 30447641 | 30448041 | 2.13 | -142646 | intergenic |
| ENSMUSG00000039568 | 4879753 | 4880403 | 2.13 | 562 | overlap |
| ENSMUSG00000022983 | 90283608 | 90284058 | 2.13 | 817 | inside |
| ENSMUSG00000059791 | 35861211 | 35862461 | 2.13 | -107 | overlap |
| ENSMUSG00000046711 | 27555015 | 27555415 | 2.13 | -1605 | upstream |
| ENSMUSG00000024014 | 29490072 | 29490522 | 2.13 | -740 | upstream |
| ENSMUSG00000024163 | 24936272 | 24936922 | 2.13 | 705 | inside |
| ENSMUSG00000033717 | 54046025 | 54046825 | 2.13 | 843 | inside |
| ENSMUSG00000084837 | 144331049 | 144331449 | 2.13 | -410 | upstream |
| ENSMUSG00000026764 | 49619473 | 49619973 | 2.13 | 175 | inside |
| ENSMUSG00000086308 | 116066799 | 116067249 | 2.13 | -426 | overlap |
| ENSMUSG00000016386 | 106693252 | 106693652 | 2.13 | -17 | overlap |
| ENSMUSG00000026791 | 32981351 | 32981801 | 2.13 | 732 | inside |
| ENSMUSG00000035513 | 29252551 | 29252951 | 2.13 | 454 | inside |
| ENSMUSG00000027184 | 103797382 | 103797782 | 2.13 | 267 | overlap |
| ENSMUSG00000026754 | 39065319 | 39065719 | 2.13 | 222 | overlap |
| ENSMUSG00000027794 | 55181696 | 55182096 | 2.13 | -332 | overlap |
| ENSMUSG00000033147 | 101924302 | 101924802 | 2.13 | 151 | overlap |
| ENSMUSG00000070867 | 114406157 | 114406557 | 2.13 | -567 | upstream |
| ENSMUSG00000001089 | 136470077 | 136470577 | 2.13 | 316 | inside |
| ENSMUSG00000070717 | 132074657 | 132075207 | 2.13 | -40 | overlap |
| ENSMUSG00000034645 | 108217293 | 108217743 | 2.13 | 629 | inside |
| ENSMUSG00000087516 | 119680292 | 119680992 | 2.13 | -8927 | intergenic |
| ENSMUSG00000033706 | 85431828 | 85432228 | 2.13 | -161 | overlap |
| ENSMUSG00000030270 | 113283074 | 113283474 | 2.13 | 767 | inside |
| ENSMUSG00000030199 | 134035765 | 134036415 | 2.13 | 65 | inside |
| ENSMUSG00000034203 | 91472960 | 91473410 | 2.13 | 463 | inside |
| ENSMUSG00000005362 | 106799549 | 106800049 | 2.13 | 525 | inside |
| ENSMUSG00000030287 | 146501921 | 146502321 | 2.13 | 302 | overlap |
| ENSMUSG00000031074 | 144838651 | 144839551 | 2.13 | 568 | inside |
| ENSMUSG00000047371 | 127345482 | 127346032 | 2.13 | -168 | upstream |
| ENSMUSG00000031480 | 22227236 | 22227636 | 2.13 | -77 | overlap |
| ENSMUSG00000031737 | 92360390 | 92360790 | 2.13 | 2594 | inside |
| ENSMUSG00000034472 | 75212973 | 75213573 | 2.13 | -971 | upstream |
| ENSMUSG00000034796 | 123117391 | 123117791 | 2.13 | 17 | inside |
| ENSMUSG00000036611 | 25481500 | 25481900 | 2.13 | -47 | overlap |
| ENSMUSG00000026504 | 176813896 | 176814296 | 2.12 | -764 | upstream |
| ENSMUSG00000026686 | 167688396 | 167689446 | 2.12 | -843 | overlap |
| ENSMUSG00000038702 | 111864600 | 111865050 | 2.12 | 309 | overlap |
| ENSMUSG00000038160 | 44268237 | 44268687 | 2.12 | -121 | overlap |
| ENSMUSG00000020102 | 125388987 | 125389487 | 2.12 | -60024 | intergenic |
| ENSMUSG00000020770 | 116030436 | 116030836 | 2.12 | 114 | inside |
| ENSMUSG00000001552 | 100397515 | 100398015 | 2.12 | 248 | overlap |
| ENSMUSG00000077590 | 65356158 | 65357208 | 2.12 | 49812 | intergenic |
| ENSMUSG00000015002 | 65787125 | 65787525 | 2.12 | 91 | inside |
| ENSMUSG00000040605 | 97356407 | 97356807 | 2.12 | -321 | overlap |
| ENSMUSG00000085783 | 62664497 | 62664897 | 2.12 | -52456 | intergenic |
| ENSMUSG00000038168 | 26105517 | 26105917 | 2.12 | 267 | overlap |


| ENSMUSG00000089883 | 13844100 | 13844650 | 2.12 | 18703 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000024974 | 53599025 | 53600225 | 2.12 | -1373 | upstream |
| ENSMUSG00000052188 | 6531165 | 6531565 | 2.12 | 846 | inside |
| ENSMUSG00000027301 | 130576473 | 130576873 | 2.12 | 300 | inside |
| ENSMUSG00000074582 | 166805748 | 166806148 | 2.12 | 160 | inside |
| ENSMUSG00000027525 | 178142221 | 178142721 | 2.12 | 23246 | inside |
| ENSMUSG00000027315 | 119237535 | 119237985 | 2.12 | 173 | inside |
| ENSMUSG00000087455 | 59347928 | 59348328 | 2.12 | 39543 | intergenic |
| ENSMUSG00000027967 | 127633149 | 127633849 | 2.12 | 13 | inside |
| ENSMUSG00000028032 | 131564411 | 131564811 | 2.12 | -357 | overlap |
| ENSMUSG00000077253 | 110682444 | 110683244 | 2.12 | -117903 | intergenic |
| ENSMUSG00000028351 | 68832764 | 68833214 | 2.12 | 121633 | inside |
| ENSMUSG00000080412 | 120302020 | 120302770 | 2.12 | 14294 | intergenic |
| ENSMUSG00000060621 | 19519914 | 19520314 | 2.12 | 1183 | inside |
| ENSMUSG00000054716 | 127249099 | 127249499 | 2.12 | 4573 | inside |
| ENSMUSG00000012889 | 84130823 | 84131273 | 2.12 | 4834 | inside |
| ENSMUSG00000014786 | 105349108 | 105349508 | 2.12 | 850 | inside |
| ENSMUSG00000085795 | 26978567 | 26979217 | 2.12 | 1231 | inside |
| ENSMUSG00000002393 | 71380681 | 71381181 | 2.12 | 1279 | inside |
| ENSMUSG00000032178 | 21367891 | 21368741 | 2.12 | -128 | overlap |
| ENSMUSG00000046997 | 97018016 | 97018716 | 2.12 | 827 | inside |
| ENSMUSG00000071072 | 128058322 | 128058872 | 2.11 | -632 | upstream |
| ENSMUSG00000044199 | 81499056 | 81499556 | 2.11 | 1076 | inside |
| ENSMUSG00000020097 | 61146431 | 61147181 | 2.11 | 1272 | inside |
| ENSMUSG00000063760 | 31608461 | 31608861 | 2.11 | 723 | inside |
| ENSMUSG00000038594 | 53379037 | 53379437 | 2.11 | 814 | inside |
| ENSMUSG00000000976 | 83753027 | 83753577 | 2.11 | -669 | upstream |
| ENSMUSG00000086058 | 82930286 | 82930786 | 2.11 | 3096 | inside |
| ENSMUSG00000077270 | 65881168 | 65881768 | 2.11 | 315078 | intergenic |
| ENSMUSG00000052632 | 21112037 | 21112587 | 2.11 | 289 | inside |
| ENSMUSG00000088158 | 80380290 | 80380890 | 2.11 | 11461 | intergenic |
| ENSMUSG00000051111 | 96132085 | 96132635 | 2.11 | 492 | overlap |
| ENSMUSG00000021359 | 40731200 | 40731600 | 2.11 | 2623 | inside |
| ENSMUSG00000036422 | 79770219 | 79770769 | 2.11 | 1093 | inside |
| ENSMUSG00000072294 | 100049861 | 100050411 | 2.11 | 99903 | inside |
| ENSMUSG00000023484 | 99055615 | 99056215 | 2.11 | 441 | inside |
| ENSMUSG00000062901 | 20097772 | 20098322 | 2.11 | 218 | inside |
| ENSMUSG00000040785 | 94370116 | 94370516 | 2.11 | -502 | upstream |
| ENSMUSG00000060475 | 12991380 | 12991880 | 2.11 | 1166 | inside |
| ENSMUSG00000016498 | 29424626 | 29425026 | 2.11 | 13707 | inside |
| ENSMUSG00000024758 | 7482963 | 7483363 | 2.11 | 326 | overlap |
| ENSMUSG00000027007 | 79635502 | 79635952 | 2.11 | 150 | inside |
| ENSMUSG00000042821 | 167538171 | 167539021 | 2.11 | -24 | overlap |
| ENSMUSG00000003662 | 127247785 | 127248285 | 2.11 | 31 | overlap |
| ENSMUSG00000027171 | 104849469 | 104849919 | 2.11 | 407 | overlap |
| ENSMUSG00000098383 | 38351095 | 38351495 | 2.11 | -2779 | upstream |
| ENSMUSG00000027597 | 155073673 | 155074173 | 2.11 | 824 | inside |
| ENSMUSG00000061809 | 24123489 | 24124039 | 2.11 | -26894 | intergenic |
| ENSMUSG00000048001 | 154960889 | 154961289 | 2.11 | -34 | overlap |
| ENSMUSG00000028919 | 141247827 | 141248277 | 2.11 | 8328 | inside |
| ENSMUSG00000042380 | 127243736 | 127244136 | 2.11 | -48 | overlap |


| ENSMUSG00000052135 | 120286457 | 120287157 | 2.11 | 892 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000029231 | 75152732 | 75153332 | 2.11 | 441 | inside |
| ENSMUSG00000029135 | 32136270 | 32136670 | 2.11 | -202 | overlap |
| ENSMUSG00000070473 | 134986137 | 134986887 | 2.11 | -77 | overlap |
| ENSMUSG00000015053 | 88198189 | 88198589 | 2.11 | 4298 | inside |
| ENSMUSG00000068303 | 85154610 | 85155310 | 2.11 | -395 | overlap |
| ENSMUSG00000030002 | 85961460 | 85961960 | 2.11 | 207 | overlap |
| ENSMUSG00000003099 | 17027624 | 17028024 | 2.11 | 300 | overlap |
| ENSMUSG00000045969 | 11555008 | 11555408 | 2.11 | -1058 | upstream |
| ENSMUSG00000087761 | 77906623 | 77907873 | 2.11 | 99088 | intergenic |
| ENSMUSG00000050912 | 7764185 | 7764585 | 2.11 | 144 | inside |
| ENSMUSG00000031963 | 23222191 | 23222591 | 2.11 | -885 | upstream |
| ENSMUSG00000032340 | 59036778 | 59037178 | 2.11 | -337 | upstream |
| ENSMUSG00000006005 | 150368597 | 150368997 | 2.10 | -24241 | intergenic |
| ENSMUSG00000097316 | 192136562 | 192137012 | 2.10 | -336 | overlap |
| ENSMUSG00000033007 | 75461064 | 75461814 | 2.10 | 10628 | inside |
| ENSMUSG00000089866 | 105026211 | 105027411 | 2.10 | -113878 | intergenic |
| ENSMUSG00000097487 | 101425165 | 101425565 | 2.10 | 168 | overlap |
| ENSMUSG00000001444 | 97114974 | 97115574 | 2.10 | 357 | overlap |
| ENSMUSG00000034271 | 85598548 | 85599098 | 2.10 | -557 | upstream |
| ENSMUSG00000021264 | 108757917 | 108758867 | 2.10 | -35056 | intergenic |
| ENSMUSG00000051166 | 98900535 | 98901035 | 2.10 | 949 | inside |
| ENSMUSG00000042622 | 79357139 | 79357639 | 2.10 | 9598 | inside |
| ENSMUSG00000035828 | 88863189 | 88863639 | 2.10 | 1003 | inside |
| ENSMUSG00000000532 | 101174188 | 101174638 | 2.10 | 84 | inside |
| ENSMUSG00000025076 | 56397120 | 56397520 | 2.10 | -9 | overlap |
| ENSMUSG00000043531 | 50677725 | 50678575 | 2.10 | 921 | inside |
| ENSMUSG00000059326 | 61225363 | 61225763 | 2.10 | 3055 | inside |
| ENSMUSG00000085322 | 168765785 | 168766285 | 2.10 | -358 | overlap |
| ENSMUSG00000076312 | 155628953 | 155629803 | 2.10 | 6073 | intergenic |
| ENSMUSG00000055612 | 72476364 | 72476814 | 2.10 | 205 | inside |
| ENSMUSG00000027217 | 93333878 | 93334378 | 2.10 | 627 | inside |
| ENSMUSG00000062319 | 6996347 | 6997597 | 2.10 | -44667 | intergenic |
| ENSMUSG00000053819 | 126596544 | 126596944 | 2.10 | 242 | inside |
| ENSMUSG00000099169 | 140304904 | 140305504 | 2.10 | 276655 | intergenic |
| ENSMUSG00000070687 | 136424557 | 136424957 | 2.10 | 1033 | inside |
| ENSMUSG00000029326 | 100039782 | 100040182 | 2.10 | -212 | overlap |
| ENSMUSG00000029504 | 110653516 | 110653966 | 2.10 | 65 | inside |
| ENSMUSG00000089415 | 77994776 | 77995326 | 2.10 | -324930 | intergenic |
| ENSMUSG00000028995 | 24029861 | 24030261 | 2.10 | 829 | inside |
| ENSMUSG00000052751 | 48596394 | 48597594 | 2.10 | 2511 | inside |
| ENSMUSG00000032652 | 134830073 | 134830523 | 2.10 | -81 | overlap |
| ENSMUSG00000043131 | 83325739 | 83326189 | 2.10 | -277 | overlap |
| ENSMUSG00000047710 | 13869258 | 13869758 | 2.10 | -383 | overlap |
| ENSMUSG00000042195 | 53771628 | 53772128 | 2.10 | 93 | inside |
| ENSMUSG00000048752 | 110858081 | 110858481 | 2.10 | 114 | inside |
| ENSMUSG00000048000 | 87327364 | 87327864 | 2.09 | 366 | inside |
| ENSMUSG00000026234 | 86358308 | 86358808 | 2.09 | 1147 | inside |
| ENSMUSG00000025938 | 12992046 | 12992496 | 2.09 | -869 | upstream |
| ENSMUSG00000065096 | 12274860 | 12275960 | 2.09 | -88550 | intergenic |
| ENSMUSG00000000686 | 77515080 | 77515530 | 2.09 | -41 | overlap |


| ENSMUSG00000085609 | 75172177 | 75172977 | 2.09 | -383 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000017376 | 78697027 | 78697477 | 2.09 | 346 | overlap |
| ENSMUSG00000017631 | 76623080 | 76623480 | 2.09 | -766 | upstream |
| ENSMUSG00000048616 | 89301968 | 89302418 | 2.09 | 591 | inside |
| ENSMUSG00000021051 | 75595111 | 75595561 | 2.09 | 1089 | inside |
| ENSMUSG00000077473 | 4982260 | 4982810 | 2.09 | 207674 | intergenic |
| ENSMUSG00000021943 | 33923476 | 33923876 | 2.09 | -111 | overlap |
| ENSMUSG00000022552 | 76350423 | 76351623 | 2.09 | 688 | overlap |
| ENSMUSG00000022964 | 91597019 | 91597419 | 2.09 | 781 | inside |
| ENSMUSG00000024664 | 10041490 | 10042090 | 2.09 | -58 | overlap |
| ENSMUSG00000085596 | 167565476 | 167565876 | 2.09 | 12162 | inside |
| ENSMUSG00000068735 | 93187064 | 93187564 | 2.09 | -484 | overlap |
| ENSMUSG00000026959 | 25318689 | 25319089 | 2.09 | 498 | inside |
| ENSMUSG00000027016 | 77816928 | 77817328 | 2.09 | 2711 | inside |
| ENSMUSG00000078578 | 135437953 | 135438653 | 2.09 | -344 | overlap |
| ENSMUSG00000087440 | 51275945 | 51276595 | 2.09 | -1626 | upstream |
| ENSMUSG00000027985 | 131110100 | 131111250 | 2.09 | -371 | overlap |
| ENSMUSG00000053965 | 122729221 | 122729921 | 2.09 | 63 | inside |
| ENSMUSG00000027715 | 36571145 | 36571845 | 2.09 | 1005 | inside |
| ENSMUSG00000037325 | 36613090 | 36613540 | 2.09 | 387 | overlap |
| ENSMUSG00000028840 | 134249397 | 134249997 | 2.09 | -3805 | upstream |
| ENSMUSG00000025858 | 139252439 | 139252939 | 2.09 | 115 | inside |
| ENSMUSG00000029152 | 73292562 | 73293012 | 2.09 | -232 | overlap |
| ENSMUSG00000023353 | 24485175 | 24485575 | 2.09 | 32998 | inside |
| ENSMUSG00000002633 | 28593395 | 28594245 | 2.09 | -126294 | intergenic |
| ENSMUSG00000077474 | 83715318 | 83715768 | 2.09 | 8271 | intergenic |
| ENSMUSG00000029575 | 114444048 | 114444498 | 2.09 | 11 | overlap |
| ENSMUSG00000092928 | 48024611 | 48025011 | 2.09 | -24092 | intergenic |
| ENSMUSG00000076327 | 128034875 | 128035725 | 2.09 | 60151 | intergenic |
| ENSMUSG00000030527 | 80688027 | 80688677 | 2.09 | 850 | inside |
| ENSMUSG00000082281 | 103966768 | 103967218 | 2.09 | 10339 | intergenic |
| ENSMUSG00000087530 | 128010009 | 128010459 | 2.09 | -4819 | upstream |
| ENSMUSG00000034330 | 117498430 | 117498880 | 2.09 | 139 | inside |
| ENSMUSG00000019464 | 83667842 | 83668242 | 2.09 | 1009 | inside |
| ENSMUSG00000031609 | 57486992 | 57487392 | 2.09 | 868 | inside |
| ENSMUSG00000036466 | 64385979 | 64386379 | 2.09 | 353 | inside |
| ENSMUSG00000086158 | 72985328 | 72985728 | 2.09 | 54 | overlap |
| ENSMUSG00000026176 | 74391945 | 74392345 | 2.08 | 436 | inside |
| ENSMUSG00000038473 | 170589852 | 170590252 | 2.08 | 9 | overlap |
| ENSMUSG00000015961 | 177795558 | 177796408 | 2.08 | 953 | inside |
| ENSMUSG00000035873 | 108333173 | 108333573 | 2.08 | 984 | inside |
| ENSMUSG00000034994 | 81176756 | 81177256 | 2.08 | 125 | inside |
| ENSMUSG00000019979 | 91082263 | 91082763 | 2.08 | 507 | inside |
| ENSMUSG00000045912 | 79613865 | 79614265 | 2.08 | 160 | overlap |
| ENSMUSG00000090266 | 116842068 | 116843018 | 2.08 | -1210 | upstream |
| ENSMUSG00000044950 | 43682427 | 43682827 | 2.08 | 429 | inside |
| ENSMUSG00000085516 | 24164050 | 24164500 | 2.08 | 8774 | inside |
| ENSMUSG00000039976 | 119227965 | 119228365 | 2.08 | 534 | inside |
| ENSMUSG00000097061 | 69196776 | 69197376 | 2.08 | -435 | overlap |
| ENSMUSG00000006356 | 113139987 | 113140387 | 2.08 | -249 | overlap |
| ENSMUSG00000048251 | 107916795 | 107917295 | 2.08 | 86807 | inside |


| ENSMUSG00000091105 | 21898703 | 21899303 | 2.08 | -121996 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000021118 | 78906761 | 78907161 | 2.08 | 203 | overlap |
| ENSMUSG00000091387 | 96924401 | 96924851 | 2.08 | -288 | overlap |
| ENSMUSG00000021391 | 49652421 | 49652871 | 2.08 | 325 | overlap |
| ENSMUSG00000021676 | 95891735 | 95892435 | 2.08 | 187 | overlap |
| ENSMUSG00000025544 | 122107177 | 122107577 | 2.08 | 139 | inside |
| ENSMUSG00000021838 | 46881905 | 46882955 | 2.08 | -949 | overlap |
| ENSMUSG00000022012 | 77156456 | 77156856 | 2.08 | -307 | overlap |
| ENSMUSG00000065760 | 49600055 | 49600455 | 2.08 | 61860 | intergenic |
| ENSMUSG00000050761 | 18621117 | 18621717 | 2.08 | 1286 | inside |
| ENSMUSG00000023143 | 5203502 | 5204052 | 2.08 | 510 | overlap |
| ENSMUSG00000071054 | 56584773 | 56585173 | 2.08 | -52 | overlap |
| ENSMUSG00000096361 | 47410008 | 47410458 | 2.08 | -355 | overlap |
| ENSMUSG00000040140 | 43629844 | 43630294 | 2.08 | 455 | inside |
| ENSMUSG00000024608 | 60748677 | 60749077 | 2.08 | -25833 | intergenic |
| ENSMUSG00000024565 | 80986048 | 80986498 | 2.08 | 530 | inside |
| ENSMUSG00000024487 | 40219159 | 40219559 | 2.08 | 240 | overlap |
| ENSMUSG00000024378 | 33330398 | 33330998 | 2.08 | -116536 | intergenic |
| ENSMUSG00000067199 | 41829640 | 41830040 | 2.08 | -528 | upstream |
| ENSMUSG00000027678 | 165992971 | 165993421 | 2.08 | 335 | inside |
| ENSMUSG00000042448 | 74763202 | 74763652 | 2.08 | 222 | inside |
| ENSMUSG00000035403 | 37786701 | 37787101 | 2.08 | 10452 | inside |
| ENSMUSG00000002100 | 91115269 | 91115719 | 2.08 | -2875 | upstream |
| ENSMUSG00000059173 | 79936936 | 79937386 | 2.08 | 192522 | inside |
| ENSMUSG00000074912 | 119337835 | 119338335 | 2.08 | -11641 | intergenic |
| ENSMUSG00000044320 | 30801215 | 30801765 | 2.08 | 2446 | inside |
| ENSMUSG00000082678 | 112979593 | 112979993 | 2.08 | 220 | overlap |
| ENSMUSG00000066224 | 41730760 | 41731360 | 2.08 | 382 | overlap |
| ENSMUSG00000029334 | 99037160 | 99037660 | 2.08 | 191 | overlap |
| ENSMUSG00000039474 | 36988768 | 36989168 | 2.08 | 214 | overlap |
| ENSMUSG00000028995 | 24030411 | 24030811 | 2.08 | 279 | overlap |
| ENSMUSG00000084934 | 141383175 | 141383925 | 2.08 | -2249 | upstream |
| ENSMUSG00000079511 | 83102089 | 83102489 | 2.08 | 472 | inside |
| ENSMUSG00000063870 | 125095975 | 125096525 | 2.08 | -6 | overlap |
| ENSMUSG00000002222 | 71440339 | 71441039 | 2.08 | 298 | overlap |
| ENSMUSG00000049583 | 87584213 | 87584713 | 2.08 | -18331 | intergenic |
| ENSMUSG00000030731 | 44352027 | 44352477 | 2.08 | -32075 | intergenic |
| ENSMUSG00000099103 | 143445141 | 143445591 | 2.08 | -2464 | upstream |
| ENSMUSG00000048617 | 84944990 | 84945990 | 2.08 | -2001 | upstream |
| ENSMUSG00000074136 | 95806789 | 95807539 | 2.08 | -41 | overlap |
| ENSMUSG00000096943 | 83896990 | 83897390 | 2.08 | 2511 | inside |
| ENSMUSG00000084128 | 106136358 | 106136808 | 2.08 | 616 | inside |
| ENSMUSG00000032591 | 108079005 | 108079505 | 2.08 | -1431 | upstream |
| ENSMUSG00000026308 | 91344850 | 91345350 | 2.07 | -6166 | intergenic |
| ENSMUSG00000067028 | 99593862 | 99594912 | 2.07 | -178903 | intergenic |
| ENSMUSG00000026028 | 58973094 | 58973544 | 2.07 | 336 | overlap |
| ENSMUSG00000025982 | 55027389 | 55027889 | 2.07 | 89 | overlap |
| ENSMUSG00000074785 | 94943633 | 94944033 | 2.07 | 945 | inside |
| ENSMUSG00000048756 | 42276473 | 42276873 | 2.07 | 282 | overlap |
| ENSMUSG00000042650 | 60536877 | 60537327 | 2.07 | 496 | inside |
| ENSMUSG00000005417 | 59662030 | 59662430 | 2.07 | 725 | inside |


| ENSMUSG00000049336 | 37236177 | 37236577 | 2.07 | -213 | upstream |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000020598 | 44328790 | 44329490 | 2.07 | -95 | overlap |
| ENSMUSG00000063632 | 27342279 | 27343229 | 2.07 | 430 | overlap |
| ENSMUSG00000050671 | 87299913 | 87300313 | 2.07 | -208 | upstream |
| ENSMUSG00000037169 | 12941321 | 12941821 | 2.07 | 593 | inside |
| ENSMUSG00000056770 | 108177495 | 108178795 | 2.07 | 1819 | inside |
| ENSMUSG00000021318 | 15463923 | 15464373 | 2.07 | 688 | inside |
| ENSMUSG00000034928 | 54693955 | 54694455 | 2.07 | -48 | upstream |
| ENSMUSG00000038009 | 99093665 | 99094115 | 2.07 | 495 | inside |
| ENSMUSG00000055745 | 84557116 | 84557566 | 2.07 | 707 | inside |
| ENSMUSG00000001076 | 99088224 | 99088674 | 2.07 | -407 | upstream |
| ENSMUSG00000058600 | 34442991 | 34443491 | 2.07 | 649 | inside |
| ENSMUSG00000041935 | 3995253 | 3995653 | 2.07 | 499 | inside |
| ENSMUSG00000071636 | 17208379 | 17208779 | 2.07 | 244 | inside |
| ENSMUSG00000047953 | 30308317 | 30308767 | 2.07 | 2462 | inside |
| ENSMUSG00000023067 | 29094398 | 29094798 | 2.07 | 3419 | inside |
| ENSMUSG00000037089 | 45563247 | 45564047 | 2.07 | -717 | overlap |
| ENSMUSG00000024242 | 80727217 | 80727617 | 2.07 | 876 | inside |
| ENSMUSG00000024335 | 34118212 | 34118912 | 2.07 | 4422 | inside |
| ENSMUSG00000047466 | 77712892 | 77713342 | 2.07 | 1118 | inside |
| ENSMUSG00000054874 | 5688066 | 5688466 | 2.07 | 842 | inside |
| ENSMUSG00000077351 | 43113751 | 43114501 | 2.07 | 304716 | intergenic |
| ENSMUSG00000027015 | 71117764 | 71119114 | 2.07 | -159 | overlap |
| ENSMUSG00000005803 | 122764985 | 122765385 | 2.07 | -252 | overlap |
| ENSMUSG00000081878 | 71786808 | 71787408 | 2.07 | -24041 | intergenic |
| ENSMUSG00000084339 | 19599812 | 19601162 | 2.07 | 15021 | intergenic |
| ENSMUSG00000074517 | 83127795 | 83128195 | 2.07 | 1451 | inside |
| ENSMUSG00000070990 | 46343376 | 46344026 | 2.07 | -233 | overlap |
| ENSMUSG00000028409 | 40757559 | 40758009 | 2.07 | 364 | overlap |
| ENSMUSG00000039813 | 46649550 | 46650150 | 2.07 | 659 | inside |
| ENSMUSG00000056596 | 133498257 | 133498707 | 2.07 | 293 | overlap |
| ENSMUSG00000059991 | 144545366 | 144545816 | 2.07 | -521 | upstream |
| ENSMUSG00000018001 | 143622574 | 143623074 | 2.07 | 127 | inside |
| ENSMUSG00000075551 | 145681225 | 145681725 | 2.07 | 38911 | intergenic |
| ENSMUSG00000029147 | 31219712 | 31220212 | 2.07 | 833 | inside |
| ENSMUSG00000062960 | 75977852 | 75978352 | 2.07 | 606 | inside |
| ENSMUSG00000029173 | 52669504 | 52669904 | 2.07 | 225 | overlap |
| ENSMUSG00000073144 | 39118444 | 39118944 | 2.07 | 36 | inside |
| ENSMUSG00000049661 | 45361603 | 45362003 | 2.07 | -4560 | upstream |
| ENSMUSG00000055652 | 75995927 | 75996377 | 2.07 | 147589 | intergenic |
| ENSMUSG00000042797 | 97737522 | 97738072 | 2.07 | 725 | inside |
| ENSMUSG00000046792 | 6155821 | 6156271 | 2.07 | 150 | overlap |
| ENSMUSG00000066697 | 71328546 | 71329796 | 2.07 | 4375 | downstream |
| ENSMUSG00000052837 | 84977905 | 84978305 | 2.07 | 843 | inside |
| ENSMUSG00000031667 | 91133609 | 91134009 | 2.07 | 856 | inside |
| ENSMUSG00000053399 | 113848330 | 113848830 | 2.07 | 285 | overlap |
| ENSMUSG00000004319 | 60982807 | 60983407 | 2.07 | 493 | overlap |
| ENSMUSG00000066687 | 48835031 | 48835481 | 2.07 | 914 | inside |
| ENSMUSG00000052331 | 54925444 | 54925944 | 2.06 | 943 | inside |
| ENSMUSG00000018417 | 51914294 | 51915244 | 2.06 | 1777 | inside |
| ENSMUSG00000020009 | 19591852 | 19592302 | 2.06 | -97 | overlap |


| ENSMUSG00000019916 | 59323503 | 59324003 | 2.06 | 207 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000071369 | 19933595 | 19934145 | 2.06 | -877 | upstream |
| ENSMUSG00000019992 | 20347502 | 20348002 | 2.06 | -317 | overlap |
| ENSMUSG00000020225 | 120201195 | 120201995 | 2.06 | -395 | overlap |
| ENSMUSG00000023068 | 52417854 | 52418354 | 2.06 | 307 | inside |
| ENSMUSG00000056316 | 29147423 | 29147873 | 2.06 | -3229 | upstream |
| ENSMUSG00000020261 | 55204227 | 55204727 | 2.06 | -123 | overlap |
| ENSMUSG00000020883 | 98149715 | 98150365 | 2.06 | 688 | inside |
| ENSMUSG00000021051 | 75596411 | 75596861 | 2.06 | -211 | upstream |
| ENSMUSG00000034928 | 54687070 | 54687470 | 2.06 | 6837 | inside |
| ENSMUSG00000041078 | 34820427 | 34820827 | 2.06 | 602 | inside |
| ENSMUSG00000041408 | 34673944 | 34674594 | 2.06 | 16 | inside |
| ENSMUSG00000046204 | 66910543 | 66911093 | 2.06 | -627 | upstream |
| ENSMUSG00000068697 | 20660908 | 20661458 | 2.06 | -4368 | upstream |
| ENSMUSG00000022568 | 76074171 | 76074571 | 2.06 | -4387 | upstream |
| ENSMUSG00000047565 | 20551946 | 20552546 | 2.06 | 114804 | intergenic |
| ENSMUSG00000094605 | 43142810 | 43143260 | 2.06 | 16577 | intergenic |
| ENSMUSG00000045038 | 86166520 | 86166920 | 2.06 | -1265 | upstream |
| ENSMUSG00000042644 | 27056765 | 27057465 | 2.06 | -539 | overlap |
| ENSMUSG00000041168 | 56626737 | 56627137 | 2.06 | 166 | overlap |
| ENSMUSG00000054723 | 56716287 | 56716687 | 2.06 | 1412 | inside |
| ENSMUSG00000052397 | 6783207 | 6783607 | 2.06 | -423 | upstream |
| ENSMUSG00000063239 | 27512001 | 27512401 | 2.06 | 1340 | inside |
| ENSMUSG00000034484 | 53176427 | 53176877 | 2.06 | 62 | inside |
| ENSMUSG00000025885 | 74442727 | 74443177 | 2.06 | 1791 | inside |
| ENSMUSG00000024927 | 5637015 | 5637515 | 2.06 | -468 | overlap |
| ENSMUSG00000027624 | 156421085 | 156422035 | 2.06 | 176 | inside |
| ENSMUSG00000075304 | 70476402 | 70476802 | 2.06 | 1479 | inside |
| ENSMUSG00000027035 | 68861964 | 68862664 | 2.06 | 523 | inside |
| ENSMUSG00000053615 | 125505835 | 125506235 | 2.06 | 746 | inside |
| ENSMUSG00000027593 | 154790749 | 154791249 | 2.06 | -347 | overlap |
| ENSMUSG00000027326 | 119047203 | 119047653 | 2.06 | 84 | inside |
| ENSMUSG00000089458 | 148167785 | 148168785 | 2.06 | 16635 | intergenic |
| ENSMUSG00000027544 | 168570693 | 168571343 | 2.06 | 30964 | inside |
| ENSMUSG00000075012 | 102451014 | 102451414 | 2.06 | 778 | inside |
| ENSMUSG00000074796 | 130697135 | 130697635 | 2.06 | 384 | overlap |
| ENSMUSG00000027777 | 68166030 | 68166930 | 2.06 | 101228 | inside |
| ENSMUSG00000040389 | 108590907 | 108591357 | 2.06 | -372 | overlap |
| ENSMUSG00000084163 | 27534515 | 27535215 | 2.06 | -8663 | intergenic |
| ENSMUSG00000052135 | 120268707 | 120269157 | 2.06 | 18642 | inside |
| ENSMUSG00000028909 | 131836665 | 131837065 | 2.06 | 1623 | inside |
| ENSMUSG00000040928 | 129189478 | 129190078 | 2.06 | 249 | overlap |
| ENSMUSG00000039137 | 63494698 | 63495098 | 2.06 | 1293 | inside |
| ENSMUSG00000049907 | 74195132 | 74195982 | 2.06 | -156 | overlap |
| ENSMUSG00000054252 | 33721908 | 33722358 | 2.06 | 184 | inside |
| ENSMUSG00000088751 | 57108318 | 57108718 | 2.06 | -2506 | upstream |
| ENSMUSG00000029192 | 36581294 | 36581744 | 2.06 | 11982 | inside |
| ENSMUSG00000030094 | 91515110 | 91515860 | 2.06 | 774 | inside |
| ENSMUSG00000071341 | 85512497 | 85513097 | 2.06 | 1092 | inside |
| ENSMUSG00000055407 | 99268366 | 99268816 | 2.06 | 919 | inside |
| ENSMUSG00000038244 | 112225462 | 112225862 | 2.06 | -394 | overlap |


| ENSMUSG00000025586 | 81453460 | 81454010 | 2.06 | 2005 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000030849 | 133121930 | 133122330 | 2.06 | 1420 | inside |
| ENSMUSG00000096938 | 11006580 | 11007180 | 2.06 | -1270 | upstream |
| ENSMUSG00000019261 | 70913905 | 70914405 | 2.06 | 7931 | inside |
| ENSMUSG00000097702 | 120598272 | 120598722 | 2.06 | -280 | overlap |
| ENSMUSG00000098322 | 15028008 | 15028458 | 2.06 | -9101 | intergenic |
| ENSMUSG00000025237 | 59617278 | 59617878 | 2.06 | -6 | overlap |
| ENSMUSG00000096786 | 109366971 | 109367521 | 2.06 | -8636 | intergenic |
| ENSMUSG00000045087 | 21244241 | 21244691 | 2.06 | 4202 | inside |
| ENSMUSG00000032396 | 64340428 | 64340978 | 2.06 | 860 | inside |
| ENSMUSG00000025945 | 65979370 | 65979970 | 2.05 | 52848 | intergenic |
| ENSMUSG00000097347 | 180777438 | 180777888 | 2.05 | -25130 | intergenic |
| ENSMUSG00000090031 | 6214346 | 6214746 | 2.05 | 244 | overlap |
| ENSMUSG00000026620 | 184956290 | 184956790 | 2.05 | 43259 | inside |
| ENSMUSG00000020037 | 84754806 | 84755206 | 2.05 | -1256 | upstream |
| ENSMUSG00000035529 | 85916706 | 85917106 | 2.05 | 239 | overlap |
| ENSMUSG00000065395 | 79711577 | 79712027 | 2.05 | -392 | overlap |
| ENSMUSG00000046215 | 103649124 | 103649524 | 2.05 | -385 | overlap |
| ENSMUSG00000005951 | 73199398 | 73199798 | 2.05 | -62 | overlap |
| ENSMUSG00000018217 | 63132938 | 63133388 | 2.05 | 3956 | inside |
| ENSMUSG00000046719 | 95513468 | 95513868 | 2.05 | 1102 | inside |
| ENSMUSG00000020902 | 68399836 | 68400336 | 2.05 | 987 | inside |
| ENSMUSG00000021140 | 81860657 | 81861157 | 2.05 | 627 | inside |
| ENSMUSG00000056553 | 116757082 | 116757982 | 2.05 | 271362 | inside |
| ENSMUSG00000048982 | 75379040 | 75379540 | 2.05 | 37741 | intergenic |
| ENSMUSG00000095777 | 78329195 | 78329795 | 2.05 | -31902 | intergenic |
| ENSMUSG00000016477 | 29985221 | 29985671 | 2.05 | 447 | overlap |
| ENSMUSG00000022176 | 54479226 | 54479626 | 2.05 | 3126 | inside |
| ENSMUSG00000090534 | 118235258 | 118235658 | 2.05 | -974 | upstream |
| ENSMUSG00000021767 | 21500141 | 21501041 | 2.05 | 18707 | inside |
| ENSMUSG00000040123 | 56811426 | 56811826 | 2.05 | 290 | overlap |
| ENSMUSG00000044819 | 120137640 | 120138140 | 2.05 | -95205 | intergenic |
| ENSMUSG00000034731 | 78731806 | 78732206 | 2.05 | -6717 | intergenic |
| ENSMUSG00000050310 | 6708119 | 6708519 | 2.05 | -262 | overlap |
| ENSMUSG00000036800 | 71169473 | 71170773 | 2.05 | 558365 | intergenic |
| ENSMUSG00000022840 | 35156174 | 35156574 | 2.05 | 1297 | inside |
| ENSMUSG00000071475 | 83499757 | 83500207 | 2.05 | -128024 | intergenic |
| ENSMUSG00000024188 | 26252911 | 26253461 | 2.05 | 1 | inside |
| ENSMUSG00000001228 | 56304144 | 56304594 | 2.05 | 823 | inside |
| ENSMUSG00000040356 | 34849851 | 34850401 | 2.05 | 359 | overlap |
| ENSMUSG00000027381 | 128125785 | 128126185 | 2.05 | -253 | overlap |
| ENSMUSG00000086436 | 80638819 | 80639269 | 2.05 | -9251 | intergenic |
| ENSMUSG00000013465 | 25210939 | 25211439 | 2.05 | 550 | inside |
| ENSMUSG00000034075 | 84714708 | 84715258 | 2.05 | 472 | overlap |
| ENSMUSG00000026932 | 26091401 | 26092051 | 2.05 | 31819 | inside |
| ENSMUSG00000036591 | 20968594 | 20969444 | 2.05 | 287 | overlap |
| ENSMUSG00000092345 | 59003352 | 59003802 | 2.05 | 142 | inside |
| ENSMUSG00000028341 | 48045220 | 48045620 | 2.05 | 67 | inside |
| ENSMUSG00000073684 | 155249683 | 155250083 | 2.05 | -119 | overlap |
| ENSMUSG00000070803 | 120667093 | 120667493 | 2.05 | 530 | inside |
| ENSMUSG00000028456 | 43058840 | 43059290 | 2.05 | -113 | overlap |


| ENSMUSG00000008305 | 72199608 | 72200158 | 2.05 | 1311 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000014030 | 44703176 | 44704076 | 2.05 | 8270 | inside |
| ENSMUSG00000055235 | 24730033 | 24730533 | 2.05 | 694 | inside |
| ENSMUSG00000000568 | 99978610 | 99979460 | 2.05 | 328 | overlap |
| ENSMUSG00000056755 | 110645539 | 110646039 | 2.05 | -42 | overlap |
| ENSMUSG00000053297 | 48619842 | 48620492 | 2.05 | -6922 | intergenic |
| ENSMUSG00000029833 | 37871183 | 37871683 | 2.05 | 372 | inside |
| ENSMUSG00000032667 | 5297853 | 5298303 | 2.05 | 602 | inside |
| ENSMUSG00000063757 | 5030604 | 5031054 | 2.05 | -3514 | upstream |
| ENSMUSG00000030541 | 80115137 | 80115587 | 2.05 | 255 | overlap |
| ENSMUSG00000048583 | 142663718 | 142664168 | 2.05 | 3098 | inside |
| ENSMUSG00000002396 | 71371473 | 71371873 | 2.05 | 175 | inside |
| ENSMUSG00000031737 | 92359190 | 92360540 | 2.05 | 1394 | inside |
| ENSMUSG00000056267 | 99243266 | 99243666 | 2.05 | -176 | overlap |
| ENSMUSG00000049742 | 7836585 | 7837135 | 2.05 | 479 | includeFeature |
| ENSMUSG00000059237 | 66805828 | 66806228 | 2.05 | 110 | overlap |
| ENSMUSG00000026638 | 193152945 | 193153495 | 2.04 | -167 | overlap |
| ENSMUSG00000026113 | 37299584 | 37300184 | 2.04 | -281 | overlap |
| ENSMUSG00000026277 | 93634867 | 93635267 | 2.04 | 855 | inside |
| ENSMUSG00000026482 | 152765847 | 152766497 | 2.04 | 504 | overlap |
| ENSMUSG00000026121 | 36557499 | 36558049 | 2.04 | 850 | inside |
| ENSMUSG00000014329 | 71159145 | 71159545 | 2.04 | 555 | inside |
| ENSMUSG00000041164 | 6389629 | 6390029 | 2.04 | 555 | inside |
| ENSMUSG00000052915 | 98795115 | 98795515 | 2.04 | -401 | upstream |
| ENSMUSG00000038351 | 74896927 | 74897527 | 2.04 | 133 | overlap |
| ENSMUSG00000003934 | 69559736 | 69560186 | 2.04 | 469 | inside |
| ENSMUSG00000020868 | 94670465 | 94670865 | 2.04 | 7050 | inside |
| ENSMUSG00000018537 | 97700115 | 97700515 | 2.04 | 382 | overlap |
| ENSMUSG00000045440 | 55600511 | 55600911 | 2.04 | 1594 | inside |
| ENSMUSG00000021264 | 108793888 | 108794288 | 2.04 | 915 | inside |
| ENSMUSG00000020973 | 69197826 | 69198326 | 2.04 | 603 | inside |
| ENSMUSG00000064138 | 77857495 | 77858245 | 2.04 | 148805 | inside |
| ENSMUSG00000021314 | 18948237 | 18948687 | 2.04 | -134 | overlap |
| ENSMUSG00000015396 | 43784671 | 43785071 | 2.04 | -436 | upstream |
| ENSMUSG00000035248 | 59822608 | 59823008 | 2.04 | 539 | inside |
| ENSMUSG00000016477 | 29983771 | 29984171 | 2.04 | 1897 | inside |
| ENSMUSG00000021506 | 55825929 | 55826329 | 2.04 | 10263 | inside |
| ENSMUSG00000025872 | 54468704 | 54469304 | 2.04 | 145 | overlap |
| ENSMUSG00000033885 | 8097599 | 8098199 | 2.04 | -614 | upstream |
| ENSMUSG00000097136 | 25607094 | 25607944 | 2.04 | -52725 | intergenic |
| ENSMUSG00000058655 | 102073870 | 102074370 | 2.04 | 97 | inside |
| ENSMUSG00000022427 | 79669373 | 79670523 | 2.04 | -1488 | upstream |
| ENSMUSG00000036678 | 102355465 | 102355865 | 2.04 | -4706 | upstream |
| ENSMUSG00000097536 | 96285170 | 96285570 | 2.04 | -675 | upstream |
| ENSMUSG00000022483 | 98004415 | 98004915 | 2.04 | 280 | overlap |
| ENSMUSG00000022797 | 32608709 | 32609159 | 2.04 | -211 | overlap |
| ENSMUSG00000022503 | 10411644 | 10412144 | 2.04 | -304 | overlap |
| ENSMUSG00000024155 | 24803915 | 24804815 | 2.04 | -467 | overlap |
| ENSMUSG00000095687 | 8147528 | 8148028 | 2.04 | 569 | inside |
| ENSMUSG00000050138 | 87796233 | 87797083 | 2.04 | 1761 | inside |
| ENSMUSG00000088493 | 37732833 | 37733333 | 2.04 | 26854 | intergenic |


| ENSMUSG00000069378 | 53463548 | 53464098 | 2.04 | -998 | upstream |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000052928 | 75696433 | 75696833 | 2.04 | 1263 | inside |
| ENSMUSG00000024759 | 7494063 | 7494513 | 2.04 | 23 | inside |
| ENSMUSG00000033768 | 6428466 | 6428916 | 2.04 | 9735 | inside |
| ENSMUSG00000025010 | 40842325 | 40842725 | 2.04 | 11046 | inside |
| ENSMUSG00000040565 | 36925613 | 36926163 | 2.04 | -466 | overlap |
| ENSMUSG00000053080 | 7417326 | 7418276 | 2.04 | -299 | overlap |
| ENSMUSG00000024869 | 4000210 | 4000760 | 2.04 | -370 | overlap |
| ENSMUSG00000024833 | 5963739 | 5964289 | 2.04 | 467 | overlap |
| ENSMUSG00000059326 | 61226663 | 61227413 | 2.04 | 1755 | inside |
| ENSMUSG00000082971 | 55631010 | 55631410 | 2.04 | -144538 | intergenic |
| ENSMUSG00000053166 | 165233376 | 165234326 | 2.04 | 1477 | inside |
| ENSMUSG00000026888 | 65022082 | 65022582 | 2.04 | 2905 | inside |
| ENSMUSG00000029419 | 25577889 | 25578289 | 2.04 | 2210 | inside |
| ENSMUSG00000027765 | 61002617 | 61003367 | 2.04 | -178 | overlap |
| ENSMUSG00000077441 | 147780003 | 147780553 | 2.04 | 574215 | intergenic |
| ENSMUSG00000028654 | 122995481 | 122996031 | 2.04 | -171 | overlap |
| ENSMUSG00000073700 | 152008713 | 152009113 | 2.04 | -90 | overlap |
| ENSMUSG00000028341 | 48043276 | 48043676 | 2.04 | -1877 | upstream |
| ENSMUSG00000028736 | 139821577 | 139822027 | 2.04 | 11951 | inside |
| ENSMUSG00000071019 | 4048633 | 4049433 | 2.04 | 28889 | intergenic |
| ENSMUSG00000041530 | 126468498 | 126468948 | 2.04 | 85 | overlap |
| ENSMUSG00000029287 | 107289066 | 107289466 | 2.04 | 563 | inside |
| ENSMUSG00000002297 | 8422143 | 8422593 | 2.04 | 573 | inside |
| ENSMUSG00000048450 | 37823918 | 37824368 | 2.04 | 665 | inside |
| ENSMUSG00000063935 | 72580562 | 72581062 | 2.04 | 522 | inside |
| ENSMUSG00000037235 | 34187428 | 34187878 | 2.04 | 292 | overlap |
| ENSMUSG00000059518 | 136987373 | 136987973 | 2.04 | 648 | inside |
| ENSMUSG00000015053 | 88189749 | 88190249 | 2.04 | -4142 | upstream |
| ENSMUSG00000049093 | 67508047 | 67508597 | 2.04 | -16192 | intergenic |
| ENSMUSG00000010797 | 18030043 | 18030493 | 2.04 | 542 | inside |
| ENSMUSG00000080562 | 27364344 | 27364744 | 2.04 | 1030 | downstream |
| ENSMUSG00000007946 | 101821409 | 101821809 | 2.04 | 3096 | inside |
| ENSMUSG00000046591 | 79660227 | 79660877 | 2.04 | 31 | inside |
| ENSMUSG00000002635 | 34196528 | 34196978 | 2.04 | 133 | overlap |
| ENSMUSG00000098839 | 45007103 | 45007803 | 2.04 | 5384 | downstream |
| ENSMUSG00000001472 | 123373139 | 123373539 | 2.04 | -685 | upstream |
| ENSMUSG00000049946 | 58966461 | 58967311 | 2.04 | 54706 | intergenic |
| ENSMUSG00000025809 | 128684639 | 128685739 | 2.04 | -1015 | overlap |
| ENSMUSG00000098560 | 121085008 | 121085408 | 2.04 | -302 | upstream |
| ENSMUSG00000041440 | 96118966 | 96119516 | 2.04 | -396 | overlap |
| ENSMUSG00000032403 | 63398728 | 63399128 | 2.04 | 516 | inside |
| ENSMUSG00000084701 | 96536916 | 96537816 | 2.04 | 12793 | intergenic |
| ENSMUSG00000039224 | 186967160 | 186967660 | 2.03 | -256 | overlap |
| ENSMUSG00000026502 | 178187813 | 178188213 | 2.03 | 396 | inside |
| ENSMUSG00000009907 | 106796300 | 106796700 | 2.03 | 428 | inside |
| ENSMUSG00000050069 | 174938158 | 174938958 | 2.03 | -16339 | intergenic |
| ENSMUSG00000026466 | 156036264 | 156036714 | 2.03 | 216 | overlap |
| ENSMUSG00000055493 | 11342812 | 11343362 | 2.03 | -592 | upstream |
| ENSMUSG00000038602 | 52690704 | 52691454 | 2.03 | 171 | inside |
| ENSMUSG00000048756 | 42275273 | 42275673 | 2.03 | 1482 | inside |


| ENSMUSG00000025366 | 128525295 | 128525745 | 2.03 | 564 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000020456 | 6291626 | 6292076 | 2.03 | -7 | overlap |
| ENSMUSG00000018899 | 53769536 | 53770186 | 2.03 | -478 | overlap |
| ENSMUSG00000017774 | 75654077 | 75654477 | 2.03 | 3573 | inside |
| ENSMUSG00000020907 | 67678186 | 67678886 | 2.03 | -17140 | intergenic |
| ENSMUSG00000037243 | 58307377 | 58307777 | 2.03 | 308 | inside |
| ENSMUSG00000004040 | 100939168 | 100939568 | 2.03 | 372 | overlap |
| ENSMUSG00000071234 | 84698677 | 84699077 | 2.03 | 130 | overlap |
| ENSMUSG00000021381 | 48664587 | 48665137 | 2.03 | 1589 | inside |
| ENSMUSG00000041112 | 20090197 | 20090647 | 2.03 | -310 | overlap |
| ENSMUSG00000059877 | 106632386 | 106633036 | 2.03 | -85326 | intergenic |
| ENSMUSG00000046908 | 55767843 | 55768243 | 2.03 | 1881 | inside |
| ENSMUSG00000025555 | 121035941 | 121036491 | 2.03 | 741 | inside |
| ENSMUSG00000021939 | 63122126 | 63122826 | 2.03 | -336 | overlap |
| ENSMUSG00000054423 | 12821348 | 12821948 | 2.03 | 1731 | inside |
| ENSMUSG00000047347 | 82516066 | 82516666 | 2.03 | -245 | overlap |
| ENSMUSG00000089804 | 37233850 | 37234300 | 2.03 | -541 | upstream |
| ENSMUSG00000033707 | 76722371 | 76722771 | 2.03 | -198 | upstream |
| ENSMUSG00000071637 | 15887160 | 15887610 | 2.03 | -126 | overlap |
| ENSMUSG00000046119 | 90514800 | 90515250 | 2.03 | -39364 | intergenic |
| ENSMUSG00000014074 | 32277436 | 32277886 | 2.03 | -23 | overlap |
| ENSMUSG00000024301 | 26916962 | 26917362 | 2.03 | -129 | overlap |
| ENSMUSG00000024462 | 37050704 | 37051104 | 2.03 | 4738 | inside |
| ENSMUSG00000024014 | 29490972 | 29491372 | 2.03 | 160 | inside |
| ENSMUSG00000097566 | 15374909 | 15375809 | 2.03 | -91 | overlap |
| ENSMUSG00000072082 | 24251165 | 24251665 | 2.03 | 244 | overlap |
| ENSMUSG00000093508 | 67143770 | 67144670 | 2.03 | -117535 | intergenic |
| ENSMUSG00000039615 | 25832251 | 25832801 | 2.03 | 1110 | inside |
| ENSMUSG00000055795 | 14291054 | 14292104 | 2.03 | -133814 | intergenic |
| ENSMUSG00000024304 | 16808204 | 16808904 | 2.03 | 1042 | inside |
| ENSMUSG00000025218 | 45541663 | 45542213 | 2.03 | 18868 | intergenic |
| ENSMUSG00000050530 | 3118643 | 3119143 | 2.03 | 4419 | inside |
| ENSMUSG00000062175 | 156840735 | 156841235 | 2.03 | 658 | inside |
| ENSMUSG00000068859 | 73273364 | 73273764 | 2.03 | 1439 | inside |
| ENSMUSG00000042662 | 152950812 | 152951512 | 2.03 | 886 | inside |
| ENSMUSG00000090625 | 174327820 | 174328570 | 2.03 | 18892 | downstream |
| ENSMUSG00000077761 | 168912009 | 168912559 | 2.03 | -29841 | intergenic |
| ENSMUSG00000082460 | 83547169 | 83547669 | 2.03 | -8494 | intergenic |
| ENSMUSG00000025782 | 10048062 | 10048712 | 2.03 | 534 | overlap |
| ENSMUSG00000028221 | 14864101 | 14864501 | 2.03 | 25 | inside |
| ENSMUSG00000028747 | 139074565 | 139075165 | 2.03 | 1005 | inside |
| ENSMUSG00000029154 | 73405840 | 73406240 | 2.03 | -236 | overlap |
| ENSMUSG00000039000 | 29568745 | 29569145 | 2.03 | -497 | upstream |
| ENSMUSG00000060708 | 36748418 | 36749018 | 2.03 | 261 | overlap |
| ENSMUSG00000029128 | 41707704 | 41708254 | 2.03 | 451 | overlap |
| ENSMUSG00000055923 | 76857682 | 76858182 | 2.03 | 47832 | intergenic |
| ENSMUSG00000037822 | 65537162 | 65537562 | 2.03 | 22 | overlap |
| ENSMUSG00000025821 | 47877284 | 47877684 | 2.03 | 80 | inside |
| ENSMUSG00000029754 | 6863543 | 6863943 | 2.03 | 209 | inside |
| ENSMUSG00000030279 | 143099123 | 143099623 | 2.03 | 984 | inside |
| ENSMUSG00000030512 | 66060687 | 66061087 | 2.03 | 388 | inside |


| ENSMUSG00000041037 | 24533237 | 24533987 | 2.03 | 2589 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000030731 | 44352877 | 44353427 | 2.03 | -31225 | intergenic |
| ENSMUSG00000038296 | 111780032 | 111780432 | 2.03 | -55 | upstream |
| ENSMUSG00000058886 | 141327411 | 141327911 | 2.03 | 314 | overlap |
| ENSMUSG00000031910 | 106870389 | 106870839 | 2.03 | 147 | inside |
| ENSMUSG00000079070 | 32883250 | 32883850 | 2.03 | 66776 | downstream |
| ENSMUSG00000079157 | 9770530 | 9771730 | 2.03 | 488 | overlap |
| ENSMUSG00000031654 | 87472473 | 87472873 | 2.03 | 119 | overlap |
| ENSMUSG00000032477 | 109875831 | 109876331 | 2.03 | 252 | inside |
| ENSMUSG00000032612 | 108347454 | 108348054 | 2.03 | -377 | overlap |
| ENSMUSG00000041064 | 65587628 | 65588078 | 2.03 | 468 | inside |
| ENSMUSG00000052428 | 167308296 | 167308796 | 2.02 | -374 | overlap |
| ENSMUSG00000026123 | 34849584 | 34850084 | 2.02 | -392 | overlap |
| ENSMUSG00000026238 | 86527564 | 86528064 | 2.02 | 828 | inside |
| ENSMUSG00000023150 | 151344489 | 151345039 | 2.02 | -9 | overlap |
| ENSMUSG00000026255 | 87264352 | 87264752 | 2.02 | -11 | overlap |
| ENSMUSG00000091476 | 177983107 | 177983507 | 2.02 | -8328 | intergenic |
| ENSMUSG00000097934 | 20890388 | 20890788 | 2.02 | 85 | overlap |
| ENSMUSG00000073530 | 159021758 | 159022208 | 2.02 | -41268 | intergenic |
| ENSMUSG00000098876 | 47420735 | 47421935 | 2.02 | 46390 | intergenic |
| ENSMUSG00000026478 | 153332539 | 153332939 | 2.02 | 247 | overlap |
| ENSMUSG00000019943 | 98913195 | 98914345 | 2.02 | -1957 | upstream |
| ENSMUSG00000006342 | 75667698 | 75668148 | 2.02 | -23690 | intergenic |
| ENSMUSG00000020133 | 80323506 | 80324106 | 2.02 | -2969 | upstream |
| ENSMUSG00000087833 | 17586626 | 17587026 | 2.02 | 38340 | intergenic |
| ENSMUSG00000017548 | 79992927 | 79993377 | 2.02 | -179 | overlap |
| ENSMUSG00000000384 | 6625576 | 6626126 | 2.02 | 491 | overlap |
| ENSMUSG00000018446 | 70982780 | 70983180 | 2.02 | 246 | overlap |
| ENSMUSG00000020176 | 12037037 | 12037787 | 2.02 | 1646 | inside |
| ENSMUSG00000020660 | 3929905 | 3930405 | 2.02 | -25046 | intergenic |
| ENSMUSG00000020646 | 24830890 | 24831340 | 2.02 | -709 | upstream |
| ENSMUSG00000048982 | 75371768 | 75372968 | 2.02 | 45013 | intergenic |
| ENSMUSG00000021248 | 85374277 | 85374827 | 2.02 | 440 | overlap |
| ENSMUSG00000064972 | 94714945 | 94715945 | 2.02 | 664006 | intergenic |
| ENSMUSG00000005583 | 82859441 | 82860441 | 2.02 | -644593 | intergenic |
| ENSMUSG00000077473 | 4983210 | 4983810 | 2.02 | 206724 | intergenic |
| ENSMUSG00000022269 | 26309041 | 26309441 | 2.02 | -7 | overlap |
| ENSMUSG00000047166 | 75909716 | 75910116 | 2.02 | -44 | overlap |
| ENSMUSG00000022629 | 91048815 | 91049315 | 2.02 | 1133 | inside |
| ENSMUSG00000037458 | 38518098 | 38518548 | 2.02 | 1168 | inside |
| ENSMUSG00000022840 | 35154774 | 35155174 | 2.02 | -103 | overlap |
| ENSMUSG00000022883 | 72663108 | 72663508 | 2.02 | -41 | overlap |
| ENSMUSG00000097139 | 92697500 | 92697950 | 2.02 | 84686 | intergenic |
| ENSMUSG00000040732 | 95585157 | 95585657 | 2.02 | 1436 | inside |
| ENSMUSG00000038037 | 10784560 | 10785260 | 2.02 | 976 | inside |
| ENSMUSG00000095407 | 68840677 | 68841227 | 2.02 | 3541 | inside |
| ENSMUSG00000024130 | 24351311 | 24352361 | 2.02 | -639 | overlap |
| ENSMUSG00000073394 | 44735324 | 44736074 | 2.02 | -521 | overlap |
| ENSMUSG00000045215 | 22344412 | 22344962 | 2.02 | -471 | overlap |
| ENSMUSG00000087960 | 19312554 | 19313754 | 2.02 | -394539 | intergenic |
| ENSMUSG00000035342 | 45028463 | 45029013 | 2.02 | 13287 | downstream |


| ENSMUSG00000056829 | 16872425 | 16872825 | 2.02 | 1405 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000042401 | 42432263 | 42432663 | 2.02 | -480 | upstream |
| ENSMUSG00000024870 | 5106853 | 5107303 | 2.02 | 143 | overlap |
| ENSMUSG00000043342 | 74698564 | 74698964 | 2.02 | 884 | inside |
| ENSMUSG00000026790 | 29889915 | 29890315 | 2.02 | 694 | inside |
| ENSMUSG00000027207 | 125858535 | 125858985 | 2.02 | -574 | upstream |
| ENSMUSG00000035877 | 160872949 | 160873399 | 2.02 | 49 | overlap |
| ENSMUSG00000063275 | 14055532 | 14056282 | 2.02 | 603 | overlap |
| ENSMUSG00000046470 | 181670280 | 181670830 | 2.02 | 1360 | inside |
| ENSMUSG00000051817 | 152397890 | 152398540 | 2.02 | 173 | overlap |
| ENSMUSG00000027544 | 168601326 | 168601876 | 2.02 | 331 | overlap |
| ENSMUSG00000039059 | 180103230 | 180103780 | 2.02 | 1258 | inside |
| ENSMUSG00000026779 | 23155615 | 23156065 | 2.02 | 409 | overlap |
| ENSMUSG00000093230 | 87617067 | 87617917 | 2.02 | -332 | includeFeature |
| ENSMUSG00000070227 | 24599267 | 24600267 | 2.02 | 37193 | intergenic |
| ENSMUSG00000039617 | 20955180 | 20955780 | 2.02 | -94265 | intergenic |
| ENSMUSG00000061143 | 52104467 | 52104967 | 2.02 | 424 | overlap |
| ENSMUSG00000047281 | 133599865 | 133600265 | 2.02 | 2303 | downstream |
| ENSMUSG00000028612 | 107846017 | 107846417 | 2.02 | -15173 | intergenic |
| ENSMUSG00000098364 | 138324747 | 138325997 | 2.02 | 1342 | overlapEnd |
| ENSMUSG00000097904 | 110386132 | 110386632 | 2.02 | 297 | inside |
| ENSMUSG00000063146 | 134551659 | 134552109 | 2.02 | 775 | inside |
| ENSMUSG00000029128 | 41566018 | 41566518 | 2.02 | 142137 | intergenic |
| ENSMUSG00000029705 | 136565937 | 136566337 | 2.02 | 1553 | inside |
| ENSMUSG00000043059 | 31200304 | 31200704 | 2.02 | 1999 | inside |
| ENSMUSG00000033726 | 85187810 | 85188410 | 2.02 | 372 | inside |
| ENSMUSG00000073155 | 35252343 | 35252743 | 2.02 | -311 | overlap |
| ENSMUSG00000052751 | 48594044 | 48594444 | 2.02 | 161 | inside |
| ENSMUSG00000038759 | 35177404 | 35177804 | 2.02 | -26 | overlap |
| ENSMUSG00000030243 | 142386823 | 142387223 | 2.02 | 264 | overlap |
| ENSMUSG00000055633 | 5052730 | 5053430 | 2.02 | 1198 | inside |
| ENSMUSG00000041420 | 16158497 | 16159047 | 2.02 | -16593 | intergenic |
| ENSMUSG00000001918 | 16781964 | 16782364 | 2.02 | 618 | inside |
| ENSMUSG00000030725 | 100159186 | 100159586 | 2.02 | -91 | overlap |
| ENSMUSG00000093405 | 65862446 | 65862946 | 2.02 | -454 | upstream |
| ENSMUSG00000030590 | 29161394 | 29161794 | 2.02 | -5160 | intergenic |
| ENSMUSG00000045777 | 142371818 | 142372418 | 2.02 | 1935 | inside |
| ENSMUSG00000005575 | 13038007 | 13038457 | 2.02 | 268 | overlap |
| ENSMUSG00000003863 | 45366506 | 45366906 | 2.02 | 513 | inside |
| ENSMUSG00000097424 | 121553822 | 121554322 | 2.02 | 9440 | intergenic |
| ENSMUSG00000031749 | 110919230 | 110919980 | 2.02 | -692 | overlap |
| ENSMUSG00000014907 | 66860440 | 66860840 | 2.02 | 223 | inside |
| ENSMUSG00000007950 | 71463209 | 71463609 | 2.02 | 448 | inside |
| ENSMUSG00000001911 | 84798973 | 84799373 | 2.02 | 1371 | inside |
| ENSMUSG00000044006 | 69881023 | 69881523 | 2.02 | 6664 | inside |
| ENSMUSG00000035606 | 102506005 | 102506405 | 2.02 | -133 | overlap |
| ENSMUSG00000049932 | 44334902 | 44335502 | 2.02 | 187 | inside |
| ENSMUSG00000032504 | 113590217 | 113590717 | 2.02 | 118042 | intergenic |
| ENSMUSG00000032128 | 37459850 | 37460400 | 2.02 | -26687 | intergenic |
| ENSMUSG00000026167 | 74791615 | 74792015 | 2.01 | 99 | inside |
| ENSMUSG00000044340 | 106172362 | 106172762 | 2.01 | 610 | inside |


| ENSMUSG00000026283 | 93804117 | 93804517 | 2.01 | 152 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000026167 | 74803215 | 74803615 | 2.01 | 11699 | inside |
| ENSMUSG00000026356 | 128416917 | 128417317 | 2.01 | 499 | inside |
| ENSMUSG00000038608 | 80758614 | 80759014 | 2.01 | -61 | upstream |
| ENSMUSG00000045005 | 64736414 | 64737464 | 2.01 | 1336 | inside |
| ENSMUSG00000035262 | 80806956 | 80807356 | 2.01 | 1708 | inside |
| ENSMUSG00000019996 | 20149345 | 20149745 | 2.01 | 874 | inside |
| ENSMUSG00000040054 | 128096737 | 128097137 | 2.01 | 3954 | inside |
| ENSMUSG00000061904 | 91123165 | 91123665 | 2.01 | 894 | inside |
| ENSMUSG00000054450 | 53478677 | 53479077 | 2.01 | -1489 | upstream |
| ENSMUSG00000020458 | 29693529 | 29694029 | 2.01 | 582 | inside |
| ENSMUSG00000020471 | 5878076 | 5878476 | 2.01 | 216 | overlap |
| ENSMUSG00000046474 | 99851065 | 99851465 | 2.01 | 543 | inside |
| ENSMUSG00000017291 | 77607677 | 77608077 | 2.01 | 138 | overlap |
| ENSMUSG00000021124 | 79171961 | 79172361 | 2.01 | 706 | inside |
| ENSMUSG00000017756 | 73763768 | 73764168 | 2.01 | 71 | inside |
| ENSMUSG00000021493 | 55513158 | 55513558 | 2.01 | 518 | inside |
| ENSMUSG00000021572 | 74061718 | 74062168 | 2.01 | 567 | inside |
| ENSMUSG00000038372 | 32338197 | 32338697 | 2.01 | 515 | inside |
| ENSMUSG00000090907 | 92426661 | 92427061 | 2.01 | 563 | inside |
| ENSMUSG00000040717 | 27038794 | 27039194 | 2.01 | -207 | overlap |
| ENSMUSG00000099315 | 69502128 | 69502628 | 2.01 | -728 | upstream |
| ENSMUSG00000065238 | 99721058 | 99721508 | 2.01 | 52750 | intergenic |
| ENSMUSG00000090691 | 6889421 | 6889971 | 2.01 | 541 | overlap |
| ENSMUSG00000033004 | 103345996 | 103346446 | 2.01 | 818 | inside |
| ENSMUSG00000064959 | 82340771 | 82341371 | 2.01 | -1882 | upstream |
| ENSMUSG00000036661 | 73512573 | 73512973 | 2.01 | 13 | inside |
| ENSMUSG00000089837 | 79833021 | 79833471 | 2.01 | 1216 | inside |
| ENSMUSG00000022999 | 98917665 | 98918315 | 2.01 | 566 | overlap |
| ENSMUSG00000022994 | 98609315 | 98609715 | 2.01 | -1682 | upstream |
| ENSMUSG00000043683 | 56257334 | 56257784 | 2.01 | 541 | inside |
| ENSMUSG00000024327 | 34031578 | 34031978 | 2.01 | 112 | overlap |
| ENSMUSG00000024304 | 16808804 | 16809454 | 2.01 | 442 | overlap |
| ENSMUSG00000025081 | 56825725 | 56826225 | 2.01 | -484 | overlap |
| ENSMUSG00000039126 | 16955725 | 16956175 | 2.01 | -393 | overlap |
| ENSMUSG00000075044 | 8251775 | 8252275 | 2.01 | -32875 | intergenic |
| ENSMUSG00000040929 | 28155220 | 28155620 | 2.01 | -144054 | intergenic |
| ENSMUSG00000035576 | 162943099 | 162943849 | 2.01 | -373 | overlap |
| ENSMUSG00000017740 | 164968285 | 164968935 | 2.01 | 7469 | inside |
| ENSMUSG00000039849 | 164879735 | 164880235 | 2.01 | 431 | inside |
| ENSMUSG00000029419 | 25578989 | 25579589 | 2.01 | 1110 | inside |
| ENSMUSG00000055897 | 173659388 | 173659838 | 2.01 | 252 | overlap |
| ENSMUSG00000026836 | 58566674 | 58567324 | 2.01 | 483 | overlap |
| ENSMUSG00000074637 | 34648467 | 34648917 | 2.01 | -1538 | upstream |
| ENSMUSG00000073752 | 130308774 | 130309624 | 2.01 | -100 | upstream |
| ENSMUSG00000028744 | 139310397 | 139310797 | 2.01 | 311 | overlap |
| ENSMUSG00000037962 | 125028637 | 125029187 | 2.01 | 25190 | intergenic |
| ENSMUSG00000037962 | 125003373 | 125003773 | 2.01 | -74 | overlap |
| ENSMUSG00000029229 | 75043618 | 75044218 | 2.01 | 1156 | inside |
| ENSMUSG00000034040 | 131306937 | 131307437 | 2.01 | 1141 | inside |
| ENSMUSG00000029673 | 132543437 | 132543887 | 2.01 | -93 | upstream |


| ENSMUSG00000047881 | 63968868 | 63969268 | 2.01 | 37 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000007415 | 3647107 | 3647657 | 2.01 | 827 | inside |
| ENSMUSG00000009471 | 46348706 | 46349106 | 2.01 | -27768 | intergenic |
| ENSMUSG00000060260 | 139247133 | 139247683 | 2.01 | -1349 | upstream |
| ENSMUSG00000037772 | 142532741 | 142533191 | 2.01 | -58 | overlap |
| ENSMUSG00000002083 | 16313797 | 16314347 | 2.01 | 4181 | inside |
| ENSMUSG00000041769 | 138830099 | 138830699 | 2.01 | -15980 | intergenic |
| ENSMUSG00000059263 | 112023549 | 112024049 | 2.01 | 43 | inside |
| ENSMUSG00000030402 | 19279274 | 19279774 | 2.01 | 775 | inside |
| ENSMUSG00000084882 | 45632443 | 45632943 | 2.01 | 634 | inside |
| ENSMUSG00000052566 | 84990359 | 84990959 | 2.01 | -236 | overlap |
| ENSMUSG00000074357 | 47674935 | 47675585 | 2.01 | -456 | overlap |
| ENSMUSG00000031849 | 70374181 | 70374831 | 2.01 | 633 | inside |
| ENSMUSG00000006276 | 72421073 | 72421473 | 2.01 | 401 | inside |
| ENSMUSG00000032024 | 40685452 | 40686102 | 2.01 | -510 | overlap |
| ENSMUSG00000057626 | 105568005 | 105569355 | 2.01 | -7243 | intergenic |
| ENSMUSG00000032065 | 50560930 | 50561330 | 2.01 | 338 | overlap |
| ENSMUSG00000032515 | 119983217 | 119983767 | 2.01 | -5967 | intergenic |
| ENSMUSG00000066877 | 43446194 | 43446594 | 2.00 | 443 | inside |
| ENSMUSG00000073586 | 123857012 | 123857412 | 2.00 | -187811 | intergenic |
| ENSMUSG00000077846 | 134799262 | 134799862 | 2.00 | -10013 | intergenic |
| ENSMUSG00000026604 | 189728110 | 189728510 | 2.00 | -158 | overlap |
| ENSMUSG00000073486 | 179802658 | 179803208 | 2.00 | -718 | upstream |
| ENSMUSG00000006301 | 74304065 | 74304465 | 2.00 | 1557 | inside |
| ENSMUSG00000026107 | 51433135 | 51433985 | 2.00 | 45264 | intergenic |
| ENSMUSG00000026566 | 165642396 | 165643146 | 2.00 | -7855 | intergenic |
| ENSMUSG00000060935 | 85102863 | 85103263 | 2.00 | 236 | inside |
| ENSMUSG00000039697 | 30802195 | 30802895 | 2.00 | 912 | inside |
| ENSMUSG00000069814 | 74619630 | 74620030 | 2.00 | 25 | inside |
| ENSMUSG00000039230 | 121451786 | 121452186 | 2.00 | -163 | overlap |
| ENSMUSG00000087042 | 97534415 | 97534965 | 2.00 | -12520 | intergenic |
| ENSMUSG00000086058 | 82931186 | 82931586 | 2.00 | 2196 | inside |
| ENSMUSG00000040548 | 106612315 | 106612765 | 2.00 | 1108 | inside |
| ENSMUSG00000048562 | 118836654 | 118837204 | 2.00 | -9675 | intergenic |
| ENSMUSG00000021385 | 49399247 | 49399647 | 2.00 | -22064 | intergenic |
| ENSMUSG00000021338 | 24281121 | 24281521 | 2.00 | -326 | upstream |
| ENSMUSG00000021569 | 73937192 | 73937642 | 2.00 | 575 | inside |
| ENSMUSG00000048904 | 56252005 | 56252455 | 2.00 | 158 | overlap |
| ENSMUSG00000035953 | 50930342 | 50930792 | 2.00 | 514 | inside |
| ENSMUSG00000023034 | 101266370 | 101267220 | 2.00 | -476 | overlap |
| ENSMUSG00000056605 | 101785770 | 101786220 | 2.00 | 688 | inside |
| ENSMUSG00000096883 | 82207016 | 82207416 | 2.00 | 5799 | inside |
| ENSMUSG00000022971 | 91372827 | 91373227 | 2.00 | 44 | inside |
| ENSMUSG00000022808 | 33251078 | 33251978 | 2.00 | -378 | overlap |
| ENSMUSG00000046962 | 97962107 | 97962707 | 2.00 | 514 | overlap |
| ENSMUSG00000008393 | 8671530 | 8671980 | 2.00 | 623 | inside |
| ENSMUSG00000002844 | 38452115 | 38452765 | 2.00 | 574 | overlap |
| ENSMUSG00000093026 | 61398409 | 61398809 | 2.00 | 596 | downstream |
| ENSMUSG00000054072 | 60339759 | 60340809 | 2.00 | -36270 | intergenic |
| ENSMUSG00000060534 | 72576509 | 72577009 | 2.00 | -225440 | intergenic |
| ENSMUSG00000024268 | 25678601 | 25679101 | 2.00 | 75382 | inside |


| ENSMUSG00000025049 | 47068175 | 47068575 | 2.00 | 427 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000007338 | 6057360 | 6057960 | 2.00 | 391 | overlap |
| ENSMUSG00000051984 | 44545354 | 44545754 | 2.00 | 510 | inside |
| ENSMUSG00000032802 | 152093635 | 152094085 | 2.00 | -11881 | intergenic |
| ENSMUSG00000074876 | 122630407 | 122630807 | 2.00 | -218 | overlap |
| ENSMUSG00000027569 | 180582334 | 180582734 | 2.00 | 1030 | inside |
| ENSMUSG00000038831 | 33371265 | 33371665 | 2.00 | 221 | overlap |
| ENSMUSG00000083325 | 150500299 | 150500799 | 2.00 | 546 | inside |
| ENSMUSG00000027434 | 147193923 | 147194323 | 2.00 | 320 | overlap |
| ENSMUSG00000056476 | 59005845 | 59006245 | 2.00 | -1133 | upstream |
| ENSMUSG00000027793 | 55054703 | 55055153 | 2.00 | 352 | overlap |
| ENSMUSG00000059857 | 110142450 | 110142850 | 2.00 | 778 | inside |
| ENSMUSG00000040809 | 106171599 | 106172099 | 2.00 | -4035 | upstream |
| ENSMUSG00000041263 | 89090403 | 89090803 | 2.00 | 2960 | inside |
| ENSMUSG00000043572 | 106638737 | 106639237 | 2.00 | -12332 | intergenic |
| ENSMUSG00000028243 | 6191115 | 6191515 | 2.00 | 17 | inside |
| ENSMUSG00000077260 | 102494477 | 102494877 | 2.00 | -242335 | intergenic |
| ENSMUSG00000025743 | 130792507 | 130792907 | 2.00 | -30 | overlap |
| ENSMUSG00000029076 | 155993071 | 155993471 | 2.00 | 199 | inside |
| ENSMUSG00000054679 | 33248107 | 33248857 | 2.00 | 39116 | intergenic |
| ENSMUSG00000028521 | 103214967 | 103215567 | 2.00 | 197 | overlap |
| ENSMUSG00000060862 | 137048415 | 137048815 | 2.00 | 386 | overlap |
| ENSMUSG00000039682 | 45493412 | 45493862 | 2.00 | 38 | inside |
| ENSMUSG00000061882 | 123930420 | 123931020 | 2.00 | 3002 | inside |
| ENSMUSG00000043059 | 31201856 | 31202256 | 2.00 | 447 | inside |
| ENSMUSG00000037822 | 65492604 | 65493004 | 2.00 | 44580 | inside |
| ENSMUSG00000051391 | 135933837 | 135934787 | 2.00 | 779 | overlap |
| ENSMUSG00000097626 | 19226267 | 19226667 | 2.00 | 288 | overlap |
| ENSMUSG00000072641 | 121869420 | 121869820 | 2.00 | -10452 | intergenic |
| ENSMUSG00000029860 | 42349692 | 42350142 | 2.00 | -136 | overlap |
| ENSMUSG00000015053 | 88193839 | 88194239 | 2.00 | -52 | overlap |
| ENSMUSG00000029687 | 47594119 | 47594669 | 2.00 | 1222 | inside |
| ENSMUSG00000030732 | 100042053 | 100042553 | 2.00 | 35649 | intergenic |
| ENSMUSG00000031068 | 137437680 | 137438280 | 2.00 | 32 | inside |
| ENSMUSG00000036862 | 105786768 | 105787318 | 2.00 | 886 | inside |
| ENSMUSG00000000131 | 126200312 | 126200712 | 2.00 | 189 | overlap |
| ENSMUSG00000043866 | 105743699 | 105744099 | 2.00 | 662 | inside |
| ENSMUSG00000077409 | 142082427 | 142082977 | 2.00 | -3990 | upstream |
| ENSMUSG00000003824 | 84871959 | 84872359 | 2.00 | -152 | overlap |
| ENSMUSG00000031887 | 105264089 | 105264589 | 2.00 | 520 | inside |
| ENSMUSG00000031673 | 102784522 | 102784972 | 2.00 | 589 | inside |
| ENSMUSG00000031681 | 79398681 | 79399131 | 2.00 | 837 | inside |
| ENSMUSG00000032309 | 55208578 | 55208978 | 2.00 | -347 | overlap |
| ENSMUSG00000032303 | 55026178 | 55026578 | 2.00 | 381 | overlap |
| ENSMUSG00000032368 | 91381248 | 91381798 | 2.00 | -15438 | intergenic |
| ENSMUSG00000093745 | 42520503 | 42520953 | 2.00 | 328 | includeFeature |
| ENSMUSG00000032462 | 99140266 | 99140666 | 2.00 | 355 | overlap |
| ENSMUSG00000004936 | 64253329 | 64253779 | 2.00 | 302 | overlap |
| ENSMUSG00000048960 | 10993707 | 10994107 | 1.99 | 242 | inside |
| ENSMUSG00000079330 | 132190212 | 132190962 | 1.99 | -1224 | upstream |
| ENSMUSG00000036206 | 89070252 | 89070752 | 1.99 | -163 | overlap |


| ENSMUSG00000052062 | 61639059 | 61639509 | 1.99 | 235 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000025925 | 15805546 | 15805996 | 1.99 | -112 | overlap |
| ENSMUSG00000026494 | 178530313 | 178530763 | 1.99 | 1188 | inside |
| ENSMUSG00000098815 | 117587011 | 117587611 | 1.99 | 25781 | intergenic |
| ENSMUSG00000041075 | 59481944 | 59482344 | 1.99 | -203 | overlap |
| ENSMUSG00000033722 | 155244308 | 155244708 | 1.99 | -11584 | intergenic |
| ENSMUSG00000081623 | 65979920 | 65980370 | 1.99 | 53005 | intergenic |
| ENSMUSG00000035206 | 80855398 | 80855898 | 1.99 | 123 | inside |
| ENSMUSG00000097007 | 11280531 | 11281131 | 1.99 | 690 | inside |
| ENSMUSG00000020232 | 81350083 | 81350483 | 1.99 | 397 | overlap |
| ENSMUSG00000048217 | 102145265 | 102145665 | 1.99 | -248 | overlap |
| ENSMUSG00000018765 | 69633148 | 69633648 | 1.99 | 158 | inside |
| ENSMUSG00000036264 | 52764127 | 52764677 | 1.99 | -507 | overlap |
| ENSMUSG00000017774 | 75655580 | 75656080 | 1.99 | 5076 | inside |
| ENSMUSG00000023764 | 3180626 | 3181026 | 1.99 | 12837 | inside |
| ENSMUSG00000086851 | 53567086 | 53567536 | 1.99 | 227 | overlap |
| ENSMUSG00000049755 | 58323127 | 58323527 | 1.99 | 219 | overlap |
| ENSMUSG00000037957 | 110738217 | 110738617 | 1.99 | 268 | inside |
| ENSMUSG00000033731 | 99392667 | 99393217 | 1.99 | 51513 | intergenic |
| ENSMUSG00000042029 | 116405172 | 116405672 | 1.99 | -230 | overlap |
| ENSMUSG00000043398 | 72070857 | 72071257 | 1.99 | 134 | overlap |
| ENSMUSG00000020672 | 30372812 | 30373262 | 1.99 | 563 | inside |
| ENSMUSG00000091387 | 96925001 | 96925601 | 1.99 | 312 | inside |
| ENSMUSG00000021423 | 37406847 | 37407997 | 1.99 | 61502 | inside |
| ENSMUSG00000038246 | 34746847 | 34747247 | 1.99 | 11997 | inside |
| ENSMUSG00000021596 | 76353358 | 76353758 | 1.99 | -31177 | intergenic |
| ENSMUSG00000021685 | 94873435 | 94873885 | 1.99 | -2167 | upstream |
| ENSMUSG00000055137 | 17694632 | 17695782 | 1.99 | 100 | overlap |
| ENSMUSG00000089827 | 80885654 | 80886054 | 1.99 | -130 | upstream |
| ENSMUSG00000087286 | 59878926 | 59880176 | 1.99 | 161565 | intergenic |
| ENSMUSG00000021983 | 60176975 | 60177375 | 1.99 | 20204 | inside |
| ENSMUSG00000002320 | 55643298 | 55643798 | 1.99 | 508 | inside |
| ENSMUSG00000097286 | 54952296 | 54952696 | 1.99 | -652 | upstream |
| ENSMUSG00000034161 | 76457825 | 76458225 | 1.99 | 387 | inside |
| ENSMUSG00000016541 | 85336571 | 85337021 | 1.99 | 190 | inside |
| ENSMUSG00000097259 | 37007498 | 37007998 | 1.99 | -25 | overlap |
| ENSMUSG00000000555 | 103366015 | 103366665 | 1.99 | 748 | inside |
| ENSMUSG00000002524 | 76079973 | 76080573 | 1.99 | 973 | inside |
| ENSMUSG00000088128 | 76178571 | 76179771 | 1.99 | 35634 | intergenic |
| ENSMUSG00000022568 | 76068716 | 76069266 | 1.99 | 1068 | inside |
| ENSMUSG00000053774 | 32332109 | 32332859 | 1.99 | -143 | overlap |
| ENSMUSG00000039427 | 5233571 | 5234021 | 1.99 | -50 | overlap |
| ENSMUSG00000040681 | 96126916 | 96127516 | 1.99 | 813 | inside |
| ENSMUSG00000041130 | 24669965 | 24670365 | 1.99 | 213 | inside |
| ENSMUSG00000024220 | 28176678 | 28177228 | 1.99 | -529 | overlap |
| ENSMUSG00000040276 | 27655172 | 27655672 | 1.99 | -337 | overlap |
| ENSMUSG00000045999 | 39848628 | 39849078 | 1.99 | 199 | overlap |
| ENSMUSG00000024112 | 25433115 | 25433515 | 1.99 | 668 | inside |
| ENSMUSG00000023828 | 12506941 | 12507391 | 1.99 | 763 | inside |
| ENSMUSG00000023951 | 46031268 | 46031818 | 1.99 | 1109 | inside |
| ENSMUSG00000024530 | 67464959 | 67465359 | 1.99 | 110 | inside |


| ENSMUSG00000056481 | 5068253 | 5068653 | 1.99 | 175 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000024776 | 34192320 | 34192770 | 1.99 | 91 | inside |
| ENSMUSG00000074923 | 118676635 | 118677135 | 1.99 | 13332 | inside |
| ENSMUSG00000068154 | 146221335 | 146221735 | 1.99 | -586 | upstream |
| ENSMUSG00000027455 | 151494529 | 151494979 | 1.99 | 347 | inside |
| ENSMUSG00000045319 | 6100275 | 6100875 | 1.99 | 29936 | inside |
| ENSMUSG00000000823 | 181592716 | 181593116 | 1.99 | 87 | overlap |
| ENSMUSG00000050896 | 84871614 | 84872014 | 1.99 | 15096 | overlapEnd |
| ENSMUSG00000026885 | 35979665 | 35980065 | 1.99 | 248 | overlap |
| ENSMUSG00000027660 | 31095496 | 31095896 | 1.99 | 438 | inside |
| ENSMUSG00000002233 | 104788871 | 104789321 | 1.99 | -140 | overlap |
| ENSMUSG00000027985 | 131111150 | 131111600 | 1.99 | 679 | inside |
| ENSMUSG00000028102 | 96635529 | 96635929 | 1.99 | 153 | inside |
| ENSMUSG00000027894 | 107516950 | 107517800 | 1.99 | 1068 | inside |
| ENSMUSG00000028710 | 115784925 | 115785325 | 1.99 | 113 | inside |
| ENSMUSG00000028811 | 129223298 | 129223948 | 1.99 | 33538 | downstream |
| ENSMUSG00000082195 | 146066779 | 146067579 | 1.99 | -745 | overlap |
| ENSMUSG00000028542 | 117835537 | 117835937 | 1.99 | 1031 | inside |
| ENSMUSG00000043753 | 89688551 | 89688951 | 1.99 | 9115 | inside |
| ENSMUSG00000028756 | 138326097 | 138326497 | 1.99 | 210 | overlap |
| ENSMUSG00000039809 | 46990800 | 46991200 | 1.99 | 1073 | inside |
| ENSMUSG00000028546 | 110186793 | 110187193 | 1.99 | 165116 | intergenic |
| ENSMUSG00000084179 | 23657525 | 23657975 | 1.99 | 2819 | downstream |
| ENSMUSG00000049686 | 123014576 | 123014976 | 1.99 | -498 | upstream |
| ENSMUSG00000016503 | 146948775 | 146949225 | 1.99 | 118 | inside |
| ENSMUSG00000029359 | 118026692 | 118027992 | 1.99 | -1051 | overlap |
| ENSMUSG00000085376 | 116005684 | 116006584 | 1.99 | -3632 | upstream |
| ENSMUSG00000001566 | 29467637 | 29468387 | 1.99 | 10882 | downstream |
| ENSMUSG00000028959 | 24444436 | 24445086 | 1.99 | 851 | inside |
| ENSMUSG00000029363 | 117388948 | 117389398 | 1.99 | 99 | overlap |
| ENSMUSG00000030166 | 119902673 | 119903123 | 1.99 | -25 | overlap |
| ENSMUSG00000096282 | 74530066 | 74530516 | 1.99 | -24902 | intergenic |
| ENSMUSG00000053907 | 72438774 | 72439224 | 1.99 | 784 | inside |
| ENSMUSG00000050917 | 144861608 | 144862008 | 1.99 | 222 | inside |
| ENSMUSG00000035354 | 99140363 | 99140763 | 1.99 | 781 | inside |
| ENSMUSG00000010476 | 137309063 | 137309463 | 1.99 | 5382 | inside |
| ENSMUSG00000097842 | 16874747 | 16875647 | 1.99 | 139 | overlap |
| ENSMUSG00000074004 | 98193413 | 98193863 | 1.99 | 6068 | inside |
| ENSMUSG00000064450 | 123102508 | 123102908 | 1.99 | -530 | upstream |
| ENSMUSG00000088312 | 42904835 | 42905335 | 1.99 | 162028 | intergenic |
| ENSMUSG00000002396 | 71375509 | 71375909 | 1.99 | 4211 | inside |
| ENSMUSG00000037300 | 124721289 | 124721789 | 1.99 | 694 | inside |
| ENSMUSG00000031487 | 27121267 | 27121867 | 1.99 | 7365 | downstream |
| ENSMUSG00000032872 | 87021600 | 87022050 | 1.99 | -414 | overlap |
| ENSMUSG00000010607 | 22164641 | 22165091 | 1.99 | 7795 | intergenic |
| ENSMUSG00000037808 | 13827441 | 13827891 | 1.99 | -275 | overlap |
| ENSMUSG00000097820 | 121759168 | 121759818 | 1.99 | 620 | overlap |
| ENSMUSG00000032366 | 67043428 | 67043828 | 1.99 | 5978 | inside |
| ENSMUSG00000067028 | 99592112 | 99592912 | 1.98 | -180653 | intergenic |
| ENSMUSG00000090071 | 74855559 | 74856309 | 1.98 | 530 | inside |
| ENSMUSG00000079330 | 132190862 | 132191262 | 1.98 | -574 | upstream |


| ENSMUSG00000067081 | 89992913 | 89993313 | 1.98 | 21753 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000067071 | 91414067 | 91414467 | 1.98 | -29 | upstream |
| ENSMUSG00000026209 | 75317319 | 75317869 | 1.98 | 318 | overlap |
| ENSMUSG00000046980 | 73099403 | 73100053 | 1.98 | -51 | includeFeature |
| ENSMUSG00000019907 | 108161973 | 108162623 | 1.98 | -427 | overlap |
| ENSMUSG00000084658 | 80381184 | 80381784 | 1.98 | -1118 | upstream |
| ENSMUSG00000097086 | 21992995 | 21994095 | 1.98 | 53990 | intergenic |
| ENSMUSG00000059901 | 61272753 | 61273253 | 1.98 | 685 | inside |
| ENSMUSG00000020788 | 72960977 | 72961377 | 1.98 | -192 | overlap |
| ENSMUSG00000050288 | 102604265 | 102605315 | 1.98 | -166 | overlap |
| ENSMUSG00000070345 | 87616827 | 87617877 | 1.98 | -337 | overlap |
| ENSMUSG00000047284 | 69901830 | 69902230 | 1.98 | 758 | inside |
| ENSMUSG00000019312 | 98458015 | 98458465 | 1.98 | 11621 | downstream |
| ENSMUSG00000040610 | 33201382 | 33201782 | 1.98 | 2206 | inside |
| ENSMUSG00000020176 | 12025576 | 12025976 | 1.98 | 13107 | inside |
| ENSMUSG00000041771 | 102128917 | 102129317 | 1.98 | 184 | inside |
| ENSMUSG00000089603 | 60385011 | 60386361 | 1.98 | -350487 | intergenic |
| ENSMUSG00000072949 | 84009077 | 84009627 | 1.98 | -425 | overlap |
| ENSMUSG00000041669 | 103242117 | 103242517 | 1.98 | 33 | overlap |
| ENSMUSG00000063200 | 43398447 | 43398847 | 1.98 | 71 | inside |
| ENSMUSG00000021423 | 37404887 | 37405437 | 1.98 | 59542 | inside |
| ENSMUSG00000051627 | 23622212 | 23622612 | 1.98 | 290 | overlap |
| ENSMUSG00000021451 | 51792597 | 51793197 | 1.98 | 1150 | inside |
| ENSMUSG00000021991 | 29722077 | 29722627 | 1.98 | -213 | upstream |
| ENSMUSG00000022255 | 34082696 | 34083346 | 1.98 | 2 | inside |
| ENSMUSG00000033170 | 78802171 | 78802571 | 1.98 | 871 | inside |
| ENSMUSG00000095440 | 101078765 | 101079265 | 1.98 | -24366 | intergenic |
| ENSMUSG00000046546 | 30599817 | 30600567 | 1.98 | 94 | inside |
| ENSMUSG00000039903 | 90830708 | 90831258 | 1.98 | 3989 | inside |
| ENSMUSG00000022760 | 17530480 | 17530880 | 1.98 | 656 | inside |
| ENSMUSG00000063952 | 28801362 | 28801762 | 1.98 | 272 | inside |
| ENSMUSG00000091705 | 35342278 | 35342678 | 1.98 | 36 | inside |
| ENSMUSG00000044279 | 57059194 | 57059644 | 1.98 | 89 | inside |
| ENSMUSG00000095325 | 32859222 | 32859622 | 1.98 | 26861 | intergenic |
| ENSMUSG00000024165 | 24960512 | 24960912 | 1.98 | 124 | overlap |
| ENSMUSG00000037013 | 14682972 | 14683422 | 1.98 | -58 | upstream |
| ENSMUSG00000040385 | 4191566 | 4192166 | 1.98 | -592 | overlap |
| ENSMUSG00000085196 | 6276560 | 6277010 | 1.98 | 45 | overlap |
| ENSMUSG00000038467 | 154645090 | 154645940 | 1.98 | -6615 | intergenic |
| ENSMUSG00000060227 | 121867149 | 121867549 | 1.98 | 179 | inside |
| ENSMUSG00000027652 | 158409999 | 158410499 | 1.98 | 151 | inside |
| ENSMUSG00000068079 | 152142485 | 152143585 | 1.98 | -1076 | overlap |
| ENSMUSG00000026740 | 18391908 | 18392558 | 1.98 | 922 | inside |
| ENSMUSG00000086537 | 174299634 | 174300034 | 1.98 | -4198 | upstream |
| ENSMUSG00000086555 | 35581901 | 35582501 | 1.98 | -23199 | intergenic |
| ENSMUSG00000045493 | 180775871 | 180776921 | 1.98 | 1029 | overlap |
| ENSMUSG00000028161 | 136671388 | 136671838 | 1.98 | 1264 | inside |
| ENSMUSG00000041220 | 129533349 | 129533849 | 1.98 | 963 | inside |
| ENSMUSG00000049119 | 5643795 | 5644395 | 1.98 | -295 | overlap |
| ENSMUSG00000036856 | 137278047 | 137278447 | 1.98 | 558 | inside |
| ENSMUSG00000091921 | 155694803 | 155695203 | 1.98 | 461 | inside |


| ENSMUSG00000094253 | 3546801 | 3547451 | 1.98 | 9711 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000028405 | 40142989 | 40143439 | 1.98 | -92 | overlap |
| ENSMUSG00000034042 | 116557393 | 116557893 | 1.98 | -265 | overlap |
| ENSMUSG00000085482 | 74251844 | 74252244 | 1.98 | -130 | upstream |
| ENSMUSG00000055761 | 20777951 | 20778351 | 1.98 | 915 | inside |
| ENSMUSG00000040659 | 141874207 | 141874707 | 1.98 | 713 | inside |
| ENSMUSG00000046671 | 134534909 | 134535309 | 1.98 | 478 | inside |
| ENSMUSG00000029430 | 129020510 | 129020910 | 1.98 | 441 | inside |
| ENSMUSG00000029310 | 104046416 | 104046916 | 1.98 | 110 | inside |
| ENSMUSG00000085639 | 148928831 | 148929681 | 1.98 | 20035 | intergenic |
| ENSMUSG00000060261 | 134313909 | 134314559 | 1.98 | 851 | inside |
| ENSMUSG00000029603 | 120711242 | 120711742 | 1.98 | 685 | inside |
| ENSMUSG00000086040 | 54429589 | 54430389 | 1.98 | -14 | overlap |
| ENSMUSG00000029999 | 86195449 | 86195899 | 1.98 | 198 | inside |
| ENSMUSG00000030019 | 119479765 | 119481065 | 1.98 | 97 | inside |
| ENSMUSG00000038058 | 54972049 | 54972749 | 1.98 | 563 | overlap |
| ENSMUSG00000030852 | 130646109 | 130646559 | 1.98 | 68625 | inside |
| ENSMUSG00000030664 | 116031509 | 116032009 | 1.98 | -470 | overlap |
| ENSMUSG00000040940 | 24902987 | 24903387 | 1.98 | 75 | inside |
| ENSMUSG00000042462 | 127253682 | 127254132 | 1.98 | 7027 | downstream |
| ENSMUSG00000095115 | 118490959 | 118491509 | 1.98 | 1016 | inside |
| ENSMUSG00000037664 | 143459568 | 143460118 | 1.98 | 1482 | inside |
| ENSMUSG00000004637 | 114439558 | 114440058 | 1.98 | -97 | overlap |
| ENSMUSG00000031902 | 106059389 | 106059789 | 1.98 | -214 | overlap |
| ENSMUSG00000056724 | 110653931 | 110654381 | 1.98 | 230 | overlap |
| ENSMUSG00000041729 | 62536278 | 62536728 | 1.98 | 766 | inside |
| ENSMUSG00000000168 | 50698730 | 50699330 | 1.98 | -38950 | intergenic |
| ENSMUSG00000074505 | 16501541 | 16501991 | 1.98 | -123310 | intergenic |
| ENSMUSG00000026511 | 182124763 | 182125163 | 1.97 | 26 | inside |
| ENSMUSG00000093782 | 82838952 | 82839452 | 1.97 | -494 | overlap |
| ENSMUSG00000090252 | 85254669 | 85255069 | 1.97 | 7818 | inside |
| ENSMUSG00000040596 | 166409752 | 166410152 | 1.97 | 111 | overlap |
| ENSMUSG00000093064 | 190651000 | 190651500 | 1.97 | -167843 | intergenic |
| ENSMUSG00000084849 | 81464383 | 81464783 | 1.97 | 3224 | downstream |
| ENSMUSG00000025795 | 121475673 | 121476323 | 1.97 | 577 | overlap |
| ENSMUSG00000043670 | 81030834 | 81031334 | 1.97 | -5172 | intergenic |
| ENSMUSG00000039232 | 12964195 | 12964595 | 1.97 | 64 | overlap |
| ENSMUSG00000043822 | 80348372 | 80348822 | 1.97 | 40 | overlap |
| ENSMUSG00000020329 | 79735606 | 79736006 | 1.97 | 10975 | downstream |
| ENSMUSG00000013858 | 79977306 | 79977906 | 1.97 | 7024 | inside |
| ENSMUSG00000020277 | 78009634 | 78010134 | 1.97 | 449 | overlap |
| ENSMUSG00000083844 | 5761978 | 5762378 | 1.97 | -151 | overlap |
| ENSMUSG00000049800 | 20542985 | 20543435 | 1.97 | -268 | overlap |
| ENSMUSG00000080152 | 58961780 | 58962180 | 1.97 | -32 | overlap |
| ENSMUSG00000001510 | 95120065 | 95120615 | 1.97 | -24 | overlap |
| ENSMUSG00000000093 | 85838229 | 85838679 | 1.97 | 5678 | inside |
| ENSMUSG00000038255 | 98327818 | 98328268 | 1.97 | 1830 | inside |
| ENSMUSG00000020773 | 116109674 | 116110324 | 1.97 | 17536 | inside |
| ENSMUSG00000072963 | 53456736 | 53457236 | 1.97 | 374 | overlap |
| ENSMUSG00000025582 | 119546868 | 119547368 | 1.97 | 885 | inside |
| ENSMUSG00000018363 | 106919574 | 106920224 | 1.97 | 1141 | inside |


| ENSMUSG00000053113 | 117949667 | 117950117 | 1.97 | 20380 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000085207 | 119942118 | 119943368 | 1.97 | 580 | overlap |
| ENSMUSG00000009079 | 5099128 | 5099878 | 1.97 | 138 | overlap |
| ENSMUSG00000043448 | 59176630 | 59177030 | 1.97 | 6583 | inside |
| ENSMUSG00000020722 | 107686515 | 107686965 | 1.97 | 30007 | intergenic |
| ENSMUSG00000064616 | 48887518 | 48887918 | 1.97 | -122365 | intergenic |
| ENSMUSG00000021108 | 73584111 | 73584611 | 1.97 | -686 | upstream |
| ENSMUSG00000021025 | 55492058 | 55492458 | 1.97 | 589 | inside |
| ENSMUSG00000034168 | 86883327 | 86883877 | 1.97 | 1487 | inside |
| ENSMUSG00000096107 | 3358394 | 3359094 | 1.97 | -2635 | upstream |
| ENSMUSG00000021763 | 113293348 | 113293748 | 1.97 | -23736 | intergenic |
| ENSMUSG00000098266 | 31626071 | 31627021 | 1.97 | 472 | includeFeature |
| ENSMUSG00000088201 | 36210491 | 36210891 | 1.97 | 5231 | intergenic |
| ENSMUSG00000099013 | 66057284 | 66057734 | 1.97 | -17268 | intergenic |
| ENSMUSG00000038879 | 34678396 | 34678796 | 1.97 | 310 | overlap |
| ENSMUSG00000023169 | 96641220 | 96641770 | 1.97 | 1693 | inside |
| ENSMUSG00000044442 | 87353978 | 87354428 | 1.97 | -207 | overlap |
| ENSMUSG00000022702 | 18877321 | 18878071 | 1.97 | 284 | inside |
| ENSMUSG00000009097 | 18582510 | 18582910 | 1.97 | 4459 | inside |
| ENSMUSG00000024036 | 31295578 | 31296478 | 1.97 | 95 | inside |
| ENSMUSG00000024462 | 37045715 | 37046465 | 1.97 | -251 | overlap |
| ENSMUSG00000002249 | 28340762 | 28341262 | 1.97 | 10043 | inside |
| ENSMUSG00000092564 | 33909078 | 33909778 | 1.97 | 239 | overlap |
| ENSMUSG00000024218 | 27909201 | 27909601 | 1.97 | 41 | overlap |
| ENSMUSG00000032688 | 65430677 | 65431177 | 1.97 | -286 | overlap |
| ENSMUSG00000024535 | 53245409 | 53246209 | 1.97 | -253 | overlap |
| ENSMUSG00000024276 | 23954336 | 23954736 | 1.97 | -352 | overlap |
| ENSMUSG00000025429 | 77794013 | 77794463 | 1.97 | -532 | upstream |
| ENSMUSG00000038299 | 32836659 | 32837409 | 1.97 | -566 | overlap |
| ENSMUSG00000025184 | 42518320 | 42518820 | 1.97 | -439 | overlap |
| ENSMUSG00000027605 | 155517673 | 155518173 | 1.97 | -275 | overlap |
| ENSMUSG00000089762 | 30472545 | 30473045 | 1.97 | 1654 | overlapEnd |
| ENSMUSG00000075415 | 31141945 | 31142345 | 1.97 | 63 | overlap |
| ENSMUSG00000098944 | 65659367 | 65659817 | 1.97 | 79 | overlapEnd |
| ENSMUSG00000004894 | 88022545 | 88023245 | 1.97 | 5038 | inside |
| ENSMUSG00000086556 | 96219944 | 96220744 | 1.97 | 224 | overlap |
| ENSMUSG00000027599 | 19073417 | 19074067 | 1.97 | 89648 | intergenic |
| ENSMUSG00000082368 | 70117114 | 70118164 | 1.97 | 69957 | intergenic |
| ENSMUSG00000045751 | 24496115 | 24496615 | 1.97 | -336 | overlap |
| ENSMUSG00000092812 | 134468647 | 134469097 | 1.97 | 283 | downstream |
| ENSMUSG00000028879 | 132884165 | 132884565 | 1.97 | 344 | overlap |
| ENSMUSG00000028367 | 57955826 | 57956226 | 1.97 | 585 | inside |
| ENSMUSG00000003644 | 133887859 | 133888259 | 1.97 | -62 | upstream |
| ENSMUSG00000059816 | 108383037 | 108383587 | 1.97 | 312 | overlap |
| ENSMUSG00000029233 | 76140382 | 76140782 | 1.97 | 111 | inside |
| ENSMUSG00000029474 | 122849976 | 122850626 | 1.97 | -212 | overlap |
| ENSMUSG00000034462 | 104460032 | 104460432 | 1.97 | 582 | inside |
| ENSMUSG00000048988 | 139907897 | 139908597 | 1.97 | -46 | overlap |
| ENSMUSG00000035187 | 101664388 | 101664838 | 1.97 | 838 | inside |
| ENSMUSG00000097191 | 149183709 | 149184259 | 1.97 | 354 | overlap |
| ENSMUSG00000048578 | 115158220 | 115158620 | 1.97 | -41 | upstream |


| ENSMUSG00000097114 | 125493975 | 125494475 | 1.97 | -449 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000005893 | 92091999 | 92092449 | 1.97 | 609 | inside |
| ENSMUSG00000078169 | 53819610 | 53820210 | 1.97 | -365 | includeFeature |
| ENSMUSG00000087136 | 82161399 | 82161899 | 1.97 | -108818 | intergenic |
| ENSMUSG00000040797 | 121413175 | 121413675 | 1.97 | 60503 | inside |
| ENSMUSG00000038784 | 35133591 | 35134091 | 1.97 | 146 | overlap |
| ENSMUSG00000048022 | 24955633 | 24956083 | 1.97 | 492 | inside |
| ENSMUSG00000030057 | 87850689 | 87851089 | 1.97 | 417 | inside |
| ENSMUSG00000030471 | 48744412 | 48745012 | 1.97 | -44591 | intergenic |
| ENSMUSG00000043671 | 35753663 | 35754163 | 1.97 | 791 | inside |
| ENSMUSG00000031783 | 94857430 | 94857930 | 1.97 | -20 | overlap |
| ENSMUSG00000031860 | 69832659 | 69833059 | 1.97 | 35 | inside |
| ENSMUSG00000096943 | 83875181 | 83875581 | 1.97 | 24320 | intergenic |
| ENSMUSG00000098976 | 71629286 | 71629686 | 1.97 | 1820 | downstream |
| ENSMUSG00000031880 | 104630658 | 104631108 | 1.97 | 663 | inside |
| ENSMUSG00000069633 | 3451604 | 3452054 | 1.97 | 16076 | intergenic |
| ENSMUSG00000071793 | 20424102 | 20424552 | 1.97 | 712 | inside |
| ENSMUSG00000055435 | 115706439 | 115707189 | 1.97 | 1355 | inside |
| ENSMUSG00000025647 | 109036271 | 109037171 | 1.97 | -2294 | upstream |
| ENSMUSG00000041268 | 54500278 | 54501478 | 1.97 | 1482 | inside |
| ENSMUSG00000045414 | 94537566 | 94537966 | 1.97 | 515 | inside |
| ENSMUSG00000033688 | 103304805 | 103305355 | 1.97 | 277 | overlap |
| ENSMUSG00000045087 | 21244591 | 21245191 | 1.97 | 3852 | inside |
| ENSMUSG00000032405 | 62980129 | 62980579 | 1.97 | 750 | inside |
| ENSMUSG00000026585 | 163779608 | 163780158 | 1.96 | 25 | inside |
| ENSMUSG00000089358 | 13896958 | 13897408 | 1.96 | -35420 | intergenic |
| ENSMUSG00000046404 | 130717267 | 130717767 | 1.96 | -60 | overlap |
| ENSMUSG00000016528 | 131096811 | 131097261 | 1.96 | 732 | inside |
| ENSMUSG00000025980 | 55088175 | 55088675 | 1.96 | 68 | overlap |
| ENSMUSG00000025956 | 64617614 | 64618014 | 1.96 | -372 | upstream |
| ENSMUSG00000025774 | 18058013 | 18058463 | 1.96 | 87889 | intergenic |
| ENSMUSG00000098811 | 81387634 | 81388034 | 1.96 | 1287 | downstream |
| ENSMUSG00000019820 | 12869303 | 12869703 | 1.96 | -7568 | intergenic |
| ENSMUSG00000020052 | 87492598 | 87492998 | 1.96 | 1062 | inside |
| ENSMUSG00000035781 | 79921163 | 79921713 | 1.96 | -4157 | upstream |
| ENSMUSG00000074734 | 116113402 | 116113852 | 1.96 | 515 | inside |
| ENSMUSG00000075410 | 116653274 | 116653924 | 1.96 | -260 | overlap |
| ENSMUSG00000020788 | 72961277 | 72961677 | 1.96 | 108 | inside |
| ENSMUSG00000038976 | 94991218 | 94991618 | 1.96 | 183 | inside |
| ENSMUSG00000059248 | 117199317 | 117199967 | 1.96 | -344 | overlap |
| ENSMUSG00000063109 | 89060186 | 89060836 | 1.96 | -4543 | upstream |
| ENSMUSG00000083555 | 20631829 | 20632229 | 1.96 | -56640 | intergenic |
| ENSMUSG00000078154 | 48837777 | 48838177 | 1.96 | -11122 | intergenic |
| ENSMUSG00000093164 | 74610336 | 74610886 | 1.96 | 8177 | intergenic |
| ENSMUSG00000025576 | 118908465 | 118908865 | 1.96 | 3132 | inside |
| ENSMUSG00000018931 | 60912980 | 60913630 | 1.96 | 1770 | inside |
| ENSMUSG00000001440 | 97187867 | 97188367 | 1.96 | 14 | overlap |
| ENSMUSG00000070407 | 63921127 | 63921527 | 1.96 | 1163 | inside |
| ENSMUSG00000054204 | 30884362 | 30884762 | 1.96 | 40 | inside |
| ENSMUSG00000035451 | 57542861 | 57543261 | 1.96 | 3260 | inside |
| ENSMUSG00000021279 | 111377687 | 111378137 | 1.96 | 31 | overlap |


| ENSMUSG00000020949 | 65073811 | 65074311 | 1.96 | 133 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000021120 | 79089358 | 79089758 | 1.96 | 312 | overlap |
| ENSMUSG00000050545 | 4768549 | 4768999 | 1.96 | 718 | inside |
| ENSMUSG00000021701 | 110394258 | 110394658 | 1.96 | -786 | upstream |
| ENSMUSG00000087143 | 78163205 | 78163805 | 1.96 | -34812 | intergenic |
| ENSMUSG00000021552 | 58273405 | 58273855 | 1.96 | 783 | inside |
| ENSMUSG00000041417 | 101767226 | 101768526 | 1.96 | 991 | overlap |
| ENSMUSG00000017485 | 16364513 | 16365363 | 1.96 | -666 | overlap |
| ENSMUSG00000063458 | 22019355 | 22019755 | 1.96 | -357 | overlap |
| ENSMUSG00000021771 | 21830805 | 21831255 | 1.96 | -464 | upstream |
| ENSMUSG00000095311 | 122406690 | 122407140 | 1.96 | -18530 | intergenic |
| ENSMUSG00000022456 | 82274966 | 82275716 | 1.96 | 31 | inside |
| ENSMUSG00000022622 | 89548019 | 89548669 | 1.96 | -20307 | intergenic |
| ENSMUSG00000064210 | 95790715 | 95791115 | 1.96 | -128 | overlap |
| ENSMUSG00000091198 | 81523039 | 81523439 | 1.96 | -23491 | intergenic |
| ENSMUSG00000038965 | 17200610 | 17201060 | 1.96 | 2039 | inside |
| ENSMUSG00000069729 | 4908157 | 4908557 | 1.96 | -86175 | intergenic |
| ENSMUSG00000024027 | 30901322 | 30901772 | 1.96 | -545 | upstream |
| ENSMUSG00000002372 | 56673073 | 56673573 | 1.96 | -152 | overlap |
| ENSMUSG00000034868 | 70990183 | 70990583 | 1.96 | 604 | inside |
| ENSMUSG00000024805 | 36347826 | 36348276 | 1.96 | -31241 | intergenic |
| ENSMUSG00000065454 | 180893966 | 180894766 | 1.96 | -65 | includeFeature |
| ENSMUSG00000026781 | 23102751 | 23103151 | 1.96 | 34583 | inside |
| ENSMUSG00000027273 | 136678723 | 136679173 | 1.96 | -34730 | intergenic |
| ENSMUSG00000027422 | 144011162 | 144011562 | 1.96 | 101 | overlap |
| ENSMUSG00000027566 | 180041834 | 180042284 | 1.96 | 599 | inside |
| ENSMUSG00000027634 | 156991499 | 156991899 | 1.96 | 557 | inside |
| ENSMUSG00000028152 | 138741575 | 138742025 | 1.96 | -620 | upstream |
| ENSMUSG00000028086 | 84814945 | 84815345 | 1.96 | -323 | overlap |
| ENSMUSG00000028060 | 88685803 | 88686703 | 1.96 | 9 | inside |
| ENSMUSG00000042579 | 89772896 | 89773946 | 1.96 | 344 | overlap |
| ENSMUSG00000045031 | 37312590 | 37312990 | 1.96 | 118 | overlap |
| ENSMUSG00000027793 | 55055203 | 55055653 | 1.96 | -148 | upstream |
| ENSMUSG00000008763 | 100685316 | 100685716 | 1.96 | 187 | overlap |
| ENSMUSG00000005813 | 138488603 | 138489053 | 1.96 | 779 | inside |
| ENSMUSG00000028709 | 115827887 | 115828287 | 1.96 | -205 | overlap |
| ENSMUSG00000034210 | 115737217 | 115737867 | 1.96 | -527 | overlap |
| ENSMUSG00000081501 | 88757126 | 88757626 | 1.96 | -718 | upstream |
| ENSMUSG00000083154 | 24289745 | 24290345 | 1.96 | 51645 | intergenic |
| ENSMUSG00000029050 | 155221489 | 155221889 | 1.96 | 1103 | inside |
| ENSMUSG00000083743 | 84545094 | 84545544 | 1.96 | -115970 | intergenic |
| ENSMUSG00000028826 | 134853177 | 134853777 | 1.96 | 168 | overlap |
| ENSMUSG00000028337 | 46601958 | 46602458 | 1.96 | 244 | overlap |
| ENSMUSG00000028849 | 126237030 | 126237430 | 1.96 | 19313 | inside |
| ENSMUSG00000048988 | 139912625 | 139913075 | 1.96 | 4682 | inside |
| ENSMUSG00000043833 | 25100825 | 25101225 | 1.96 | -158 | overlap |
| ENSMUSG00000029359 | 118028442 | 118028842 | 1.96 | 699 | inside |
| ENSMUSG00000044134 | 121852220 | 121852720 | 1.96 | 3192 | inside |
| ENSMUSG00000072754 | 110363174 | 110363574 | 1.96 | 9078 | intergenic |
| ENSMUSG00000036377 | 76657782 | 76658182 | 1.96 | 99 | inside |
| ENSMUSG00000048988 | 139908597 | 139909147 | 1.96 | 654 | inside |


| ENSMUSG00000029405 | 92082875 | 92083275 | 1.96 | 860 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000001566 | 29477189 | 29477789 | 1.96 | 1330 | inside |
| ENSMUSG00000029823 | 38550993 | 38551543 | 1.96 | -341 | overlap |
| ENSMUSG00000068551 | 48437905 | 48438305 | 1.96 | 7920 | inside |
| ENSMUSG00000050654 | 116578665 | 116579065 | 1.96 | 4366 | downstream |
| ENSMUSG00000093052 | 90716289 | 90716789 | 1.96 | -34769 | intergenic |
| ENSMUSG00000038077 | 126740215 | 126740765 | 1.96 | 447 | overlap |
| ENSMUSG00000053442 | 66896639 | 66897089 | 1.96 | -118 | upstream |
| ENSMUSG00000055409 | 50175093 | 50176443 | 1.96 | 200229 | inside |
| ENSMUSG00000093238 | 79534987 | 79535587 | 1.96 | 29723 | intergenic |
| ENSMUSG00000092381 | 19714614 | 19715114 | 1.96 | -402 | overlap |
| ENSMUSG00000002083 | 16312314 | 16312714 | 1.96 | 2698 | inside |
| ENSMUSG00000046058 | 28267394 | 28267894 | 1.96 | -487 | overlap |
| ENSMUSG00000025584 | 81213624 | 81214024 | 1.96 | 28 | inside |
| ENSMUSG00000047085 | 44444612 | 44445112 | 1.96 | 15594 | inside |
| ENSMUSG00000032875 | 100930849 | 100931249 | 1.96 | 1312 | inside |
| ENSMUSG00000003872 | 45369803 | 45370403 | 1.96 | 780 | inside |
| ENSMUSG00000034656 | 84414409 | 84415009 | 1.96 | 25969 | inside |
| ENSMUSG00000036442 | 105854958 | 105855458 | 1.96 | -145 | overlap |
| ENSMUSG00000025521 | 64947286 | 64947736 | 1.96 | 101 | inside |
| ENSMUSG00000049717 | 9977472 | 9977872 | 1.96 | 214 | overlap |
| ENSMUSG00000061313 | 25753667 | 25754117 | 1.96 | 613 | inside |
| ENSMUSG00000025044 | 39905168 | 39905768 | 1.96 | -262490 | intergenic |
| ENSMUSG00000032012 | 43744252 | 43744702 | 1.96 | -324 | overlap |
| ENSMUSG00000036867 | 64020278 | 64020678 | 1.96 | 1781 | inside |
| ENSMUSG00000097083 | 39650274 | 39650674 | 1.95 | 98 | inside |
| ENSMUSG00000026455 | 134455492 | 134455942 | 1.95 | -39 | overlap |
| ENSMUSG00000034353 | 91179962 | 91180462 | 1.95 | 140 | inside |
| ENSMUSG00000005886 | 13374096 | 13374746 | 1.95 | -13 | upstream |
| ENSMUSG00000037503 | 34801682 | 34802132 | 1.95 | 41383 | intergenic |
| ENSMUSG00000026670 | 170174596 | 170174996 | 1.95 | 361 | overlap |
| ENSMUSG00000040675 | 3973047 | 3973447 | 1.95 | -71 | overlap |
| ENSMUSG00000009092 | 75893113 | 75893513 | 1.95 | -285 | overlap |
| ENSMUSG00000020183 | 117629095 | 117629495 | 1.95 | -405 | upstream |
| ENSMUSG00000020185 | 110745254 | 110745704 | 1.95 | -185 | overlap |
| ENSMUSG00000020114 | 119239361 | 119239761 | 1.95 | 694 | inside |
| ENSMUSG00000062866 | 13402145 | 13402695 | 1.95 | 72251 | inside |
| ENSMUSG00000020402 | 52361480 | 52361880 | 1.95 | 620 | inside |
| ENSMUSG00000055805 | 103170574 | 103171324 | 1.95 | -533 | overlap |
| ENSMUSG00000050830 | 11113526 | 11114176 | 1.95 | -697 | upstream |
| ENSMUSG00000020869 | 94629776 | 94630176 | 1.95 | 9 | inside |
| ENSMUSG00000020721 | 107547676 | 107548276 | 1.95 | -254 | overlap |
| ENSMUSG00000034449 | 84828527 | 84829027 | 1.95 | 467 | overlap |
| ENSMUSG00000048732 | 100471924 | 100472624 | 1.95 | 817 | inside |
| ENSMUSG00000000120 | 95587286 | 95587686 | 1.95 | 449 | inside |
| ENSMUSG00000054517 | 116130115 | 116130665 | 1.95 | 1013 | inside |
| ENSMUSG00000045546 | 84416898 | 84417298 | 1.95 | -296 | overlap |
| ENSMUSG00000071379 | 17691098 | 17691498 | 1.95 | 284 | inside |
| ENSMUSG00000085622 | 71015740 | 71016340 | 1.95 | 92 | overlap |
| ENSMUSG00000021240 | 84616927 | 84617327 | 1.95 | 539 | inside |
| ENSMUSG00000002997 | 32060630 | 32061030 | 1.95 | 666 | inside |


| ENSMUSG00000099100 | 67221557 | 67222157 | 1.95 | -308 | upstream |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000005320 | 55152679 | 55153479 | 1.95 | 39 | inside |
| ENSMUSG00000077380 | 95335811 | 95336361 | 1.95 | 3193 | downstream |
| ENSMUSG00000034575 | 69534937 | 69535337 | 1.95 | -1098 | upstream |
| ENSMUSG00000021838 | 46883455 | 46883855 | 1.95 | 601 | inside |
| ENSMUSG00000048279 | 61173175 | 61173575 | 1.95 | 34718 | inside |
| ENSMUSG00000048582 | 57057193 | 57057593 | 1.95 | 837 | inside |
| ENSMUSG00000022203 | 54925643 | 54926043 | 1.95 | 1145 | inside |
| ENSMUSG00000022994 | 98609565 | 98610165 | 1.95 | -1932 | upstream |
| ENSMUSG00000037465 | 38301050 | 38301500 | 1.95 | -343 | upstream |
| ENSMUSG00000033653 | 21423350 | 21423750 | 1.95 | 232 | inside |
| ENSMUSG00000042644 | 27057315 | 27057765 | 1.95 | 11 | inside |
| ENSMUSG00000086534 | 29841898 | 29842298 | 1.95 | -48186 | intergenic |
| ENSMUSG00000005370 | 87974856 | 87975256 | 1.95 | -194 | overlap |
| ENSMUSG00000024335 | 34120462 | 34121112 | 1.95 | 2172 | inside |
| ENSMUSG00000024241 | 80480303 | 80480703 | 1.95 | 150 | overlap |
| ENSMUSG00000035765 | 75018433 | 75019033 | 1.95 | -339 | overlap |
| ENSMUSG00000024576 | 61555033 | 61555483 | 1.95 | -241 | overlap |
| ENSMUSG00000049173 | 60596288 | 60596738 | 1.95 | -4500 | upstream |
| ENSMUSG00000033382 | 20895612 | 20896062 | 1.95 | 466 | inside |
| ENSMUSG00000033768 | 6418710 | 6419110 | 1.95 | -21 | overlap |
| ENSMUSG00000025231 | 46397020 | 46397470 | 1.95 | 124 | inside |
| ENSMUSG00000004085 | 72286014 | 72286414 | 1.95 | 377 | inside |
| ENSMUSG00000068267 | 131179390 | 131179840 | 1.95 | 622 | inside |
| ENSMUSG00000038860 | 33129715 | 33130415 | 1.95 | 1939 | inside |
| ENSMUSG00000040016 | 157566804 | 157567404 | 1.95 | -88 | overlap |
| ENSMUSG00000086181 | 39099567 | 39100917 | 1.95 | -211308 | intergenic |
| ENSMUSG00000090817 | 98428089 | 98428589 | 1.95 | 28942 | intergenic |
| ENSMUSG00000061175 | 79567740 | 79568140 | 1.95 | 56 | overlap |
| ENSMUSG00000028081 | 86142317 | 86143317 | 1.95 | 385 | overlap |
| ENSMUSG00000080771 | 30069472 | 30070322 | 1.95 | -124175 | intergenic |
| ENSMUSG00000038024 | 86748270 | 86748670 | 1.95 | -285 | overlap |
| ENSMUSG00000028876 | 124885530 | 124885980 | 1.95 | 4631 | inside |
| ENSMUSG00000034926 | 106560543 | 106560993 | 1.95 | -495 | upstream |
| ENSMUSG00000084869 | 132048259 | 132048659 | 1.95 | 81 | inside |
| ENSMUSG00000028635 | 120148337 | 120148837 | 1.95 | -12869 | intergenic |
| ENSMUSG00000078582 | 119814575 | 119815075 | 1.95 | 80779 | intergenic |
| ENSMUSG00000029056 | 154964263 | 154964813 | 1.95 | 140 | inside |
| ENSMUSG00000051279 | 9843625 | 9844125 | 1.95 | -747 | upstream |
| ENSMUSG00000040761 | 141537509 | 141537909 | 1.95 | 1088 | inside |
| ENSMUSG00000036087 | 72913812 | 72914262 | 1.95 | -492 | upstream |
| ENSMUSG00000029578 | 142629597 | 142630047 | 1.95 | 55 | inside |
| ENSMUSG00000015880 | 45669568 | 45670518 | 1.95 | -354 | overlap |
| ENSMUSG00000089809 | 99728588 | 99728988 | 1.95 | 477 | inside |
| ENSMUSG00000061898 | 143180522 | 143180922 | 1.95 | 253 | overlap |
| ENSMUSG00000029467 | 122502520 | 122502920 | 1.95 | -295 | upstream |
| ENSMUSG00000052751 | 48601534 | 48601984 | 1.95 | 7651 | downstream |
| ENSMUSG00000030059 | 97178499 | 97178949 | 1.95 | 625 | inside |
| ENSMUSG00000052512 | 49246056 | 49247006 | 1.95 | 337340 | inside |
| ENSMUSG00000060621 | 19523433 | 19524083 | 1.95 | 4702 | inside |
| ENSMUSG00000040177 | 131362009 | 131362459 | 1.95 | 689 | inside |


| ENSMUSG00000036459 | 34133213 | 34133613 | 1.95 | 55 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000054808 | 28961644 | 28962194 | 1.95 | 696 | inside |
| ENSMUSG00000030850 | 130519382 | 130519782 | 1.95 | 579 | inside |
| ENSMUSG00000030660 | 116443349 | 116443749 | 1.95 | 109 | overlap |
| ENSMUSG00000031004 | 135715990 | 135716390 | 1.95 | 389 | overlap |
| ENSMUSG00000051550 | 4993961 | 4994511 | 1.95 | 2197 | inside |
| ENSMUSG00000038502 | 44868803 | 44869303 | 1.95 | 985 | inside |
| ENSMUSG00000076144 | 84084609 | 84085059 | 1.95 | -1490 | upstream |
| ENSMUSG00000074247 | 71468673 | 71469123 | 1.95 | -526 | upstream |
| ENSMUSG00000087408 | 70329981 | 70331331 | 1.95 | 14194 | inside |
| ENSMUSG00000031511 | 11758058 | 11758608 | 1.95 | 29953 | inside |
| ENSMUSG00000031447 | 13159480 | 13159880 | 1.95 | 345 | inside |
| ENSMUSG00000046413 | 91800423 | 91801173 | 1.95 | 8 | inside |
| ENSMUSG00000031799 | 72135190 | 72135590 | 1.95 | 215 | inside |
| ENSMUSG00000031486 | 27085917 | 27086417 | 1.95 | 334 | inside |
| ENSMUSG00000086067 | 84559259 | 84559959 | 1.95 | 57925 | intergenic |
| ENSMUSG00000000743 | 123212422 | 123212922 | 1.95 | 341 | overlap |
| ENSMUSG00000038542 | 13104530 | 13104980 | 1.95 | 929 | inside |
| ENSMUSG00000053716 | 106368254 | 106368854 | 1.95 | -378 | overlap |
| ENSMUSG00000032435 | 114688767 | 114689167 | 1.95 | -23 | overlap |
| ENSMUSG00000054693 | 70678728 | 70679128 | 1.95 | -269 | overlap |
| ENSMUSG00000045414 | 94538766 | 94539266 | 1.95 | -685 | upstream |
| ENSMUSG00000066456 | 83145628 | 83146028 | 1.95 | 979 | inside |
| ENSMUSG00000032018 | 42264002 | 42264552 | 1.95 | 298 | overlap |
| ENSMUSG00000032265 | 85327628 | 85328178 | 1.95 | -504 | upstream |
| ENSMUSG00000031935 | 15279241 | 15279741 | 1.95 | 626 | inside |
| ENSMUSG00000053286 | 151428489 | 151429089 | 1.94 | -159 | overlap |
| ENSMUSG00000041642 | 136131556 | 136131956 | 1.94 | 167 | inside |
| ENSMUSG00000004768 | 33719388 | 33719938 | 1.94 | -494 | overlap |
| ENSMUSG00000089418 | 141599311 | 141599711 | 1.94 | 284956 | intergenic |
| ENSMUSG00000098273 | 118837742 | 118838242 | 1.94 | -116478 | intergenic |
| ENSMUSG00000026584 | 163929863 | 163930263 | 1.94 | 763 | inside |
| ENSMUSG00000077237 | 108327312 | 108327912 | 1.94 | 434338 | intergenic |
| ENSMUSG00000034066 | 93512111 | 93512561 | 1.94 | 32 | inside |
| ENSMUSG00000041570 | 136345461 | 136345861 | 1.94 | 643 | inside |
| ENSMUSG00000018199 | 143776764 | 143777614 | 1.94 | 304 | overlap |
| ENSMUSG00000077608 | 163049358 | 163050458 | 1.94 | 5593 | downstream |
| ENSMUSG00000035754 | 79960265 | 79960665 | 1.94 | 113 | inside |
| ENSMUSG00000020042 | 85387534 | 85388034 | 1.94 | 720 | inside |
| ENSMUSG00000025422 | 127090895 | 127091345 | 1.94 | 11988 | inside |
| ENSMUSG00000071359 | 22731745 | 22732145 | 1.94 | 193 | overlap |
| ENSMUSG00000001120 | 76960648 | 76961948 | 1.94 | 1239 | overlap |
| ENSMUSG00000047712 | 8518303 | 8518703 | 1.94 | 522 | inside |
| ENSMUSG00000051043 | 114851526 | 114852226 | 1.94 | 374 | inside |
| ENSMUSG00000020849 | 75733030 | 75733530 | 1.94 | 161 | inside |
| ENSMUSG00000038976 | 94992018 | 94992418 | 1.94 | 983 | inside |
| ENSMUSG00000020700 | 106084986 | 106085386 | 1.94 | 373 | inside |
| ENSMUSG00000087013 | 85831730 | 85832230 | 1.94 | 493 | overlap |
| ENSMUSG00000043099 | 75168127 | 75168577 | 1.94 | 1392 | inside |
| ENSMUSG00000041654 | 113649874 | 113650274 | 1.94 | 205 | overlap |
| ENSMUSG00000034520 | 102818215 | 102818715 | 1.94 | 1485 | inside |


| ENSMUSG00000049659 | 20741376 | 20741926 | 1.94 | 213 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000048562 | 118848704 | 118849104 | 1.94 | 2375 | inside |
| ENSMUSG00000020647 | 4476770 | 4477220 | 1.94 | 412 | overlap |
| ENSMUSG00000021054 | 75735457 | 75736007 | 1.94 | 272 | overlap |
| ENSMUSG00000038683 | 41000147 | 41001347 | 1.94 | -876 | overlap |
| ENSMUSG00000052293 | 100651514 | 100651964 | 1.94 | 171 | inside |
| ENSMUSG00000021719 | 105053786 | 105054236 | 1.94 | 1144 | inside |
| ENSMUSG00000038991 | 38527747 | 38528147 | 1.94 | 1077 | inside |
| ENSMUSG00000036282 | 49172705 | 49173155 | 1.94 | 479 | inside |
| ENSMUSG00000021994 | 28505727 | 28506127 | 1.94 | 977 | inside |
| ENSMUSG00000021775 | 18238849 | 18239249 | 1.94 | 278 | overlap |
| ENSMUSG00000047888 | 80711121 | 80711571 | 1.94 | -198 | overlap |
| ENSMUSG00000097938 | 54147141 | 54147641 | 1.94 | -111159 | intergenic |
| ENSMUSG00000001281 | 102224222 | 102224672 | 1.94 | 7713 | inside |
| ENSMUSG00000097875 | 38989755 | 38990405 | 1.94 | 16430 | inside |
| ENSMUSG00000077468 | 31053996 | 31054496 | 1.94 | 140342 | intergenic |
| ENSMUSG00000071632 | 18836329 | 18836879 | 1.94 | -249 | overlap |
| ENSMUSG00000041215 | 20141249 | 20141649 | 1.94 | 186 | inside |
| ENSMUSG00000040605 | 97356757 | 97357157 | 1.94 | 29 | inside |
| ENSMUSG00000022811 | 33381178 | 33381578 | 1.94 | 403 | inside |
| ENSMUSG00000022799 | 38712740 | 38713340 | 1.94 | 534 | overlap |
| ENSMUSG00000022974 | 91043878 | 91044278 | 1.94 | 665 | inside |
| ENSMUSG00000005580 | 4418945 | 4419995 | 1.94 | 1553 | inside |
| ENSMUSG00000079507 | 35320115 | 35320965 | 1.94 | -290 | overlap |
| ENSMUSG00000024317 | 21002141 | 21002541 | 1.94 | 800 | inside |
| ENSMUSG00000088493 | 37722209 | 37722659 | 1.94 | 16230 | intergenic |
| ENSMUSG00000048799 | 53744409 | 53744809 | 1.94 | 138 | overlap |
| ENSMUSG00000024892 | 4509560 | 4510460 | 1.94 | -912 | upstream |
| ENSMUSG00000006463 | 4878660 | 4879210 | 1.94 | -8 | overlap |
| ENSMUSG00000052188 | 6498003 | 6498903 | 1.94 | 34008 | intergenic |
| ENSMUSG00000045045 | 4615689 | 4616089 | 1.94 | -22 | upstream |
| ENSMUSG00000087182 | 18693988 | 18694438 | 1.94 | -44 | overlap |
| ENSMUSG00000057914 | 14559332 | 14560132 | 1.94 | -44974 | intergenic |
| ENSMUSG00000085946 | 152048690 | 152049090 | 1.94 | 28118 | intergenic |
| ENSMUSG00000027238 | 121806762 | 121807162 | 1.94 | 325 | overlap |
| ENSMUSG00000027404 | 130178685 | 130179085 | 1.94 | 718 | inside |
| ENSMUSG00000065485 | 29845769 | 29846219 | 1.94 | -42 | upstream |
| ENSMUSG00000078137 | 118703029 | 118703829 | 1.94 | 934 | inside |
| ENSMUSG00000026915 | 37703489 | 37703939 | 1.94 | 370 | overlap |
| ENSMUSG00000081056 | 119872699 | 119873199 | 1.94 | 5704 | downstream |
| ENSMUSG00000015335 | 30093365 | 30093765 | 1.94 | 283 | overlap |
| ENSMUSG00000026805 | 28916119 | 28916619 | 1.94 | 549 | inside |
| ENSMUSG00000089444 | 81597196 | 81598046 | 1.94 | -283531 | intergenic |
| ENSMUSG00000028271 | 142764804 | 142765304 | 1.94 | -422 | overlap |
| ENSMUSG00000098515 | 129213499 | 129214199 | 1.94 | 3493 | downstream |
| ENSMUSG00000004891 | 87971095 | 87971545 | 1.94 | 17 | inside |
| ENSMUSG00000089149 | 19335575 | 19336175 | 1.94 | 4802 | downstream |
| ENSMUSG00000014599 | 107759744 | 107760294 | 1.94 | 725 | inside |
| ENSMUSG00000089027 | 120740771 | 120742071 | 1.94 | -130678 | intergenic |
| ENSMUSG00000028070 | 88058367 | 88058767 | 1.94 | 128 | overlap |
| ENSMUSG00000027947 | 89913045 | 89913445 | 1.94 | 117 | overlap |


| ENSMUSG00000027840 | 104961021 | 104961471 | 1.94 | 688 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000068696 | 116251871 | 116252271 | 1.94 | 1613 | inside |
| ENSMUSG00000061143 | 52101867 | 52102517 | 1.94 | 3024 | inside |
| ENSMUSG00000028488 | 85204894 | 85205344 | 1.94 | -232 | overlap |
| ENSMUSG00000006445 | 141300877 | 141301377 | 1.94 | -363 | overlap |
| ENSMUSG00000039852 | 150281833 | 150282383 | 1.94 | 187 | inside |
| ENSMUSG00000024793 | 152131413 | 152132063 | 1.94 | 15479 | intergenic |
| ENSMUSG00000029049 | 155150589 | 155151089 | 1.94 | 64012 | intergenic |
| ENSMUSG00000028476 | 43874996 | 43875546 | 1.94 | -534 | overlap |
| ENSMUSG00000028664 | 136836383 | 136836983 | 1.94 | -395 | upstream |
| ENSMUSG00000028943 | 152152113 | 152152563 | 1.94 | 258 | overlap |
| ENSMUSG00000028842 | 126429618 | 126430018 | 1.94 | -62 | upstream |
| ENSMUSG00000028551 | 109666375 | 109666975 | 1.94 | 814 | inside |
| ENSMUSG00000042500 | 126532804 | 126533204 | 1.94 | 668 | inside |
| ENSMUSG00000029345 | 112307687 | 112308087 | 1.94 | -18671 | intergenic |
| ENSMUSG00000075703 | 30231995 | 30232595 | 1.94 | -586 | overlap |
| ENSMUSG00000037373 | 33274228 | 33274628 | 1.94 | 776 | inside |
| ENSMUSG00000036285 | 77309232 | 77309782 | 1.94 | 852 | inside |
| ENSMUSG00000053121 | 29735767 | 29736167 | 1.94 | 169 | overlap |
| ENSMUSG00000029832 | 51421834 | 51422384 | 1.94 | -10836 | intergenic |
| ENSMUSG00000059187 | 95951847 | 95952347 | 1.94 | -161307 | intergenic |
| ENSMUSG00000000182 | 127080242 | 127080692 | 1.94 | 7340 | inside |
| ENSMUSG00000045095 | 94283049 | 94283599 | 1.94 | 273 | overlap |
| ENSMUSG00000030086 | 89595010 | 89595710 | 1.94 | 642 | overlap |
| ENSMUSG00000030093 | 91410739 | 91411189 | 1.94 | 624 | inside |
| ENSMUSG00000035585 | 3685604 | 3686054 | 1.94 | -7771 | intergenic |
| ENSMUSG00000033967 | 12927833 | 12928233 | 1.94 | 417 | inside |
| ENSMUSG00000053877 | 127512212 | 127512912 | 1.94 | 185 | inside |
| ENSMUSG00000030374 | 16809977 | 16811027 | 1.94 | -5912 | upstream |
| ENSMUSG00000033917 | 118705299 | 118705849 | 1.94 | 479 | overlap |
| ENSMUSG00000003873 | 45466643 | 45467043 | 1.94 | 255 | overlap |
| ENSMUSG00000031548 | 23411286 | 23411686 | 1.94 | -216 | overlap |
| ENSMUSG00000031782 | 94838139 | 94838539 | 1.94 | -182 | overlap |
| ENSMUSG00000055707 | 70476459 | 70476959 | 1.94 | 484 | overlap |
| ENSMUSG00000063049 | 47675635 | 47676035 | 1.94 | -79 | upstream |
| ENSMUSG00000081316 | 37554054 | 37554454 | 1.94 | 234638 | intergenic |
| ENSMUSG00000032185 | 21546141 | 21546591 | 1.94 | -753 | upstream |
| ENSMUSG00000042138 | 37489300 | 37489850 | 1.94 | -21 | overlap |
| ENSMUSG00000098171 | 93594197 | 93594647 | 1.94 | -201827 | intergenic |
| ENSMUSG00000032120 | 44319452 | 44319952 | 1.94 | 830 | inside |
| ENSMUSG00000032244 | 62811678 | 62812078 | 1.94 | -30 | upstream |
| ENSMUSG00000032112 | 44407402 | 44408052 | 1.94 | 146 | overlap |
| ENSMUSG00000032026 | 48479881 | 48480731 | 1.94 | 730 | overlap |
| ENSMUSG00000097334 | 57076828 | 57077278 | 1.94 | -815 | upstream |
| ENSMUSG00000076999 | 105567011 | 105568311 | 1.93 | -7676 | intergenic |
| ENSMUSG00000034220 | 92831106 | 92831806 | 1.93 | -539 | overlap |
| ENSMUSG00000026514 | 181352708 | 181353108 | 1.93 | 80 | inside |
| ENSMUSG00000026239 | 86581869 | 86582419 | 1.93 | 760 | inside |
| ENSMUSG00000055676 | 180332302 | 180332702 | 1.93 | -1752 | upstream |
| ENSMUSG00000066842 | 150993314 | 150993714 | 1.93 | 121 | overlap |
| ENSMUSG00000010609 | 180255646 | 180256146 | 1.93 | 7792 | inside |


| ENSMUSG00000035011 | 81143913 | 81144413 | 1.93 | 8693 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000058297 | 60106653 | 60107053 | 1.93 | 434 | inside |
| ENSMUSG00000058806 | 61978930 | 61979380 | 1.93 | 178 | overlap |
| ENSMUSG00000007850 | 50378379 | 50378829 | 1.93 | 1389 | inside |
| ENSMUSG00000056687 | 109363065 | 109363665 | 1.93 | 589 | overlap |
| ENSMUSG00000046719 | 95514318 | 95514718 | 1.93 | 252 | overlap |
| ENSMUSG00000015837 | 50210486 | 50210886 | 1.93 | 341 | overlap |
| ENSMUSG00000044066 | 20249326 | 20249776 | 1.93 | 103 | overlap |
| ENSMUSG00000020941 | 103267265 | 103267665 | 1.93 | 207 | overlap |
| ENSMUSG00000044548 | 71309211 | 71309711 | 1.93 | -673 | upstream |
| ENSMUSG00000020654 | 4132954 | 4133354 | 1.93 | -149 | overlap |
| ENSMUSG00000021277 | 111166733 | 111167133 | 1.93 | 363 | inside |
| ENSMUSG00000021136 | 81027018 | 81027418 | 1.93 | 210 | inside |
| ENSMUSG00000086022 | 11455689 | 11456189 | 1.93 | -390 | overlap |
| ENSMUSG00000091264 | 41249521 | 41249921 | 1.93 | -339 | overlap |
| ENSMUSG00000000253 | 45507021 | 45507621 | 1.93 | -423 | overlap |
| ENSMUSG00000021488 | 55210005 | 55210455 | 1.93 | 223 | inside |
| ENSMUSG00000076431 | 28951997 | 28952397 | 1.93 | 1716 | inside |
| ENSMUSG00000097775 | 62759175 | 62760025 | 1.93 | -1326 | upstream |
| ENSMUSG00000040760 | 26970499 | 26970899 | 1.93 | 733 | inside |
| ENSMUSG00000006289 | 50924351 | 50924751 | 1.93 | 542 | inside |
| ENSMUSG00000036046 | 84987166 | 84987716 | 1.93 | 1385 | inside |
| ENSMUSG00000032988 | 79252175 | 79252575 | 1.93 | 2586 | inside |
| ENSMUSG00000056258 | 66285589 | 66286539 | 1.93 | 635 | overlap |
| ENSMUSG00000055065 | 79545839 | 79546739 | 1.93 | 902 | inside |
| ENSMUSG00000039830 | 91227277 | 91227677 | 1.93 | 1727 | inside |
| ENSMUSG00000062713 | 94084927 | 94085327 | 1.93 | -333 | overlap |
| ENSMUSG00000039345 | 8470610 | 8471110 | 1.93 | -178 | overlap |
| ENSMUSG00000022858 | 22265250 | 22265650 | 1.93 | 755 | inside |
| ENSMUSG00000022898 | 94526816 | 94527216 | 1.93 | 14 | overlap |
| ENSMUSG00000022892 | 85173519 | 85173919 | 1.93 | 188 | overlap |
| ENSMUSG00000043445 | 24469811 | 24470411 | 1.93 | -662 | upstream |
| ENSMUSG00000002365 | 5840157 | 5841057 | 1.93 | -1171 | upstream |
| ENSMUSG00000061232 | 33999572 | 34000122 | 1.93 | 761 | inside |
| ENSMUSG00000059409 | 46704308 | 46704758 | 1.93 | 694 | inside |
| ENSMUSG00000001525 | 35836765 | 35837215 | 1.93 | 1541 | inside |
| ENSMUSG00000024563 | 76241277 | 76241677 | 1.93 | -303 | overlap |
| ENSMUSG00000034006 | 80256283 | 80256683 | 1.93 | 2991 | inside |
| ENSMUSG00000057506 | 44146126 | 44146526 | 1.93 | 320 | overlap |
| ENSMUSG00000085444 | 110157052 | 110157452 | 1.93 | -39837 | intergenic |
| ENSMUSG00000081070 | 3713649 | 3714049 | 1.93 | 27287 | intergenic |
| ENSMUSG00000087294 | 38286565 | 38287065 | 1.93 | 310 | inside |
| ENSMUSG00000087524 | 131545673 | 131546223 | 1.93 | -15941 | intergenic |
| ENSMUSG00000017817 | 163339673 | 163340423 | 1.93 | 58320 | inside |
| ENSMUSG00000027297 | 119758223 | 119758723 | 1.93 | 2208 | inside |
| ENSMUSG00000087679 | 26140306 | 26140706 | 1.93 | 215 | overlap |
| ENSMUSG00000089251 | 44191367 | 44192117 | 1.93 | 265798 | intergenic |
| ENSMUSG00000027722 | 37419495 | 37419895 | 1.93 | -401 | upstream |
| ENSMUSG00000037610 | 31948830 | 31949680 | 1.93 | 46323 | inside |
| ENSMUSG00000074513 | 85887317 | 85887817 | 1.93 | 199 | overlap |
| ENSMUSG00000091329 | 28781003 | 28781803 | 1.93 | 105 | overlap |


| ENSMUSG00000039701 | 122984144 | 122984694 | 1.93 | 303 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000027895 | 107458944 | 107459494 | 1.93 | -46 | upstream |
| ENSMUSG00000028527 | 101419425 | 101419925 | 1.93 | 148 | inside |
| ENSMUSG00000092724 | 9668775 | 9669175 | 1.93 | -504 | upstream |
| ENSMUSG00000042198 | 3938527 | 3938977 | 1.93 | -361 | overlap |
| ENSMUSG00000039911 | 149954573 | 149954973 | 1.93 | 470 | inside |
| ENSMUSG00000086774 | 28815062 | 28815462 | 1.93 | 14 | overlap |
| ENSMUSG00000098563 | 91371151 | 91371551 | 1.93 | 2212 | downstream |
| ENSMUSG00000040860 | 141040309 | 141041009 | 1.93 | 20241 | inside |
| ENSMUSG00000054920 | 65107762 | 65108162 | 1.93 | 194 | inside |
| ENSMUSG00000010825 | 143372878 | 143373378 | 1.93 | 15540 | inside |
| ENSMUSG00000041870 | 114772570 | 114773020 | 1.93 | -2107 | upstream |
| ENSMUSG00000040274 | 3340657 | 3341557 | 1.93 | -3236 | upstream |
| ENSMUSG00000058291 | 138619365 | 138619815 | 1.93 | 396 | overlap |
| ENSMUSG00000029408 | 124095442 | 124095842 | 1.93 | 356 | overlap |
| ENSMUSG00000029207 | 66618368 | 66618868 | 1.93 | 460 | overlap |
| ENSMUSG00000016128 | 151233376 | 151233776 | 1.93 | 460 | inside |
| ENSMUSG00000093910 | 143293150 | 143293700 | 1.93 | -794 | upstream |
| ENSMUSG00000077474 | 83783752 | 83784302 | 1.93 | -60163 | intergenic |
| ENSMUSG00000042599 | 39205255 | 39205655 | 1.93 | 1534 | inside |
| ENSMUSG00000084772 | 17064633 | 17065133 | 1.93 | 363 | overlap |
| ENSMUSG00000034832 | 83441099 | 83441599 | 1.93 | 579 | inside |
| ENSMUSG00000051184 | 5015561 | 5016011 | 1.93 | 60 | inside |
| ENSMUSG00000037337 | 29018913 | 29019713 | 1.93 | 36059 | intergenic |
| ENSMUSG00000039176 | 79464340 | 79465340 | 1.93 | 2022 | inside |
| ENSMUSG00000098839 | 45007653 | 45008053 | 1.93 | 4834 | downstream |
| ENSMUSG00000031700 | 85492131 | 85492631 | 1.93 | -445 | overlap |
| ENSMUSG00000054823 | 25601517 | 25602067 | 1.93 | -84 | overlap |
| ENSMUSG00000071138 | 27318967 | 27319617 | 1.93 | -25427 | intergenic |
| ENSMUSG00000036990 | 79640031 | 79640531 | 1.93 | 413 | inside |
| ENSMUSG00000083285 | 23057900 | 23058900 | 1.93 | -69556 | intergenic |
| ENSMUSG00000038872 | 108791989 | 108792689 | 1.93 | 77345 | inside |
| ENSMUSG00000031458 | 13889522 | 13890022 | 1.93 | 749 | inside |
| ENSMUSG00000004383 | 73353323 | 73353773 | 1.93 | 217 | overlap |
| ENSMUSG00000051495 | 126593089 | 126593489 | 1.93 | 897 | inside |
| ENSMUSG00000001911 | 84799273 | 84799723 | 1.93 | 1071 | inside |
| ENSMUSG00000032009 | 14276641 | 14277041 | 1.93 | 340 | inside |
| ENSMUSG00000025887 | 5475435 | 5476085 | 1.93 | 130005 | intergenic |
| ENSMUSG00000032411 | 96195816 | 96196516 | 1.93 | -459 | overlap |
| ENSMUSG00000032498 | 111271131 | 111271531 | 1.93 | 477 | inside |
| ENSMUSG00000019471 | 21149791 | 21150441 | 1.93 | 115 | overlap |
| ENSMUSG00000032218 | 70421228 | 70421728 | 1.93 | 326 | overlap |
| ENSMUSG00000026062 | 40681194 | 40681594 | 1.92 | -518 | upstream |
| ENSMUSG00000026229 | 86064808 | 86065208 | 1.92 | 421 | inside |
| ENSMUSG00000026208 | 75360019 | 75360869 | 1.92 | -310 | overlap |
| ENSMUSG00000026049 | 44102382 | 44102782 | 1.92 | 59 | overlap |
| ENSMUSG00000026393 | 138619512 | 138620062 | 1.92 | 184 | overlap |
| ENSMUSG00000041859 | 20819838 | 20820288 | 1.92 | 440 | overlap |
| ENSMUSG00000025404 | 127380004 | 127380404 | 1.92 | -323 | overlap |
| ENSMUSG00000049764 | 76032583 | 76032983 | 1.92 | -71 | overlap |
| ENSMUSG00000020190 | 80676463 | 80676863 | 1.92 | -4487 | upstream |


| ENSMUSG00000074785 | 94944033 | 94944433 | 1.92 | 545 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000097086 | 21994345 | 21994845 | 1.92 | 52640 | intergenic |
| ENSMUSG00000015202 | 7211582 | 7211982 | 1.92 | 655 | inside |
| ENSMUSG00000020057 | 88356656 | 88357106 | 1.92 | 22424 | inside |
| ENSMUSG00000037533 | 54522880 | 54523280 | 1.92 | 33 | inside |
| ENSMUSG00000000617 | 50851429 | 50851829 | 1.92 | 744 | inside |
| ENSMUSG00000037926 | 77215577 | 77216077 | 1.92 | -710 | upstream |
| ENSMUSG00000086459 | 23497658 | 23498158 | 1.92 | -95 | overlap |
| ENSMUSG00000019368 | 4017126 | 4017576 | 1.92 | -14336 | intergenic |
| ENSMUSG00000000282 | 74831248 | 74831648 | 1.92 | 328 | inside |
| ENSMUSG00000020937 | 103101074 | 103101524 | 1.92 | 584 | inside |
| ENSMUSG00000034621 | 102555567 | 102556567 | 1.92 | 825 | overlap |
| ENSMUSG00000086058 | 82929086 | 82930286 | 1.92 | 4296 | inside |
| ENSMUSG00000063251 | 99627615 | 99628065 | 1.92 | 624 | inside |
| ENSMUSG00000089247 | 8058135 | 8059485 | 1.92 | -53119 | intergenic |
| ENSMUSG00000021259 | 108334367 | 108334917 | 1.92 | -10 | overlap |
| ENSMUSG00000020650 | 31634630 | 31635130 | 1.92 | 28 | overlap |
| ENSMUSG00000052593 | 21373316 | 21373716 | 1.92 | 316 | overlap |
| ENSMUSG00000051367 | 73045040 | 73045840 | 1.92 | 8847 | inside |
| ENSMUSG00000021287 | 111813843 | 111814243 | 1.92 | 30 | overlap |
| ENSMUSG00000021385 | 49399997 | 49400547 | 1.92 | -21314 | intergenic |
| ENSMUSG00000019726 | 13590489 | 13591239 | 1.92 | 80 | inside |
| ENSMUSG00000021696 | 108214521 | 108215021 | 1.92 | 117 | inside |
| ENSMUSG00000001504 | 72628371 | 72628821 | 1.92 | -449 | overlap |
| ENSMUSG00000032846 | 107890371 | 107890771 | 1.92 | -307 | upstream |
| ENSMUSG00000034522 | 65343712 | 65344162 | 1.92 | -14964 | intergenic |
| ENSMUSG00000063895 | 60251243 | 60251643 | 1.92 | 264 | overlap |
| ENSMUSG00000021944 | 63240326 | 63240726 | 1.92 | 31366 | inside |
| ENSMUSG00000022201 | 12118398 | 12118798 | 1.92 | 567 | inside |
| ENSMUSG00000022367 | 56692891 | 56693291 | 1.92 | 1648 | inside |
| ENSMUSG00000036800 | 71650071 | 71650971 | 1.92 | 77767 | inside |
| ENSMUSG00000039100 | 31531141 | 31531641 | 1.92 | -104 | upstream |
| ENSMUSG00000068167 | 64450347 | 64450747 | 1.92 | 28801 | intergenic |
| ENSMUSG00000022537 | 30550467 | 30550867 | 1.92 | 375 | overlap |
| ENSMUSG00000073411 | 35263422 | 35264222 | 1.92 | 692 | inside |
| ENSMUSG00000032855 | 24549728 | 24550128 | 1.92 | -222 | overlap |
| ENSMUSG00000084880 | 47672111 | 47672611 | 1.92 | -15499 | intergenic |
| ENSMUSG00000048915 | 62880708 | 62881458 | 1.92 | 609 | overlap |
| ENSMUSG00000079553 | 33890222 | 33890672 | 1.92 | 439 | overlap |
| ENSMUSG00000024456 | 37934827 | 37935377 | 1.92 | 649 | inside |
| ENSMUSG00000025231 | 46441925 | 46442425 | 1.92 | 45029 | inside |
| ENSMUSG00000024639 | 16133375 | 16133775 | 1.92 | 544 | inside |
| ENSMUSG00000024978 | 55098790 | 55099190 | 1.92 | 661 | inside |
| ENSMUSG00000027314 | 119322373 | 119322773 | 1.92 | -3411 | upstream |
| ENSMUSG00000027134 | 112238952 | 112239402 | 1.92 | -516 | upstream |
| ENSMUSG00000027349 | 117249835 | 117250235 | 1.92 | 96 | inside |
| ENSMUSG00000037843 | 157944049 | 157944499 | 1.92 | 29396 | inside |
| ENSMUSG00000016458 | 105127008 | 105127458 | 1.92 | 479 | inside |
| ENSMUSG00000082930 | 40917819 | 40918669 | 1.92 | -40572 | intergenic |
| ENSMUSG00000087264 | 70562264 | 70562714 | 1.92 | 1093 | inside |
| ENSMUSG00000075270 | 76338664 | 76339064 | 1.92 | 110 | overlap |


| ENSMUSG00000004113 | 24762228 | 24762878 | 1.92 | 924 | inside |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000046470 | 181671330 | 181671730 | 1.92 | 310 | overlap |
| ENSMUSG00000041997 | 70824678 | 70825378 | 1.92 | 1050 | inside |
| ENSMUSG00000074796 | 130697535 | 130697985 | 1.92 | -16 | upstream |
| ENSMUSG00000074682 | 152414349 | 152414999 | 1.92 | 695 | inside |
| ENSMUSG00000046688 | 127789834 | 127790234 | 1.92 | -38 | overlap |
| ENSMUSG00000027555 | 14641567 | 14642067 | 1.92 | -160 | overlap |
| ENSMUSG00000029056 | 154984113 | 154984563 | 1.92 | 19990 | downstream |
| ENSMUSG00000085863 | 101291675 | 101292325 | 1.92 | 7454 | intergenic |
| ENSMUSG00000028522 | 103113807 | 103114257 | 1.92 | -583 | upstream |
| ENSMUSG00000028790 | 129742074 | 129742474 | 1.92 | 229 | overlap |
| ENSMUSG00000089051 | 108031837 | 108032237 | 1.92 | 9963 | intergenic |
| ENSMUSG00000028741 | 139352007 | 139352457 | 1.92 | 569 | inside |
| ENSMUSG00000029466 | 122422492 | 122422942 | 1.92 | 799 | inside |
| ENSMUSG00000058558 | 107900338 | 107900738 | 1.92 | -164 | overlap |
| ENSMUSG00000000916 | 135369731 | 135370131 | 1.92 | -222 | overlap |
| ENSMUSG00000029345 | 112326109 | 112326509 | 1.92 | -249 | overlap |
| ENSMUSG00000034981 | 91517381 | 91517881 | 1.92 | -234 | overlap |
| ENSMUSG00000028949 | 24604569 | 24604969 | 1.92 | -2557 | upstream |
| ENSMUSG00000043614 | 135089673 | 135090123 | 1.92 | -11407 | intergenic |
| ENSMUSG00000055204 | 90366125 | 90366525 | 1.92 | 60 | overlap |
| ENSMUSG00000043323 | 110431785 | 110432185 | 1.92 | 16718 | inside |
| ENSMUSG00000029863 | 42264792 | 42265392 | 1.92 | -193 | overlap |
| ENSMUSG00000030256 | 145865883 | 145866333 | 1.92 | -325 | upstream |
| ENSMUSG00000099103 | 143445491 | 143446041 | 1.92 | -2114 | upstream |
| ENSMUSG00000002083 | 16309133 | 16309883 | 1.92 | -483 | overlap |
| ENSMUSG00000030562 | 87292166 | 87292716 | 1.92 | 46070 | inside |
| ENSMUSG00000033676 | 57590426 | 57590876 | 1.92 | -92 | overlap |
| ENSMUSG00000025505 | 141328061 | 141328461 | 1.92 | 322 | inside |
| ENSMUSG00000092071 | 89632453 | 89633153 | 1.92 | 671 | overlap |
| ENSMUSG00000030583 | 29518504 | 29519054 | 1.92 | 137 | overlap |
| ENSMUSG00000030678 | 127025268 | 127025718 | 1.92 | 1211 | inside |
| ENSMUSG00000031570 | 25719708 | 25720108 | 1.92 | -353 | overlap |
| ENSMUSG00000019139 | 70598581 | 70598981 | 1.92 | 4100 | downstream |
| ENSMUSG00000039067 | 107587630 | 107588230 | 1.92 | 852 | inside |
| ENSMUSG00000000792 | 70892540 | 70892940 | 1.92 | 217 | overlap |
| ENSMUSG00000045636 | 41133168 | 41133668 | 1.92 | 558 | inside |
| ENSMUSG00000096188 | 104394789 | 104395239 | 1.92 | 1018 | inside |
| ENSMUSG00000032410 | 95954616 | 95955066 | 1.92 | -144 | overlap |
| ENSMUSG00000032468 | 99568116 | 99568516 | 1.92 | 783 | inside |
| ENSMUSG00000099275 | 110638231 | 110638831 | 1.92 | -2116 | upstream |
| ENSMUSG00000036867 | 64020828 | 64021278 | 1.92 | 1231 | inside |
| ENSMUSG00000049307 | 14751491 | 14751891 | 1.92 | 631 | inside |
| ENSMUSG00000038412 | 121857817 | 121858217 | 1.92 | 183 | overlap |
| ENSMUSG00000032582 | 107872554 | 107873054 | 1.92 | 683 | inside |
| ENSMUSG00000067336 | 59763150 | 59763600 | 1.91 | -1129 | upstream |
| ENSMUSG00000026072 | 40266543 | 40266993 | 1.91 | 41463 | inside |
| ENSMUSG00000070644 | 133363556 | 133364256 | 1.91 | -16 | overlap |
| ENSMUSG00000089534 | 17511257 | 17511657 | 1.91 | -8744 | intergenic |
| ENSMUSG00000050069 | 175015157 | 175016107 | 1.91 | -93338 | intergenic |
| ENSMUSG00000099119 | 171064458 | 171064958 | 1.91 | 194 | includeFeature |


| ENSMUSG00000057173 | 39756549 | 39757149 | 1.91 | -35552 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000085184 | 172562007 | 172563057 | 1.91 | -3878 | upstream |
| ENSMUSG00000042772 | 152901889 | 152902589 | 1.91 | 757 | inside |
| ENSMUSG00000026360 | 144003839 | 144004239 | 1.91 | 322 | overlap |
| ENSMUSG00000026509 | 182517446 | 182517896 | 1.91 | 49 | overlap |
| ENSMUSG00000026121 | 36558049 | 36558499 | 1.91 | 300 | overlap |
| ENSMUSG00000055197 | 74882209 | 74882659 | 1.91 | 3210 | inside |
| ENSMUSG00000035027 | 81106083 | 81106483 | 1.91 | 168 | inside |
| ENSMUSG00000019952 | 99107299 | 99107849 | 1.91 | 263 | inside |
| ENSMUSG00000069539 | 89685513 | 89685913 | 1.91 | 772 | inside |
| ENSMUSG00000019803 | 42582554 | 42582954 | 1.91 | 1078 | inside |
| ENSMUSG00000039497 | 34207223 | 34207673 | 1.91 | 328 | overlap |
| ENSMUSG00000056758 | 120474945 | 120475545 | 1.91 | 1524 | inside |
| ENSMUSG00000004934 | 81154234 | 81154634 | 1.91 | 13689 | inside |
| ENSMUSG00000033416 | 75516783 | 75517283 | 1.91 | 1189 | inside |
| ENSMUSG00000020709 | 80153827 | 80154277 | 1.91 | -278 | overlap |
| ENSMUSG00000020359 | 51584627 | 51585027 | 1.91 | -130 | overlap |
| ENSMUSG00000087111 | 3146026 | 3146476 | 1.91 | 3978 | downstream |
| ENSMUSG00000018849 | 35979588 | 35980138 | 1.91 | 939 | inside |
| ENSMUSG00000085564 | 50602580 | 50603230 | 1.91 | -132 | upstream |
| ENSMUSG00000020902 | 68385327 | 68385777 | 1.91 | 15496 | inside |
| ENSMUSG00000020435 | 3863626 | 3864076 | 1.91 | 277 | overlap |
| ENSMUSG00000020925 | 102697565 | 102698065 | 1.91 | 217 | overlap |
| ENSMUSG00000096279 | 24251540 | 24252190 | 1.91 | 641 | includeFeature |
| ENSMUSG00000051367 | 73045940 | 73046490 | 1.91 | 7947 | inside |
| ENSMUSG00000034168 | 86884063 | 86884563 | 1.91 | 751 | inside |
| ENSMUSG00000091105 | 21899203 | 21899603 | 1.91 | -122496 | intergenic |
| ENSMUSG00000038175 | 45389697 | 45390197 | 1.91 | -45 | overlap |
| ENSMUSG00000039242 | 13954549 | 13954999 | 1.91 | -125 | overlap |
| ENSMUSG00000021326 | 21180421 | 21180821 | 1.91 | 976 | inside |
| ENSMUSG00000042167 | 93219485 | 93220285 | 1.91 | -27202 | intergenic |
| ENSMUSG00000069272 | 23570671 | 23571071 | 1.91 | 549 | inside |
| ENSMUSG00000021466 | 63572358 | 63573158 | 1.91 | -6838 | intergenic |
| ENSMUSG00000041014 | 39473325 | 39473775 | 1.91 | -237 | upstream |
| ENSMUSG00000021978 | 65097848 | 65098248 | 1.91 | 258 | overlap |
| ENSMUSG00000021944 | 63240926 | 63241326 | 1.91 | 30766 | inside |
| ENSMUSG00000036218 | 92397220 | 92397620 | 1.91 | 305 | inside |
| ENSMUSG00000063704 | 75998421 | 75998821 | 1.91 | 4652 | inside |
| ENSMUSG00000022472 | 82015971 | 82016421 | 1.91 | 17891 | inside |
| ENSMUSG00000022476 | 81925966 | 81926466 | 1.91 | 247 | overlap |
| ENSMUSG00000022451 | 94589738 | 94590138 | 1.91 | 151 | overlap |
| ENSMUSG00000089979 | 18066130 | 18066530 | 1.91 | -3084 | upstream |
| ENSMUSG00000022792 | 16302560 | 16303010 | 1.91 | -405 | overlap |
| ENSMUSG00000090882 | 60948288 | 60948738 | 1.91 | -110473 | intergenic |
| ENSMUSG00000022663 | 45158901 | 45159351 | 1.91 | 116 | inside |
| ENSMUSG00000014039 | 97851480 | 97851930 | 1.91 | 370 | overlap |
| ENSMUSG00000036304 | 43978959 | 43979509 | 1.91 | 134 | overlap |
| ENSMUSG00000047434 | 31081278 | 31081728 | 1.91 | 154 | overlap |
| ENSMUSG00000022894 | 85899858 | 85900458 | 1.91 | 1267 | inside |
| ENSMUSG00000024172 | 55445345 | 55445745 | 1.91 | -37 | overlap |
| ENSMUSG00000092612 | 36230348 | 36231298 | 1.91 | 1422 | downstream |


| ENSMUSG00000067235 | 35470078 | 35470478 | 1.91 | -11 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000089487 | 72464770 | 72465170 | 1.91 | 34648 | intergenic |
| ENSMUSG00000024070 | 79020853 | 79021253 | 1.91 | -37 | upstream |
| ENSMUSG00000024077 | 78736606 | 78737006 | 1.91 | 590 | inside |
| ENSMUSG00000023845 | 17623793 | 17624293 | 1.91 | 696 | inside |
| ENSMUSG00000046668 | 35847889 | 35848289 | 1.91 | 18071 | inside |
| ENSMUSG00000098276 | 76145609 | 76146009 | 1.91 | -24943 | intergenic |
| ENSMUSG00000042705 | 46958848 | 46959248 | 1.91 | -14 | overlap |
| ENSMUSG00000032656 | 56925183 | 56925633 | 1.91 | 365 | overlap |
| ENSMUSG00000024598 | 58209377 | 58209777 | 1.91 | 549 | inside |
| ENSMUSG00000003228 | 60906863 | 60907463 | 1.91 | 17114 | inside |
| ENSMUSG00000071657 | 8837320 | 8837770 | 1.91 | -147 | overlap |
| ENSMUSG00000064105 | 46761475 | 46761875 | 1.91 | -121 | overlap |
| ENSMUSG00000023307 | 37206770 | 37207170 | 1.91 | -773 | upstream |
| ENSMUSG00000024900 | 3325260 | 3325710 | 1.91 | 1959 | inside |
| ENSMUSG00000049401 | 180589166 | 180589666 | 1.91 | -79 | overlap |
| ENSMUSG00000086449 | 35719689 | 35720139 | 1.91 | -99501 | intergenic |
| ENSMUSG00000027259 | 121139907 | 121140557 | 1.91 | -521 | overlap |
| ENSMUSG00000027018 | 71388982 | 71389582 | 1.91 | 24 | inside |
| ENSMUSG00000059842 | 154613135 | 154613535 | 1.91 | -162 | overlap |
| ENSMUSG00000015647 | 180225730 | 180226230 | 1.91 | 129 | overlap |
| ENSMUSG00000028180 | 157533775 | 157534225 | 1.91 | -385 | overlap |
| ENSMUSG00000027562 | 14886425 | 14886875 | 1.91 | -1 | overlap |
| ENSMUSG00000037814 | 40949490 | 40949890 | 1.91 | -1141 | upstream |
| ENSMUSG00000025757 | 40744567 | 40745067 | 1.91 | 72 | inside |
| ENSMUSG00000027799 | 56135895 | 56136395 | 1.91 | 47806 | inside |
| ENSMUSG00000033882 | 82875930 | 82876430 | 1.91 | 553 | inside |
| ENSMUSG00000028745 | 139192177 | 139192577 | 1.91 | -722 | upstream |
| ENSMUSG00000042380 | 127214354 | 127214754 | 1.91 | -29430 | intergenic |
| ENSMUSG00000050212 | 126149398 | 126149798 | 1.91 | 1654 | inside |
| ENSMUSG00000046637 | 154855503 | 154855903 | 1.91 | -697 | upstream |
| ENSMUSG00000036052 | 42949826 | 42950376 | 1.91 | 12 | inside |
| ENSMUSG00000082388 | 98246831 | 98247481 | 1.91 | -11541 | intergenic |
| ENSMUSG00000041135 | 16163483 | 16163883 | 1.91 | 164 | overlap |
| ENSMUSG00000054659 | 33189317 | 33189717 | 1.91 | 420 | inside |
| ENSMUSG00000070576 | 111416073 | 111416823 | 1.91 | -1289 | upstream |
| ENSMUSG00000070639 | 105415682 | 105416082 | 1.91 | -93 | overlap |
| ENSMUSG00000034118 | 130079487 | 130079987 | 1.91 | 6161 | inside |
| ENSMUSG00000029095 | 35757486 | 35757886 | 1.91 | -394 | overlap |
| ENSMUSG00000029122 | 37336312 | 37336912 | 1.91 | 582 | overlap |
| ENSMUSG00000058153 | 112577065 | 112577465 | 1.91 | 133 | overlap |
| ENSMUSG00000044221 | 88674882 | 88675682 | 1.91 | 1289 | inside |
| ENSMUSG00000029505 | 110770016 | 110770666 | 1.91 | 701 | inside |
| ENSMUSG00000001632 | 113306449 | 113306849 | 1.91 | -732 | upstream |
| ENSMUSG00000044927 | 87981089 | 87981939 | 1.91 | 393 | overlap |
| ENSMUSG00000030376 | 16144648 | 16145398 | 1.91 | 14348 | inside |
| ENSMUSG00000002068 | 38106854 | 38107304 | 1.91 | 680 | inside |
| ENSMUSG00000037606 | 143739927 | 143740327 | 1.91 | 17058 | inside |
| ENSMUSG00000070462 | 83884053 | 83884453 | 1.91 | 288 | overlap |
| ENSMUSG00000055323 | 126776532 | 126776932 | 1.91 | 286 | overlap |
| ENSMUSG00000031962 | 122870289 | 122870689 | 1.91 | 21915 | downstream |


| ENSMUSG00000080348 | 43869718 | 43870118 | 1.91 | -169532 | intergenic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000031511 | 11727472 | 11727872 | 1.91 | -633 | upstream |
| ENSMUSG00000001472 | 123373389 | 123373839 | 1.91 | -435 | overlap |
| ENSMUSG00000040028 | 4325022 | 4325622 | 1.91 | 78 | overlap |
| ENSMUSG00000092842 | 65108023 | 65108423 | 1.91 | -14109 | intergenic |
| ENSMUSG00000069867 | 122630289 | 122630889 | 1.91 | -7550 | intergenic |
| ENSMUSG00000003316 | 111258472 | 111258872 | 1.91 | 744 | inside |
| ENSMUSG00000003657 | 110167939 | 110168489 | 1.91 | 267 | overlap |
| ENSMUSG00000003575 | 70451605 | 70452055 | 1.91 | -12026 | intergenic |
| ENSMUSG00000006676 | 108488304 | 108488854 | 1.91 | -2372 | upstream |
| ENSMUSG00000032336 | 58582178 | 58582778 | 1.91 | -62 | overlap |
| ENSMUSG00000057895 | 122922767 | 122923167 | 1.91 | -305 | overlap |
| ENSMUSG00000087817 | 112981341 | 112981741 | 1.91 | -422134 | intergenic |
| ENSMUSG00000038119 | 35420900 | 35421300 | 1.91 | -228 | overlap |
| ENSMUSG00000032459 | 98601366 | 98601816 | 1.91 | 313 | overlap |
| ENSMUSG00000092963 | 107231354 | 107231804 | 1.91 | 354 | includeFeature |
| ENSMUSG00000079559 | 51278580 | 51278980 | 1.91 | -26 | upstream |
| ENSMUSG00000042207 | 134556617 | 134557017 | 1.90 | -3554 | upstream |
| ENSMUSG00000035595 | 80171406 | 80171906 | 1.90 | -1538 | upstream |
| ENSMUSG00000019943 | 98915545 | 98915945 | 1.90 | 393 | inside |
| ENSMUSG00000020308 | 79669106 | 79669806 | 1.90 | -304 | overlap |
| ENSMUSG00000020124 | 123196637 | 123197137 | 1.90 | 298 | overlap |
| ENSMUSG00000058537 | 58225162 | 58225562 | 1.90 | 544 | inside |
| ENSMUSG00000043999 | 30885279 | 30885729 | 1.90 | -79 | overlap |
| ENSMUSG00000062115 | 60105636 | 60106536 | 1.90 | 623 | inside |
| ENSMUSG00000082587 | 38164179 | 38165329 | 1.90 | -54890 | intergenic |
| ENSMUSG00000009073 | 4848876 | 4849526 | 1.90 | 660 | inside |
| ENSMUSG00000056962 | 116843568 | 116843968 | 1.90 | -119 | upstream |
| ENSMUSG00000072825 | 112721316 | 112722016 | 1.90 | -858 | upstream |
| ENSMUSG00000020653 | 24651440 | 24651890 | 1.90 | 69 | inside |
| ENSMUSG00000097758 | 110187484 | 110187934 | 1.90 | 90584 | intergenic |
| ENSMUSG00000021488 | 55211705 | 55212455 | 1.90 | 1923 | inside |
| ENSMUSG00000051335 | 43303077 | 43303577 | 1.90 | 1095 | inside |
| ENSMUSG00000005148 | 99299193 | 99299593 | 1.90 | 502 | inside |
| ENSMUSG00000046160 | 91270242 | 91270742 | 1.90 | 473 | inside |
| ENSMUSG00000063239 | 27512251 | 27512751 | 1.90 | 1090 | inside |
| ENSMUSG00000049090 | 84087321 | 84087721 | 1.90 | -837 | upstream |
| ENSMUSG00000024529 | 52528927 | 52529377 | 1.90 | 940 | inside |
| ENSMUSG00000078201 | 25255456 | 25255856 | 1.90 | 17 | inside |
| ENSMUSG00000038467 | 154657073 | 154657573 | 1.90 | 5368 | inside |
| ENSMUSG00000027439 | 148680990 | 148681390 | 1.90 | -33 | overlap |
| ENSMUSG00000065083 | 120730049 | 120730849 | 1.90 | -89214 | intergenic |
| ENSMUSG00000048647 | 119547329 | 119547729 | 1.90 | 298 | overlap |
| ENSMUSG00000026878 | 35200715 | 35201265 | 1.90 | 405 | overlap |
| ENSMUSG00000004897 | 87906095 | 87906495 | 1.90 | -226 | overlap |
| ENSMUSG00000046743 | 38885846 | 38886246 | 1.90 | -1094 | upstream |
| ENSMUSG00000028078 | 86920330 | 86920930 | 1.90 | 554 | overlap |
| ENSMUSG00000028478 | 44012476 | 44012976 | 1.90 | 8024 | inside |
| ENSMUSG00000015247 | 53011808 | 53012258 | 1.90 | -72 | overlap |
| ENSMUSG00000055761 | 20778251 | 20778751 | 1.90 | 615 | inside |
| ENSMUSG00000087383 | 46345100 | 46345550 | 1.90 | 415 | overlap |


| ENSMUSG00000047221 | 21424911 | 21425411 | 1.90 | -47 | overlap |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ENSMUSG00000037736 | 66745718 | 66746318 | 1.90 | -117 | overlap |
| ENSMUSG00000029120 | 36867912 | 36868462 | 1.90 | -601 | upstream |
| ENSMUSG00000029725 | 137779223 | 137779623 | 1.90 | 374 | inside |
| ENSMUSG00000041609 | 115732342 | 115732792 | 1.90 | -721 | upstream |
| ENSMUSG00000029467 | 122500920 | 122501420 | 1.90 | 1305 | inside |
| ENSMUSG00000015882 | 45857140 | 45857540 | 1.90 | 475 | inside |
| ENSMUSG00000049694 | 90309647 | 90310047 | 1.90 | 8428 | downstream |
| ENSMUSG00000001521 | 128355665 | 128356215 | 1.90 | 186 | overlap |
| ENSMUSG00000030091 | 91116799 | 91117199 | 1.90 | 30 | overlap |
| ENSMUSG00000044030 | 19004657 | 19005157 | 1.90 | 592 | inside |
| ENSMUSG00000002083 | 16299377 | 16299877 | 1.90 | -10239 | intergenic |
| ENSMUSG00000076051 | 44852606 | 44853056 | 1.90 | 2907 | downstream |
| ENSMUSG00000031812 | 121590691 | 121591091 | 1.90 | 330 | inside |
| ENSMUSG00000037070 | 78508631 | 78509931 | 1.90 | 297 | overlap |
| ENSMUSG00000001300 | 8660830 | 8661330 | 1.90 | -57 | upstream |
| ENSMUSG00000003410 | 22050191 | 22050641 | 1.90 | 1832 | inside |

Table 4.2

| Gene list | Number of <br> expressed <br> genes | Expressed <br> genes near DE <br> Sono-seq peak | Overlap | Expected Overlap |
| :--- | :---: | :---: | :---: | :---: |
| Differential RNA-seq <br> (Chapter 2) | 823 | 86 | $5.1 \%$ | $4.6 \%$ |
| diffSplice (Chapter 2) | 118 | 20 | $1.2 \%$ | $0.7 \%$ |
| Autism (SFARI) | 420 | 59 | $3.5 \%$ | $2.3 \%$ |
| Epilepsy (GenEpi) | 83 | 5 | $0.3 \%$ | $0.5 \%$ |

Table 4.3

| Data set | Number of regions in the <br> genome | Percent overlap with differential <br> Sono-seq regions |
| :--- | :---: | :---: |
| Sono-seq Hippocampus <br> (Abel lab) | $\mathbf{3 6 1 8 6}$ | $\mathbf{8 . 5 \%}$ |
| H3K9/14ac Hippocampus <br> (Abel lab) | $\mathbf{3 8 2 1 1}$ | $\mathbf{5 . 1 \%}$ |
| H3K27ac Adult Cortex | 35684 | $3.2 \%$ |
| H3K27ac E14 Whole brain | 36505 | $3.1 \%$ |
| H3K4me1 Adult Cortex | 90067 | $0.5 \%$ |
| H3K4me1 E14 Whole brain | 131408 | $1.3 \%$ |
| H3K4me3 Adult Cortex | 18981 | $7.8 \%$ |
| H3K4me3 E14 Whole brain | 19939 | $8.7 \%$ |
| H3K27me3 E14 Whole brain | 12020 | $8.7 \%$ |
| H3K36me3 E14 Whole brain | 168449 | $0.1 \%$ |
| Pol2 Adult Cortex | 19448 | $4.6 \%$ |
| DNASE I Cerebrum | 147444 | $0.6 \%$ |

## CHAPTER 5: Conclusions And Future Directions

This work focuses on the regulation of gene expression during memory consolidation, the process that converts labile short-term memories to stable long-term memories. Memory consolidation has been known to involve transcription for decades, yet the targets and regulation of this transcription have remained unclear. The advancement of high-throughput sequencing technology in recent years has greatly influenced our ability to study these changes on a genome-wide scale. This work aimed to define the transcriptional profile that occurs in response to contextual and spatial learning and determine how this profile is regulated. In Chapter 1, I introduced what is known about the regulation of gene expression by histone acetylation during memory consolidation. In Chapter 2, I used RNA-seq with a newly designed normalization method to discover the genes regulated during memory consolidation. We also used this data to probe for exonlevel changes and discovered large-scale regulation of alternative splicing following training. In Chapter 3, I compared the gene expression profile during memory consolidation after contextual learning to that after spatial learning in the hippocampus. I found that differentially expressed genes are similar between training paradigms but the temporal dynamics of these changes differ. In Chapter 4, I used ChIP-seq, Sono-seq, and MNase-seq to determine how genes were being regulated during memory consolidation. We found that chromatin accessibility changes without large changes in H3K9/14ac or nucleosome positioning. In Chapter 5, I will review my findings and suggest future directions for research on this topic. Additionally, I will propose a model for how gene regulation could occur during memory consolidation.

### 5.1 Gene expression changes during memory consolidation

Studies of genome-wide gene expression changes after hippocampal learning have been previously attempted using microarrays [70, 71, 110-112], but there have been few genes discovered by this method and little overlap between labs. Two recent studies have used RNA-seq to study gene expression in mutant mice after learning [76, 160]. The advent of next-generation sequencing technology to study gene expression using RNA-seq provides a number of benefits over these previous studies. RNA-seq produces better resolution than microarrays, the ability to detect novel transcripts, and the ability to quantify alternative splicing. In addition, the variance between sequencing runs could prove to be less substantial than between microarray runs that rely on hybridization. Therefore, normalizing RNA-seq data in a standard way should produce reproducible results between labs and between training paradigms.

In Chapter 2, we used RNA-seq to study gene expression at 30 minutes after contextual fear conditioning. This is a time point at which our lab has observed maximum gene expression differences after learning [61, 70]. We discovered that standard RNAseq normalization procedures are unable to capture the difference between untrained and trained groups. This leads to a small list of differentially regulated genes that may or may not be caused by the learning event. Therefore, we applied the recently published remove unwanted variation (RUV) normalization [114] that is an improved normalization method for noisy data sets such as the whole hippocampal samples used in our study. Briefly, this method of normalization includes an additional factor to account for unwanted variation by using negative control genes that are known not to be altered by training. RUV allowed us to differentiate between trained and untrained groups, meaning that any differentially expressed genes were likely the result of the contextual fear training. We discovered that this normalization method greatly improved the number of
novel genes detected as different after learning. Importantly, this analysis also increased the proportion of positive control genes discovered, suggesting that it was functioning as expected. Because RUV normalization makes fear conditioning the major source of variation between samples, the list of genes differentially expressed 30 minutes after learning using RUV normalization will provide a reproducible set of genes showing changes in response to learning. This method of normalization can be applied to all RNA-seq studies and will greatly improve detection power and reliability of results in future studies of brain function.

The regulation of gene expression is a highly complex process that includes transcription of a primary transcript, 5' capping, polyadenylation, and splicing into a mature mRNA. Alternative splicing is a coordinated process by which different transcripts can be produced from the same gene. Regulation of alternative splicing has been recognized in circadian function [161], addiction [162], and neurodegeneration [163]. There are also multiple individual examples of alternative splicing regulating learning and memory [134, 164-169], indicating this process may be an important regulatory step in the nervous system. However, no genome-wide studies have been used to investigate the regulation of alternative splicing during memory consolidation. Because RNA-seq also provides the ability to study exon-specific events such as those occurring by differential splicing, we applied RUV normalization to exon-specific analysis and demonstrated numerous exon-specific expression changes occurring during memory consolidation. We validated a number of these changes, including Ania-3 (a poorly studied isoform of Homer1), translational regulator Las1l and RNA-binding protein Rbm3. We believe that this analysis provides the first description of large-scale differential exon usage in response to learning. Although transcription of splicing factors is regulated by fear conditioning [134], it is unclear whether these changes would be 167
translated quickly enough to cause the splicing changes observed. It is possible that histone modification changes could be altering exon usage [135, 137], but more work is necessary to show these marks change in response to training. Future studies can be conducted to see if transcripts containing or excluding the identified differential exon lead to changes in localization or function of the protein.

The major question that remained about the genes discovered by our RNA-seq analysis was whether these changes depend on the training paradigm used. In other words, are the genes regulated by contextual fear conditioning the same as those regulated by other hippocampus-dependent learning tasks? To answer this question, we used a high-throughput qPCR approach in Chapter 3. The goal of this study was to compare the targets and temporal profile of gene expression after training for objectlocation memory (OLM), a spatial learning task, to that of fear conditioning, a contextual learning task. We discovered that while gene targets are regulated in a similar manner after OLM, the temporal dynamics of these gene expression changes differs from that observed after fear conditioning. A subset of genes regulated 30 minutes after OLM remain elevated 2 hours after training, while these same genes return to baseline by 2 hours after fear conditioning. Although the stress of a footshock during contextual fear training may be expected to produce a larger transcriptional response, it appears that the three training trials used for OLM result in longer lasting transcriptional changes. Therefore, we hypothesize that a common set of targets are regulated by all forms of hippocampal learning, but the timing of these changes can differ based on the paradigm being tested.

### 5.2 Chromatin accessibility during memory and the histone code hypothesis

Although the mechanism of highly coordinated regulation of specific transcripts during memory consolidation remains a mystery, epigenetic mechanisms are beginning to be implicated in this process. Histone modifications [51, 52, 156], histone variants [160], DNA methylation [170], miRNA regulation [107], and nucleosome positioning [76] have all been implicated in regulation of hippocampus-dependent learning. Histone acetylation, an activating histone modification, is the best studied epigenetic modification during memory consolidation. Work from our lab and others has implicated the histone acetyltransferase CBP as a positive regulator of learning [26,27] and the class I histone deacetylase (HDAC) proteins as negative regulators [55, 58, 63]. Thus, it appears that more histone acetylation during memory consolidation leads to enhanced long-term memory and less histone acetylation leads to impaired long-term memory. Histone acetylation is thought to decrease the interaction between the positively charged lysine residue of the histone and negatively charged DNA backbone, thereby increasing accessibility of chromatin in the surrounding region [35]. In Chapter 4, we used highthroughput sequencing to study both the histone acetylation and chromatin accessibility changes that occur 30 minutes after contextual fear memory.

A pilot ChIP-seq experiment investigating three histone acetylation marks found that H3K9/14ac, a mark we have previously studied [55], displayed the largest number of changes at promoters. We followed up this result by studying H3K9/14ac genome-wide in a large cohort of mice. We also investigated whether changes in chromatin accessibility, which would be the anticipated result of histone acetylation, occur at the same time. To our surprise, we found only a modest increase in $\mathrm{H} 3 \mathrm{~K} 9 / 14 \mathrm{ac}$ surrounding the transcription start site of genes after fear conditioning. This small increase was not large enough to be significant at any one particular gene but could be seen when
averaging across all genes. However, we found a large increase in chromatin accessibility surrounding the transcription start site of genes using Sono-seq [82]. This increase was significant at 3064 regions in the genome. These regions are often found within gene bodies and are enriched in genes that show alternative splicing after learning and those implicated in autism, a known cognitive disorder. We believe that these sites of increased chromatin accessibility represent sites in the genome with active chromatin reorganization occurring during memory consolidation.

There are a number of potential causes for this increase in chromatin accessibility after learning. The most obvious explanation would be a shift in nucleosome position. Less nucleosomes in a region would be expected to increase accessibility to that region. Therefore, we used MNase-seq to map nucleosome positioning throughout the genome. We found no difference in nucleosome positioning in response to learning, indicating that this is not the primary force driving the increase in chromatin accessibility. Changes in transcription factor occupancy, including CREB, could also be driving this change in accessibility. However, it is unclear whether increased transcription factor occupancy would result in increases or decreases in Sono-seq signal. Also, there are many more differential peaks than genes showing altered gene expression, so the transcription factor would have to be selectively active at a subset of genes.

Finally, histone modifications besides H3K9/14ac may be responsible for this increase in accessibility. The histone code hypothesis, put forward in 2000 by Strahl and Allis [30], states that "distinct histone modifications, on one or more tails, act sequentially or in combination to form a 'histone code' that is, read by other proteins to bring about distinct downstream events." This hypothesis has since been updated to acknowledge that combinatorial modifications probably do not create a specific "code" but rather a "language" that is dependent on surrounding context [155]. In either case, small changes
in any particular histone modification could lead to large changes in downstream function through combinatorial interactions with other marks, which would match our results. H3K9/14ac alone may not be significant enough to regulate the accessibility of chromatin, but instead may act in concert with a large number of other histone modifications to regulate this accessibility. Hippocampal learning may therefore be changing histone modifications in such a manner as to increase accessibility and prepare for transcription to occur.

We propose that the increase in chromatin accessibility 30 minutes after learning may be a set of combinatorial histone modifications removing a gate that allows transcription to occur. In this model (Figure 5.1), a number of sites would be "opened" after learning but only a subset of those sites would be bound by the factors necessary to drive transcription, regulate alternative splicing, or maintain that "open" state for later transcription. This leads to the intriguing question of whether there is a specific histone "memory code" that hippocampal neurons use to regulate activity after a learning event. This "memory code" could be a storage mechanism for long-term memory, with neurons exhibiting a particular epigenomic code ready to be rapidly activated during memory recall. This would be a large departure from the classic view of memory being stored at particular synapses through strengthening or weakening of connections, which has a limited number of possibilities at any individual synapse. Given the tremendous array of possible histone modifications, it seems likely that each neuron will have an individual code that could regulate the ability of that cell to participate in memory traces.

Although there is still a lot of work needed to test this possibility, there are technological advances that are going to make this possible in the near future. First, the cost of sequencing is getting cheaper every year and numerous histone modifications could be tested and overlaid to look for patterns in response to learning. Second, there
has been a major advancement in mass spectrometry-based quantification of histone modifications. This technology will allow users to test the relative abundance of all histone modifications throughout the genome in response to learning [171]. In addition, new techniques can quantify combinatorial modifications occurring on the same histone molecule, truly testing the histone code hypothesis [172, 173]. Future studies will use these novel technologies to test whether a specific "memory code" exists in response to a learning event. Novel genome-targeting technologies [174] can then be used to test whether disrupting this code at particular genes changes the response of these genes to a learning event.

### 5.3 Future Directions: The need for sorting technologies for brain epigenomic research

The study of epigenomics in the brain is a difficult, but promising, avenue for future research in to learning and memory, addiction, depression, neurodevelopmental and neurodegenerative disease. The brain is a complex organ that contains numerous cell types functioning in coordinated fashion to elicit specific responses. Our data make it clear that the changes in response to a behavior such as contextual or spatial learning are small. We believe this is due to a dilution of signal caused by the small number of cells responding to any particular memory trace. Transcription is an amplification step, where one copy of DNA can lead to multiple copies of RNA. This allows mRNA studies to identify reliable changes throughout the genome. However, epigenomic changes such as $\mathrm{H} 3 \mathrm{~K} 9 / 14 \mathrm{ac}$ do not involve an amplification step and are much more susceptible to being lost by cellular dilution. Therefore, future studies of the epigenome in the brain will need to rely on sorting technologies to study the cells of interest for that particular
experiment. Efforts are being made by a number of labs to produce a reliable method of sorting for this type of research. Flourescence-activated cell sorting has been using in combination with ChIP-qPCR [175] or ChIP-seq [176] to study the epigenome of cells of interest, and biotin tagging of the nuclear envelope can immunoprecipitate whole nuclei [177].

Combining technologies will lead to a greater understanding of the role of the epigenome in regulating long-term memory. RNA-seq could be performed on the same sorted cells used by the technology above. Technologies to sort cells for chromatin analysis could be combined with translating ribosome affinity purification (TRAP), which is a technique that purifies actively translating mRNA [178, 179]. When combined with the sorting strategies above, information on the timing of epigenomic changes could be compared to mRNA levels and translation in a cell-type specific manner. This would provide the first timeline of gene regulation, transcription, and translation during memory consolidation and would be a powerful method for determining the crucial steps regulating the transcription and translation necessary for long-term memory formation.

## Figure Legends

Figure 5.1. Model for the regulation of chromatin accessibility after learning. Learning causes changes in many histone modifications including H3K9/14ac, H3K4me3 and H3K27me3. These modifications act in concert to increase in chromatin accessibility in a large number of genes throughout the genome, opening the "gate" of repression. Transcription factors including CREB and CBP, splicing factors, and basal transcriptional machinery can then bind to their intended targets, leading to increased transcription of only a subset of "open" regions. Green indicates an induced gene and red indicated an inactive gene.

## Contributions

This chapter was written by Shane G. Poplawski with suggestions by Ted Abel.

Figures
Figure 5.1


## APPENDIX: Cell-type Specific Epigenomics In The Brain


#### Abstract

Epigenetics, the modification of gene expression without altering the underlying DNA sequence, plays a crucial role in regulating brain function including memory, drug addiction, and neurodegenerative disease. Despite important breakthroughs in epigenetics in the brain, the tools to study epigenetic regulation in specific cellular subpopulations do not currently exist. This is due to the complex heterogeneity of the brain, which can obscure important signals that occur in specific subsets of cells. To solve this problem, we are using a tetO-regulated, HA-tagged histone H3.3. Histone H3.3 incorporates preferentially into chromatin in actively transcribed regions independently of DNA replication, while H 3.1 and H 3.2 incorporate into silenced regions - a phenomenon known as histone barcoding [180]. The tetracycline transactivator (tTA), which allows expression of tetO-regulated transgenes, can be controlled in a cellspecific manner using cell-type specific promoters and temporally regulated by administration of doxycycline. Therefore, the tagged histone H 3.3 will be a marker of active chromatin specifically in cells of interest. We are using the CaMKIl - -tTA driver line to express this tagged histone in excitatory forebrain neurons. ChIP for the HA tag will isolate nucleosomes bound to active regions of the excitatory neuron genome for further investigation. Our work initially focuses on excitatory neurons because epigenetic changes in these cells have been linked to memory storage, but future studies will extend this approach to interneurons, glia and cells recently activated by experience. Understanding the histone modifications that occur during memory consolidation may uncover novel therapeutic targets for diseases in which cognitive deficits occur, including schizophrenia and Alzheimer's disease. In addition to addressing the important question


of which combinations of histone modifications change after memory, this approach promises to provide tools that can be used by researchers in all fields that struggle with cellular heterogeneity.

## Introduction

The formation and storage of long-term memory depends on the hippocampus. This occurs through a process involving distinct stages of acquisition, consolidation, maintenance and retrieval [2]. Through years of research, it has been demonstrated that memory consolidation requires transcription and translation to convert labile short-term memories to stable long-term memories [11, 16]. Although transcription has been known to be involved in memory formation for many years, the processes involved in regulating this transcription have only recently been uncovered in the field of epigenetics. Epigenetics refers to the set of mechanisms that control the regulation of gene expression without altering the underlying DNA sequence. Epigenetic regulation guides the binding of transcriptional machinery to the proper genetic loci. The best-studied epigenetic process in the brain is the post-translational modification of histone proteins within the nucleosome [50-52, 156]. Histones can be modified by the addition of acetylation, phosphorylation, methylation and other marks to achieve a combinatorial "histone code" that regulates transcription [30]. Many of these modifications demonstrate positive or negative correlations with transcriptional level [181]. Histone acetylation, an activating mark, plays a critical role in long-term memory. Decreasing histone acetylation by genetic deletion of the histone acetyltransferase CREB-binding protein (CBP) in the hippocampus reduces long-term memory [23, 24, 26-28, 149], whereas increasing histone acetylation by pharmacological inhibition of histone deacetylases (HDACs), which remove acetyl groups, enhances long-term memory [51, 55, 56, 63]. These published studies all used extracts prepared from the hippocampus which contain excitatory neurons, inhibitory neurons and glia.

A major problem for the field of epigenetics in the brain and other organs is the complex cellular heterogeneity of the tissue. The hippocampus contains excitatory neurons, inhibitory neurons, and glia, each of which can be divided into smaller subpopulations. We propose using histone barcoding to isolate chromatin of interest from specific cellular subpopulations. Prior attempts to study chromatin dynamics in particular cell types in mice have relied on fluorescence-activated cell sorting (FACS), which is an arduous procedure that has the potential to alter chromatin dynamics and gene expression [175]. Alternatively, immunopanning (using antibodies to purify cells based on external receptors) has been used to isolate specific cells from heterogeneous tissue to study RNA expression [182]. This technique could be used to study epigenetic regulation, but the time and dissociation steps required could themselves alter both histone modifications and gene expression [183]. Therefore, use of a single, simple approach adaptable to multiple cell types is necessary to avoid technique-driven biases.

Canonical histone proteins can be replaced by variant forms. Histone H3 has three variants in the mouse, H 3.1 and H 3.2 , which differ by only one amino acid, and H 3.3 , which differs by four amino acids from H3.2. Despite this high sequence similarity, histones H3.1 and H3.2 incorporate into chromatin only during DNA replication whereas H 3.3 incorporates independently of replication. Further, H 3.3 has been shown to preferentially incorporate into actively expressed regions of chromatin, whereas H3.1 and H3.2 are found in non-transcribed regions [184, 185]. This phenomenon is termed the "H3 barcode hypothesis," which states that genomic regions are "barcoded" with H3 histone variants to serve as a method for long-term cellular memory of transcriptional states [180]. We have developed a mouse model expressing a tagged histone H3.3 in specific cell populations to isolate actively transcribed regions of the genome selectively
from cells of interest. This mouse model uses a tetO-driven tagged H 3.3 in combination with a tTA driver line of choice [186]. This system has the advantage of being temporally controlled by administration of doxycycline and provides modularity so that this mouse line can be used to isolate any cell population of interest.

Memory is thought to depend critically on transcriptional changes in excitatory neurons. NMDA and AMPA receptors, which are activated by excitatory neurotransmitter glutamate, are known to be essential for synaptic plasticity and memory [187, 188]. Additionally, the selective deletion of CBP or HDACs selectively in excitatory neurons using transgenes driven by the CaMKIla promoter impairs or enhances long-term memory respectively [24, 26]. Therefore, our work initially focuses on the set of excitatory neurons marked by the CaMKIla promoter. However, we anticipate that this tool will be widely applicable to the study of epigenetics in the context of neurological and psychiatric disorders, metabolic disorders, cardiac disease, developmental disorders and cancer.

We were able to produce mouse lines that express the HA-tagged H3.3. It was found that using the tTA system allowed for expression of tagged H 3.3 to be restricted to adulthood by doxycycline and that expression only occurred in CaMKIla positive neurons. Also, we found that ChIP for the HA tag of H3.3 isolates promoters from genes expressed in neurons selectively, proving that we can sort nucleosomes at the chromatin level.

## Materials and Methods

## Subjects

tetO-H3.3-HA mice were created using a mouse-codon optimized H3.3-HA sequence from Geneart (Life Technologies Grand Island, NY) cloned into the mm400 plasmid [61, 186]. The plasmid was cut using Not1, the proper band was gel excised and sequenced to ensure it was mutation-free. The gel-extracted DNA was submitted to the Transgenic and Chimeric Mouse Facility at UPENN, where it was injected into C57BL/6J mouse eggs for the production of transgenic mice. Mice were genotyped using the following primers: F: 5'-GCGTCCATCTGGTCAGAAA-3', R: 5'-TGGAATCTCAGGTCGGTCTT-3'. 4 pups expressing the tetO-H3.3-transgene were obtained and tested for expression. All tested lines expressed the transgene, but only 3 lines appeared to have a typical CaMKIIa pattern. Line 52 was chosen for future experiments based on high expression and good breeding. All mice were on a 12 hr light/ 12hr dark schedule (lights on 7AM) with food and water available ad libitum. All procedures were approved by the University of Pennsylvania Institutional Animal Care and Use Committee.

## Immunohistochemistry

All mice were bred in breeding cages with doxycycline food. At weaning, some mice were removed from doxycycline while others remained on doxycycline food. Mice were transcardially perfused with 4\% paraformaldehyde in PBS, postfixed overnight, and cryoprotected in $30 \%$ sucrose for 2 days. Brains were flash frozen in 2-methylbutane on dry ice and $30 \mu \mathrm{M}$ coronal sections were prepared using a cryostat. Free floating sections were permeabilized using $0.1 \%$ Triton $\mathrm{X}-100$ (TX-100) in PBS for 5 min and autofluorescence was quenched by addition of $1 \% \mathrm{H} 2 \mathrm{O} 2$ in PBS 15 min at room temperature. Sections were blocked in $8 \%$ normal goat serum (NGS) with $0.3 \%$ TX-100 for 50 min at room temperature and washed $3 x$ in PBS for 5 min each. Sections were incubated with the HA antibody (1:500, Roche Clone 3F10) in $2 \%$ NGS and $0.3 \%$ TX-

100 by rocking overnight at $4^{\circ} \mathrm{C}$. After washing, sections were incubated in PBST containing biotinylated goat anti-rat (1:1000, Jackson ImmunoResearch \# 112-065-003) for 2 hrs at room temperature. Sections were washed again, and incubated in ABC solution (Vector \#PK-4000) for 1.5hrs at room temperature. Sections were washed again and incubated in 2 ml DAB solution ( $0.2 \mathrm{mg} / \mathrm{ml}$ ). 100 ul of $0.1 \% \mathrm{H} 2 \mathrm{O} 2$ was added and sections were incubated for 20 min to complete staining. Sections were mounted, dried overnight, coverslipped and visualized using a light microscope.

## Immunofluorescence

Immunofluorescence was performed as described for immunohistochemistry to the primary antibody stage. The following primary antibodies were used: HA, CaMKIla (1:1000 Santa Cruz \#sc-32288), GFAP (1:1000 Millipore\#MAB3402X [alexafluor 488 conjugated]), Pvalb (1:1000 Abcam \#ab11427) and rocked in the dark overnight at $4^{\circ} \mathrm{C}$. After washing, the following secondary antibodies were used: Goat anti-rat Alexa Fluor 555 for HA (1:1000 Invitrogen \#A-21434), goat anti-mouse Alexa Fluor 488 for CaMKIl $\alpha$ (1:1000 Invitrogen\#A-11001), and goat anti-rabbit Alexa Fluor 488 for Pvalb (1:1000 Invitrogen\#A-11034). 10 drops of DAPI (Life Technologies \#R37606) were added at this time as well and sections were incubated in the dark at room temperature for 2 hrs . After washing, cells were slide mounted, dried overnight in the dark, coverslipped using permafluor, and dried at $4^{\circ} \mathrm{C}$ for two days. Images were collected using a Leica Widefield Microscope at the CDB Microscopy Core at UPENN.

## Chromatin immunoprecipitation (ChIP)

Mice underwent cervical dislocation and hippocampal dissection. Hippocampi were finely chopped using a razorblade and incubated in $2 \%$ PFA for 10 min at room temperature.
$100 \mu \mathrm{l} 1 \mathrm{M}$ glycine was added to quench the reaction, and crosslinked tissue was washed 3 X in PBS with protease inhibitors. Crosslinked tissue was then frozen at $-80^{\circ} \mathrm{C}$. After thawing on ice, tissue was dounce homogenized in 1 ml ChIP cell lysis buffer ( 10 mM Tris-HCl pH 8.1, $10 \mathrm{mM} \mathrm{NaCl}, 3 \mathrm{mM} \mathrm{MgCl} 2,0.5 \% \mathrm{NP}-40$ ), centrifuged at 5500 g and the supernatant was removed. The pellet was redissovled in $200 \mu$ l ChIP nuclear lysis buffer ( 50 mM Tris- $\mathrm{HCl} \mathrm{pH} 8.1,5 \mathrm{mM}$ EDTA, $1 \%$ SDS) by pipetting. The samples were transferred to TPX tubes (Diagenode, Denville, NJ) and left on ice for 10 min. Samples were sonicated using a Bioruptor sonicator (Diagenode) with two 15 minute cycles of 1 min on, 1.5 min off. Samples were centrifuged at max speed for 5 min at $4^{\circ} \mathrm{C}$ and the supernatant was collected. $50 \mu \mathrm{l}$ of chromatin was used per IP and $5 \mu \mathrm{l}$ of chromatin was set aside for input. $2 \mu \mathrm{~g}$ HA antibody (Roche Clone 3F10) and $430 \mu \mathrm{l}$ ChIP dilution buffer (16.7 mM Tris-HCl pH 8.1, 1.1\% TX-100, 0.01\% SDS, $167 \mathrm{mM} \mathrm{NaCl}, 1.2 \mathrm{mM}$ EDTA) was added to each IP and rocked overnight at 4C. $100 \mu \mathrm{l}$ of a $50 \%$ slurry of protein G beads was added and rocked for 2 hr . Beads were then washed for 5 min at $4^{\circ} \mathrm{C}$ in the following order: Low salt buffer ( $0.1 \%$ SDS, $1 \%$ TX-100, 2 mM EDTA, 20 mM Tris- HCl $\mathrm{pH} 8,150 \mathrm{mM} \mathrm{NaCl}$ ), High salt buffer ( $0.1 \%$ SDS, $1 \%$ TX-100, 2 mM EDTA, 20 mM Tris$\mathrm{HCl} \mathrm{pH} 8,500 \mathrm{mM} \mathrm{NaCl}$ ), LiCl buffer (1\% NP-40, $1 \%$ deoxycholate, 1 mM EDTA, 10 mM Tris-HCl pH8, 250 mM LiCl ) and twice in 1X TE buffer. DNA was eluted using $200 \mu \mathrm{l}$ ChIP elution buffer ( $1 \%$ SDS/0.1M NaHCO3) at room temperature, $8 \mu \mathrm{l} 5 \mathrm{M} \mathrm{NaCl}$ was added to each sample, and reverse crosslinking was performed overnight at $65^{\circ} \mathrm{C}$. The next day, $4 \mu \mathrm{l} 0.5 \mathrm{M}$ EDTA, $8 \mu \mathrm{l} 1 \mathrm{M}$ Tris-HCI pH 7.5 and $1 \mu \mathrm{l}$ proteinase K were added to each sample and incubated for 1 hr at $55^{\circ} \mathrm{C}$. The Qiagen Qiaquick PCR Purification Kit was then used to clean up DNA and elution was performed using $200 \mu$ water.

## Quantitative PCR

All reactions were performed on 384 well plates and run using a Viia7 Real Time PCR System (Life Technologies). Each $5 \mu \mathrm{l}$ reaction contained $2.5 \mu \mathrm{l}$ Fast SYBR Green Master Mix (Life Technologies \#4385614), $0.25 \mu \mathrm{l} 5 \mu \mathrm{M}$ primer mix and $2.25 \mu \mathrm{l}$ ChIP or input DNA. The following primers were used: Nr4a1 F: 5'-GGAGCCTAGTGGGTCTGGAAGC-3' R: 5'- GGAGCGCGGATTGTTTGATCT-3', Nr4a2 F: 5'-GGGCTTGGGGGCGATGGTTC-3' R: 5'-AGGATCCGGCAACAGGTGCG-3', Nr4a3 F: 5'- GAGGGAGGAGGAGGGTGACGTA-3' R: 5'-CATAGAGTGCCTGGAATGCGAGA-3' SNAP25 F: 5’- AGCCCCGGGCAAACAACTCG3' R: 5'- TTGGGTTTGCAGGGCTAGGGC-3', GFAP F: 5'-GCTGTTCCCTCGGCCCTCTCT-3' R: 5'-CACCAGCCTGGCTTCGCCAT-3', Olig2 F: 5’-AGGGAGTGGGGGCCTTCTGC-3' R: 5’-CCTCCTGTTTCCCGCTGCCG-3', Apcs F: 5’-AGACCCAGCTGCAGAATGGAGA-3' R: 5'- TGCTGGGAAGGGAAGAGCTGC-3', Fgb F: 5'- ACGAGACCTCCGAGACAGGGC-3' R: 5'- TGTGGACACAGGGGGTTCCTCG-3', Line1 F: 5'-AAACGAGGAGTTGGTTCTTTGAG-3' R: 5'-TTTGTCCCTGTGCCCTTTAGTGA-3', SNAP25 5' F: 5'-CAGCAGCCTCCATGCCCCAC-3' R: 5'- CTGAGCTCCCGCCATCGCAC-3', SNAP25 3' F: 5'- ACGCATGCTCAGTATTGGGACACT-3' $\quad$ R: 5'-ACACAGCTGCAGGTTTTGCTGGT-3', SNAP25 TES F: 5'-TCACACCAGAAAACACAGTCTGCAT-3' R: 5’- ACCAAGCCAAAGTGTCCATTGTCAT3', Nr4a1 Exon1 F: 5'- TCTGGACGCACCCGTGACCT-3' R: 5'-CCCTCGCTGCCACCTGAAGC-3', Nr4a1 TES F: 5'- GGACAGCGGCTAACCCAGGGA3' R: 5'- ACCTGAGACCCAAGGCCAGGTC-3'. Data were normalized as percent input and compared to get fold input of H3.3-HA expressing mice to non-expressing mice (\%Input Double Transgenic/\%Input Single Transgenic).

## Results

Strategy for isolation of active chromatin from excitatory neurons

We created mouse lines expressing HA-tagged histone H 3.3 , which serves as a "barcode" for actively transcribed genomic regions [184, 185], under the control of the tetracycline transactivator (tTA) system to drive expression in specific cell types. Standard chromatin immunoprecipitation (ChIP) to isolate the tagged histone H 3.3 will enable us to enrich chromatin at actively transcribed genes specifically from the cells of interest (Figure A.1).

## Expression of the tagged H3.3 is limited to excitatory neurons in adulthood

Because tTA can be suppressed by doxycycline, we raised pups on a doxycycline diet until weaning and removed them to standard chow at that time. This should allow for expression of $\mathrm{H} 3.3-\mathrm{HA}$ to be restricted to adulthood. We tested this by immunohistochemistry for the HA tag of a mouse removed from doxycycline at weaning (postnatal day 28) and perfused at 2 months of age compared to a mouse left on doxycycline until 2 months of age (Figure A.2). Mice weaned onto standard chow showed highest transgene expression in the dentate gyrus, but also expression in CA1 and CA3. Mice that remained on doxycycline diet show no transgene expression. We then wanted determine if this expression was specific to excitatory neurons expressing CaMKIIa. We used immunofluorescence to compare the overlap between the HA tag on H3.3 to CaMKIla, GFAP (a marker of astrocytes), and Pvalb (a marker for inhibitory neurons) (Figure A.3). As expected, we found that $\mathrm{H} 3.3-\mathrm{HA}$ was restricted to CaMKIla
cells and was not expressed in glia or inhibitory neurons. This suggests that ChIP for the HA tag should selectively isolate only chromatin from excitatory neurons.

Chromatin immunoprecipitation of H3.3-HA isolates active regions of the excitatory neuron genome

Because H3.3 is known to incorporate into regions of active transcription, we anticipated that ChIP for the HA tag would enrich for regions around genes that are actively transcribed in excitatory neurons. Indeed, we found that ChIP for H3.3-HA from whole hippocampus enriches for promoters of genes active in excitatory neurons over those active specifically in glia or other tissues (Figure A.4). We also wanted to test whether this enrichment was particular to promoter regions of the genes or whether $\mathrm{H} 3.3-\mathrm{HA}$ incorporated throughout the gene body. We found that gene promoters had the highest proportion of H3.3-HA incorporation (Figure A.5). These data suggest that this method of chromatin sorting will isolate promoters of expressed genes selectively from excitatory neurons for further study.

## Discussion

Although previous attempts have been made to combine sorting with epigenetic analysis in the brain [175], these attempts relied on FACS sorting that has the potential to distort results [183]. FACS sorting relies on harsh cell separating techniques that could alter chromatin state. In addition, crosslinking is necessary to preserve signal for FACS, so antibodies which work better with native ChIP [189] cannot be used. Our goal was to provide a method of chromatin sorting that relies only on standard chromatin 187
immunoprecipitation. This would dramatically improve the ability of researchers to discover epigenetic modifications that occur in specific subpopulations of cells in the brain. To this end, we produced a mouse model expressing an HA-tagged histone H3.3 specifically in excitatory neurons. Histone H 3.3 has two major advantages that make it a great tool for studies in the brain: First, H3.3 can incorporate into chromatin outside of cell replication, which is crucial in post-mitotic neurons. Second, H 3.3 preferentially incorporates into active regions of chromatin [184, 185] and therefore will isolate the chromatin most likely to be altered during learning. Because transcription is required for learning [108], we would expect actively transcribed regions of the genome to show the largest changes in histone alterations.

This tool is particularly exciting because of the use of histone H 3.3 , which preferentially occupies actively transcribed regions of the genome in cell [184, 185] and neuronal culture [190]. However, there have been few studies investigating where histone variants are located in the neuronal genome [160]. Thus, our experiments promise to uncover new mechanisms of gene regulation in neurons and will enable us to probe for specific histone modifications within active regions of the genome. Immunoprecipitation of the HA tag would be expected pull down nucleosomes specifically from active regions of the excitatory neuron genome, which can then be studied for histone modification changes. This would be particularly useful for transient modifications such as histone acetylation, which can be easily altered and may not survive FACS sorting.

We found that this mouse model does indeed express only in excitatory neurons and that ChIP for H3.3-HA enriches for promoters of genes active in excitatory neurons. Therefore, we believe this will be an important tool for studying histone modifications that
occur in the hippocampus in response to learning. Our goal is to combine this approach with mass spectrometry for combinatorial histone modifications [171-173] to unveil the histone code [30] that regulates learning in active regions of excitatory neurons.

As expected from previous research in our lab using the tTA system [191, 192], we were able to restrict expression to adulthood using doxycycline prior to weaning. H3.3-HA expression at 2 months of age is higher in the dentate gyrus than CA1 neurons, which are commonly studied in memory [51, 52, 108, 141]. This may be due to the dentate gyrus being a region of active neurogenesis or simply a result of the region of transgene insertion being more open in the dentate. Although CA1 is more commonly studied, the dentate gyrus shows histone acetylation differences of a similar magnitude [53] and therefore is likely to use the same histone code to regulate transcription. We found no overlap with glia or inhibitory neurons, indicating this tool should be able to isolate changes in excitatory neurons selectively.

Most importantly, we found that ChIP for H3.3-HA isolates promoters of genes active in excitatory neurons. This isolated chromatin could be studied in a number of ways. ChIP-seq could be used to determine whether H3.3 localization changes in response to stimuli. ChIP for the HA tag followed by mass spectrometry could be used to study all histone modifications at once and determine the code that changes in response to learning to regulate expression of these active genes. Finally, sequential ChIP could be used to study localization of a particular modification only at active regions of the excitatory neuron genome. We anticipate that this tool will greatly advance epigenomic analysis in the brain in response to learning. Also, because of the modularity of the tTA system, any tTA line can be crossed with tetO-H3.3-HA to drive expression in different cell types. GFAP-tTA could be used to drive expression in glia [193], Fos-tTA in recently
activated cells [194], or Myh6-tTA in cardiac myocytes [195]. Therefore, this tool has broad implications in epigenomic analysis of any tissue that displays cellular heterogeneity.

## Figure Legends

Figure A.1. Using a tagged histone H 3.3 as a novel epigenetic tool to sort chromatin. A) HA-tagged H 3.3 is controlled by the tetO promoter, which is only active in the presence of tTA. Using CaMKIla-tTA gives expression only in forebrain excitatory neurons. B) The tagged histone approach allows standard chromatin immunoprecipitation to select chromatin only from the cells expressing the construct and does not rely on harsh sorting techniques. C) Depiction of the "H3 barcode hypothesis", which states that H3 isoforms serve as long-term marks to regulate activity at genomic regions. H3.3 incorporates at active sites of the genome.

Figure A.2. Doxycyline suppresses expression of H3.3-HA. Both mice were kept on doxycycline until weaning and perfused at 2 months of age, but one mouse was removed from doxycycline at weaning while one remained on doxycycline. Expression (brown staining) only occurs in the mouse removed from doxycycline, indicating that expression can be restricted to adulthood (post-weaning). Expression is higher in dentate gyrus than CA1. Cresyl violet (purple) was used as a counterstain to show cell layers.

Figure A.3. Expression of $\mathrm{H} 3.3-\mathrm{HA}$ is limited to excitatory neurons. Each panel is shown as green channel only (top), red channel only (middle) and overlap (bottom) A) There is nearly complete overlap (yellow) of H3.3-HA (red) and CaMKIla (green) as expected. B) There is no overlap (yellow) between H3.3-HA (red) and astrocyte marker GFAP (green). C) There is no overlap (yellow) between H3.3-HA (red) and inhibitory
neuron marker Pvalb (green). Pvalb positive cells appear to have processes surrounding excitatory neurons, but are not within those cell bodies.

Figure A.4. ChIP for H3.3-HA enriches for promoters of genes active in excitatory neurons. ChIP was performed for the HA tag of H3.3-HA from whole hippocampus and primers against promoters from various genes were tested. On average, neuronally expressed genes (red) were $6.8 x$ higher than genes expressed in glia (blue), 13.4 x higher than liver-specific genes (yellow) and 18x higher than genes not expressed in all cell types (green). Data are presented as fold ChIP enrichment over a single transgenic mouse that does not express $\mathrm{H} 3.3-\mathrm{HA}$. $\mathrm{N}=3$ mice were used and error bars represent s.e.m.

Figure A.5. ChIP for H3.3-HA enriches for promoters over gene bodies. To determine whether this enrichment was particular to promoters, we tested primers throughout two neuronally expressed genes. Enrichment is biased toward the promoter of both genes tested. TES=transcripition end site, $5^{\prime}=5^{\prime}$ UTR, $3^{\prime}=3^{\prime}$ UTR.

## Contributions

This chapter was written by Shane G. Poplawski with suggestions by Ted Abel. Experiments were performed by Shane G. Poplawski, Anna McNally and Brittany Mayweather.

Figures
Figure A. 1


C


Figure A. 2


Figure A. 3


Figure A. 4


Figure A. 5


## BIBLIOGRAPHY

1. Scoville, W.B. and B. Milner, Loss of recent memory after bilateral hippocampal lesions. J Neurol Neurosurg Psychiatry, 1957. 20(1): p. 11-21.
2. Abel, T. and K.M. Lattal, Molecular mechanisms of memory acquisition, consolidation and retrieval. Curr Opin Neurobiol, 2001. 11(2): p. 180-7.
3. Maren, S., Neurobiology of Pavlovian fear conditioning. Annu Rev Neurosci, 2001. 24: p. 897-931.
4. Maren, S. and G.J. Quirk, Neuronal signalling of fear memory. Nat Rev Neurosci, 2004. 5(11): p. 844-52.
5. Phillips, R.G. and J.E. LeDoux, Differential contribution of amygdala and hippocampus to cued and contextual fear conditioning. Behav Neurosci, 1992. 106(2): p. 274-85.
6. Anagnostaras, S.G., S. Maren, and M.S. Fanselow, Temporally graded retrograde amnesia of contextual fear after hippocampal damage in rats: withinsubjects examination. J Neurosci, 1999. 19(3): p. 1106-14.
7. Kim, J.J. and M.S. Fanselow, Modality-specific retrograde amnesia of fear. Science, 1992. 256(5057): p. 675-7.
8. Frankland, P.W. and B. Bontempi, The organization of recent and remote memories. Nat Rev Neurosci, 2005. 6(2): p. 119-30.
9. McGaugh, J.L., Time-dependent processes in memory storage. Science, 1966. 153(742): p. 1351-8.
10. McGaugh, J.L., Memory--a century of consolidation. Science, 2000. 287(5451): p. 248-51.
11. Agranoff, B.W., et al., Actinomycin D blocks formation of memory of shockavoidance in goldfish. Science, 1967. 158(808): p. 1600-1.
12. Bourtchouladze, R., et al., Different Training Procedures Recruit Either One or Two Critical Periods for Contextual Memory Consolidation, Each of Which Requires Protein Synthesis and PKA. Learning \& Memory, 1998. 5(4): p. 365374.
13. Igaz, L.M., et al., Two Time Periods of Hippocampal mRNA Synthesis Are Required for Memory Consolidation of Fear-Motivated Learning. J. Neurosci., 2002. 22(15): p. 6781-6789.
14. Alberini, C.M., Transcription factors in long-term memory and synaptic plasticity. Physiol Rev, 2009. 89(1): p. 121-45.
15. Stanciu, M., J. Radulovic, and J. Spiess, Phosphorylated cAMP response element binding protein in the mouse brain after fear conditioning: relationship to Fos production. Brain Res Mol Brain Res, 2001. 94(1-2): p. 15-24.
16. Bourtchouladze, R., et al., Different training procedures recruit either one or two critical periods for contextual memory consolidation, each of which requires protein synthesis and PKA. Learn Mem, 1998. 5(4-5): p. 365-74.
17. Vazdarjanova, A. and J.F. Guzowski, Differences in hippocampal neuronal population responses to modifications of an environmental context: evidence for distinct, yet complementary, functions of CA3 and CA1 ensembles. J Neurosci, 2004. 24(29): p. 6489-96.
18. Guzowski, J.F., et al., Environment-specific expression of the immediate-early gene Arc in hippocampal neuronal ensembles. Nat Neurosci, 1999. 2(12): p. 1120-4.
19. Yin, J.C., et al., Induction of a dominant negative CREB transgene specifically blocks long-term memory in Drosophila. Cell, 1994. 79(1): p. 49-58.
20. Bourtchuladze, R., et al., Deficient long-term memory in mice with a targeted mutation of the cAMP-responsive element-binding protein. Cell, 1994. 79(1): p. 59-68.
21. Graves, L., et al., Behavioral analysis of CREB alphadelta mutation on a B6/129 F1 hybrid background. Hippocampus, 2002. 12(1): p. 18-26.
22. Chawla, S., et al., CBP: a signal-regulated transcriptional coactivator controlled by nuclear calcium and CaM kinase IV. Science, 1998. 281(5382): p. 1505-9.
23. Alarcon, J.M., et al., Chromatin acetylation, memory, and LTP are impaired in CBP+/- mice: a model for the cognitive deficit in Rubinstein-Taybi syndrome and its amelioration. Neuron, 2004. 42(6): p. 947-59.
24. Korzus, E., M.G. Rosenfeld, and M. Mayford, CBP histone acetyltransferase activity is a critical component of memory consolidation. Neuron, 2004. 42(6): p. 961-72.
25. Oike, Y., et al., Truncated CBP protein leads to classical Rubinstein-Taybi syndrome phenotypes in mice: implications for a dominant-negative mechanism. Hum Mol Genet, 1999. 8(3): p. 387-96.
26. Wood, M.A., et al., Transgenic mice expressing a truncated form of CREBbinding protein (CBP) exhibit deficits in hippocampal synaptic plasticity and memory storage. Learn Mem, 2005. 12(2): p. 111-9.
27. Wood, M.A., et al., A transcription factor-binding domain of the coactivator CBP is essential for long-term memory and the expression of specific target genes. Learn Mem, 2006. 13(5): p. 609-17.
28. Barrett, R.M., et al., Hippocampal Focal Knockout of CBP Affects Specific Histone Modifications, Long-Term Potentiation, and Long-Term Memory. Neuropsychopharmacology, 2011.
29. Berger, S.L., The complex language of chromatin regulation during transcription. Nature, 2007. 447(7143): p. 407-12.
30. Strahl, B.D. and C.D. Allis, The language of covalent histone modifications. Nature, 2000. 403(6765): p. 41-5.
31. Jenuwein, T. and C.D. Allis, Translating the histone code. Science, 2001. 293(5532): p. 1074-80.
32. Wood, M.A., J.D. Hawk, and T. Abel, Combinatorial chromatin modifications and memory storage: a code for memory? Learn Mem, 2006. 13(3): p. 241-4.
33. Peixoto, L. and T. Abel, The role of histone acetylation in memory formation and cognitive impairments. Neuropsychopharmacology, 2013. 38(1): p. 62-76.
34. Vaissiere, T., C. Sawan, and Z. Herceg, Epigenetic interplay between histone modifications and DNA methylation in gene silencing. Mutat Res, 2008. 659(1-2): p. 40-8.
35. Hong, L., et al., Studies of the DNA binding properties of histone H 4 amino terminus. Thermal denaturation studies reveal that acetylation markedly reduces the binding constant of the H4 "tail" to DNA. J Biol Chem, 1993. 268(1): p. 30514.
36. Lee, D.Y., et al., A positive role for histone acetylation in transcription factor access to nucleosomal DNA. Cell, 1993. 72(1): p. 73-84.
37. Portelli, C., A model of the mechanisms of memory. Physiologie, 1975. 12(4): p. 313-6.
38. Abel, T. and R.S. Zukin, Epigenetic targets of HDAC inhibition in neurodegenerative and psychiatric disorders. Curr Opin Pharmacol, 2008. 8(1): p. 57-64.
39. Brennan, F.X. and T. Abel, The cAMP/PKA signaling pathway and the modeling of human memory disorders in mice. Advances in Psychology, 2008. 139: p. 301316.
40. Maurice, T., et al., Altered memory capacities and response to stress in p300/CBP-associated factor (PCAF) histone acetylase knockout mice. Neuropsychopharmacology, 2008. 33(7): p. 1584-602.
41. Oliveira, A.M., et al., Transgenic mice expressing an inhibitory truncated form of p300 exhibit long-term memory deficits. Learn Mem, 2007. 14(9): p. 564-72.
42. Oliveira, A.M., et al., Subregion-specific p300 conditional knock-out mice exhibit long-term memory impairments. Learn Mem, 2011. 18(3): p. 161-9.
43. Oliveira, A.M., et al., Post-training reversible inactivation of the hippocampus enhances novel object recognition memory. Learn Mem, 2010. 17(3): p. 155-60.
44. Kwok, R.P.S., et al., Nuclear protein CBP is a coactivator for the transcription factor CREB. 1994. 370(6486): p. 223-226.
45. Parker, D., et al., Role of secondary structure in discrimination between constitutive and inducible activators. Mol Cell Biol, 1999. 19(8): p. 5601-7.
46. Petrij, F., et al., Rubinstein-Taybi syndrome caused by mutations in the transcriptional co-activator CBP. Nature, 1995. 376(6538): p. 348-51.
47. Bourtchouladze, R., et al., A mouse model of Rubinstein-Taybi syndrome: defective long-term memory is ameliorated by inhibitors of phosphodiesterase 4. Proc Natl Acad Sci U S A, 2003. 100(18): p. 10518-22.
48. Stefanko, D.P., et al., Modulation of long-term memory for object recognition via HDAC inhibition. Proc Natl Acad Sci U S A, 2009. 106(23): p. 9447-52.
49. Chen, G., et al., CREB binding protein is required for both short-term and longterm memory formation. J Neurosci, 2010. 30(39): p. 13066-77.
50. Schmitt, M. and H. Matthies, [Biochemical studies on histones of the central nervous system. III. Incorporation of [14C]-acetate into the histones of different rat brain regions during a learning experiment]. Acta Biol Med Ger, 1979. 38(4): p. 683-9.
51. Levenson, J.M., et al., Regulation of histone acetylation during memory formation in the hippocampus. J Biol Chem, 2004. 279(39): p. 40545-59.
52. Chwang, W.B., et al., ERK/MAPK regulates hippocampal histone phosphorylation following contextual fear conditioning. Learn Mem, 2006. 13(3): p. 322-8.
53. Peleg, S., et al., Altered histone acetylation is associated with age-dependent memory impairment in mice. Science, 2010. 328(5979): p. 753-6.
54. Jin, Q., et al., Distinct roles of GCN5/PCAF-mediated H3K9ac and CBP/p300mediated H3K18/27ac in nuclear receptor transactivation. EMBO J, 2011. 30(2): p. 249-62.
55. Vecsey, C.G., et al., Histone deacetylase inhibitors enhance memory and synaptic plasticity via CREB:CBP-dependent transcriptional activation. J Neurosci, 2007. 27(23): p. 6128-40.
56. Hawk, J.D., C. Florian, and T. Abel, Post-training intrahippocampal inhibition of class I histone deacetylases enhances long-term object-location memory. Learn Mem, 2011. 18(6): p. 367-70.
57. Colon-Cesario, W.I., et al., Knockdown of Nurr1 in the rat hippocampus: implications to spatial discrimination learning and memory. Learn Mem, 2006. 13(6): p. 734-44.
58. McQuown, S.C., et al., HDAC3 is a critical negative regulator of long-term memory formation. J Neurosci, 2011. 31(2): p. 764-74.
59. Rojas, P., et al., Adult mice with reduced Nurr1 expression: an animal model for schizophrenia. Mol Psychiatry, 2007. 12(8): p. 756-66.
60. Hawk, J.D. and T. Abel, The role of NR4A transcription factors in memory formation. Brain Res Bull, 2011. 85(1-2): p. 21-9.
61. Hawk, J.D., et al., NR4A nuclear receptors support memory enhancement by histone deacetylase inhibitors. J Clin Invest, 2012. 122(10): p. 3593-602.
62. Fischer, A., et al., Targeting the correct HDAC(s) to treat cognitive disorders. Trends Pharmacol Sci, 2010. 31(12): p. 605-17.
63. Guan, J.S., et al., HDAC2 negatively regulates memory formation and synaptic plasticity. Nature, 2009. 459(7243): p. 55-60.
64. Nelson, E.D., E.T. Kavalali, and L.M. Monteggia, MeCP2-dependent transcriptional repression regulates excitatory neurotransmission. Curr Biol, 2006. 16(7): p. 710-6.
65. Tischmeyer, W. and R. Grimm, Activation of immediate early genes and memory formation. Cell Mol Life Sci, 1999. 55(4): p. 564-74.
66. Hall, J., K.L. Thomas, and B.J. Everitt, Rapid and selective induction of BDNF expression in the hippocampus during contextual learning. Nat Neurosci, 2000. 3(6): p. 533-5.
67. Fleischmann, A., et al., Impaired long-term memory and NR2A-type NMDA receptor-dependent synaptic plasticity in mice lacking c-Fos in the CNS. J Neurosci, 2003. 23(27): p. 9116-22.
68. Jones, M.W., et al., A requirement for the immediate early gene Zif268 in the expression of late LTP and long-term memories. Nat Neurosci, 2001. 4(3): p. 289-96.
69. Gorski, J.A., et al., Learning deficits in forebrain-restricted brain-derived neurotrophic factor mutant mice. Neuroscience, 2003. 121(2): p. 341-54.
70. Keeley, M.B., et al., Differential transcriptional response to nonassociative and associative components of classical fear conditioning in the amygdala and hippocampus. Learn Mem, 2006. 13(2): p. 135-42.
71. Levenson, J.M., et al., A bioinformatics analysis of memory consolidation reveals involvement of the transcription factor c-rel. J Neurosci, 2004. 24(16): p. 3933-43.
72. Lemberger, T., et al., CREB has a context-dependent role in activity-regulated transcription and maintains neuronal cholesterol homeostasis. FASEB J, 2008. 22(8): p. 2872-9.
73. Leach, P.T., et al., Gadd45b knockout mice exhibit selective deficits in hippocampus-dependent long-term memory. Learn Mem, 2012. 19(8): p. 319-24.
74. Burghardt, N.S. and E.P. Bauer, Acute and chronic effects of selective serotonin reuptake inhibitor treatment on fear conditioning: implications for underlying fear circuits. Neuroscience, 2013. 247: p. 253-72.
75. Save, E., et al., Object exploration and reactions to spatial and nonspatial changes in hooded rats following damage to parietal cortex or hippocampal formation. Behav Neurosci, 1992. 106(3): p. 447-56.
76. Vogel-Ciernia, A., et al., The neuron-specific chromatin regulatory subunit BAF53b is necessary for synaptic plasticity and memory. Nat Neurosci, 2013. 16(5): p. 552-61.
77. Bousiges, O., et al., Spatial memory consolidation is associated with induction of several lysine-acetyltransferase (histone acetyltransferase) expression levels and H2B/H4 acetylation-dependent transcriptional events in the rat hippocampus. Neuropsychopharmacology, 2010. 35(13): p. 2521-37.
78. Mortazavi, A., et al., Mapping and quantifying mammalian transcriptomes by RNA-Seq. Nat Methods, 2008. 5(7): p. 621-8.
79. Boyle, A.P., et al., High-resolution mapping and characterization of open chromatin across the genome. Cell, 2008. 132(2): p. 311-22.
80. Giresi, P.G., et al., FAIRE (Formaldehyde-Assisted Isolation of Regulatory Elements) isolates active regulatory elements from human chromatin. Genome Res, 2007. 17(6): p. 877-85.
81. Buenrostro, J.D., et al., Transposition of native chromatin for fast and sensitive epigenomic profiling of open chromatin, DNA-binding proteins and nucleosome position. Nat Methods, 2013. 10(12): p. 1213-8.
82. Auerbach, R.K., et al., Mapping accessible chromatin regions using Sono-Seq. Proc Natl Acad Sci U S A, 2009. 106(35): p. 14926-31.
83. Park, C.S., H. Rehrauer, and I.M. Mansuy, Genome-wide analysis of H4K5 acetylation associated with fear memory in mice. BMC Genomics, 2013. 14: p. 539.
84. Grant, P.A., et al., Expanded lysine acetylation specificity of Gcn5 in native complexes. J Biol Chem, 1999. 274(9): p. 5895-900.
85. Tafrova, J.I. and S.T. Tafrov, Human histone acetyltransferase 1 (Hat1) acetylates lysine 5 of histone H2A in vivo. Mol Cell Biochem, 2014.
86. Winkler, G.S., et al., Elongator is a histone H 3 and H 4 acetyltransferase important for normal histone acetylation levels in vivo. Proc Natl Acad Sci U S A, 2002. 99(6): p. 3517-22.
87. Bruhat, A., et al., ATF2 is required for amino acid-regulated transcription by orchestrating specific histone acetylation. Nucleic Acids Res, 2007. 35(4): p. 1312-21.
88. Smith, E.R., et al., The drosophila MSL complex acetylates histone H 4 at lysine 16, a chromatin modification linked to dosage compensation. Mol Cell Biol, 2000. 20(1): p. 312-8.
89. Kueh, A.J., et al., HBO1 is required for H3K14 acetylation and normal transcriptional activity during embryonic development. Mol Cell Biol, 2011.31(4): p. 845-60.
90. Voss, A.K., et al., Moz and retinoic acid coordinately regulate H3K9 acetylation, Hox gene expression, and segment identity. Dev Cell, 2009. 17(5): p. 674-86.
91. Champagne, N., et al., Identification of a human histone acetyltransferase related to monocytic leukemia zinc finger protein. J Biol Chem, 1999. 274(40): p. 2852836.
92. Kimura, A. and M. Horikoshi, Tip60 acetylates six lysines of a specific class in core histones in vitro. Genes Cells, 1998. 3(12): p. 789-800.
93. Henry, R.A., Y.M. Kuo, and A.J. Andrews, Differences in specificity and selectivity between CBP and p300 acetylation of histone $H 3$ and $H 3 / H 4$. Biochemistry, 2013. 52(34): p. 5746-59.
94. Spencer, T.E., et al., Steroid receptor coactivator-1 is a histone acetyltransferase. Nature, 1997. 389(6647): p. 194-8.
95. Doi, M., J. Hirayama, and P. Sassone-Corsi, Circadian regulator CLOCK is a histone acetyltransferase. Cell, 2006. 125(3): p. 497-508.
96. Mizzen, C.A., et al., The TAF(II)250 subunit of TFIID has histone acetyltransferase activity. Cell, 1996. 87(7): p. 1261-70.
97. Hsieh, Y.J., et al., The TFIIIC90 subunit of TFIIIC interacts with multiple components of the RNA polymerase III machinery and contains a histonespecific acetyltransferase activity. Mol Cell Biol, 1999. 19(11): p. 7697-704.
98. Lahn, B.T., et al., Previously uncharacterized histone acetyltransferases implicated in mammalian spermatogenesis. Proc Natl Acad Sci U S A, 2002. 99(13): p. 8707-12.
99. Toleman, C., et al., Characterization of the histone acetyltransferase (HAT) domain of a bifunctional protein with activable O-GlcNAcase and HAT activities. J Biol Chem, 2004. 279(51): p. 53665-73.
100. Yang, X., et al., HAT4, a Golgi apparatus-anchored B-type histone acetyltransferase, acetylates free histone H 4 and facilitates chromatin assembly. Mol Cell, 2011. 44(1): p. 39-50.
101. Bahari-Javan, S., et al., HDAC1 regulates fear extinction in mice. J Neurosci, 2012. 32(15): p. 5062-73.
102. Morris, M.J., et al., Loss of histone deacetylase 2 improves working memory and accelerates extinction learning. J Neurosci, 2013. 33(15): p. 6401-11.
103. Fitzsimons, H.L., et al., The histone deacetylase HDAC4 regulates long-term memory in Drosophila. PLoS One, 2013. 8(12): p. e83903.
104. Agis-Balboa, R.C., et al., Loss of HDAC5 impairs memory function: implications for Alzheimer's disease. J Alzheimers Dis, 2013. 33(1): p. 35-44.
105. Govindarajan, N., et al., Reducing HDAC6 ameliorates cognitive deficits in a mouse model for Alzheimer's disease. EMBO Mol Med, 2013. 5(1): p. 52-63.
106. Selenica, M.L., et al., Histone deacetylase 6 inhibition improves memory and reduces total tau levels in a mouse model of tau deposition. Alzheimers Res Ther, 2014. 6(1): p. 12.
107. Gao, J., et al., A novel pathway regulates memory and plasticity via SIRT1 and miR-134. Nature, 2010. 466(7310): p. 1105-9.
108. Igaz, L.M., et al., Two time periods of hippocampal mRNA synthesis are required for memory consolidation of fear-motivated learning. J Neurosci, 2002. 22(15): p. 6781-9.
109. Peixoto, L., et al. Transcriptome analysis reveals differences in processes that regulate gene expression during memory consolidation and retrieval. in Neuroscience 2013 Abstracts. 2013. San Diego, CA: Society for Neuroscience.
110. Cavallaro, S., et al., Memory-specific temporal profiles of gene expression in the hippocampus. Proc Natl Acad Sci U S A, 2002. 99(25): p. 16279-84.
111. Mei, B., et al., Distinct gene expression profiles in hippocampus and amygdala after fear conditioning. Brain Res Bull, 2005. 67(1-2): p. 1-12.
112. Klur, S., et al., Hippocampal-dependent spatial memory functions might be lateralized in rats: An approach combining gene expression profiling and reversible inactivation. Hippocampus, 2009. 19(9): p. 800-16.
113. Barnes, P., A. Kirtley, and K.L. Thomas, Quantitatively and qualitatively different cellular processes are engaged in CA1 during the consolidation and reconsolidation of contextual fear memory. Hippocampus, 2012. 22(2): p. 149-71.
114. Risso, D., et al., Normalization of RNA-seq data using factor analysis of control genes or samples. Nat Biotechnol, 2014.
115. Leff, S.E., M.G. Rosenfeld, and R.M. Evans, Complex transcriptional units: diversity in gene expression by alternative RNA processing. Annu Rev Biochem, 1986. 55: p. 1091-117.
116. Ellis, J.D., et al., Tissue-specific alternative splicing remodels protein-protein interaction networks. Mol Cell, 2012. 46(6): p. 884-92.
117. Papandrikopoulou, A., et al., Embryonic MAP2 lacks the cross-linking sidearm sequences and dendritic targeting signal of adult MAP2. Nature, 1989. 340(6235): p. 650-2.
118. Jaskolski, F., et al., Subunit composition and alternative splicing regulate membrane delivery of kainate receptors. J Neurosci, 2004. 24(10): p. 2506-15.
119. Ehlers, M.D., et al., Splice variant-specific interaction of the NMDA receptor subunit NR1 with neuronal intermediate filaments. J Neurosci, 1998. 18(2): p. 720-30.
120. Lubin, F.D., T.L. Roth, and J.D. Sweatt, Epigenetic regulation of BDNF gene transcription in the consolidation of fear memory. J Neurosci, 2008. 28(42): p. 10576-86.
121. Mahan, A.L., et al., Epigenetic modulation of Homer1a transcription regulation in amygdala and hippocampus with pavlovian fear conditioning. J Neurosci, 2012. 32(13): p. 4651-9.
122. Rogelj, B., et al., Contextual fear conditioning regulates the expression of brainspecific small nucleolar RNAs in hippocampus. Eur J Neurosci, 2003. 18(11): p. 3089-96.
123. Wu, T.D. and S. Nacu, Fast and SNP-tolerant detection of complex variants and splicing in short reads. Bioinformatics, 2010. 26(7): p. 873-81.
124. Anders, S., P. P.T, and H. W., HTSeq - A Python framework to work with highthroughput sequencing data. bioRxiv preprint, 2014.
125. Robinson, M.D., D.J. McCarthy, and G.K. Smyth, edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. Bioinformatics, 2010. 26(1): p. 139-40.
126. Huang da, W., B.T. Sherman, and R.A. Lempicki, Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. Nat Protoc, 2009. 4(1): p. 44-57.
127. Huang da, W., B.T. Sherman, and R.A. Lempicki, Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. Nucleic Acids Res, 2009. 37(1): p. 1-13.
128. Anders, S., A. Reyes, and W. Huber, Detecting differential usage of exons from RNA-seq data. Genome Res, 2012. 22(10): p. 2008-17.
129. Fodor, A.A., T.L. Tickle, and C. Richardson, Towards the uniform distribution of null $P$ values on Affymetrix microarrays. Genome Biol, 2007. 8(5): p. R69.
130. Mizuno, K. and K.P. Giese, Hippocampus-dependent memory formation: do memory type-specific mechanisms exist? J Pharmacol Sci, 2005. 98(3): p. 191-7.
131. Platenik, J., N. Kuramoto, and Y. Yoneda, Molecular mechanisms associated with long-term consolidation of the NMDA signals. Life Sci, 2000. 67(4): p. 33564.
132. Vecsey, C.G., et al., Genomic analysis of sleep deprivation reveals translational regulation in the hippocampus. Physiol Genomics, 2012. 44(20): p. 981-91.
133. Rozic, G., et al., Dynamic changes in neurexins' alternative splicing: role of Rhoassociated protein kinases and relevance to memory formation. PLoS One, 2011. 6(4): p. e18579.
134. Antunes-Martins, A., et al., Sex-dependent up-regulation of two splicing factors, Psf and Srp20, during hippocampal memory formation. Learn Mem, 2007. 14(10): p. 693-702.
135. Zhou, H.L., et al., Regulation of alternative splicing by local histone modifications: potential roles for RNA-guided mechanisms. Nucleic Acids Res, 2014. 42(2): p. 701-13.
136. Zhu, S., et al., Modeling exon expression using histone modifications. PLoS One, 2013. 8(6): p. e67448.
137. Luco, R.F., et al., Regulation of alternative splicing by histone modifications. Science, 2010. 327(5968): p. 996-1000.
138. Ramamoorthi, K., et al., Npas 4 regulates a transcriptional program in CA3 required for contextual memory formation. Science, 2011. 334(6063): p. 1669-75.
139. Lonergan, M.E., et al., Time-dependent expression of Arc and zif268 after acquisition of fear conditioning. Neural Plast, 2010. 2010: p. 139891.
140. Fordyce, D.E., et al., Genetic and activity-dependent regulation of zif268 expression: association with spatial learning. Hippocampus, 1994. 4(5): p. 55968.
141. Pittenger, C., et al., Reversible inhibition of CREB/ATF transcription factors in region CA1 of the dorsal hippocampus disrupts hippocampus-dependent spatial memory. Neuron, 2002. 34(3): p. 447-62.
142. Florian, C., N. Mons, and P. Roullet, CREB antisense oligodeoxynucleotide administration into the dorsal hippocampal CA3 region impairs long- but not short-term spatial memory in mice. Learn Mem, 2006. 13(4): p. 465-72.
143. Haettig, J., et al., HDAC inhibition modulates hippocampus-dependent long-term memory for object location in a CBP-dependent manner. Learn Mem, 2011. 18(2): p. 71-9.
144. McNulty, S.E., et al., Differential roles for Nr4a1 and Nr4a2 in object location vs. object recognition long-term memory. Learn Mem, 2012. 19(12): p. 588-92.
145. Mizuno, K., et al., Long-lasting regulation of hippocampal Bdnf gene transcription after contextual fear conditioning. Genes Brain Behav, 2012. 11(6): p. 651-9.
146. Sultan, F.A., et al., Genetic deletion of Gadd45b, a regulator of active DNA demethylation, enhances long-term memory and synaptic plasticity. J Neurosci, 2012. 32(48): p. 17059-66.
147. Steward, O., et al., Synaptic activation causes the mRNA for the IEG Arc to localize selectively near activated postsynaptic sites on dendrites. Neuron, 1998. 21(4): p. 741-51.
148. Agranoff, B.W. and P.D. Klinger, Puromycin Effect on Memory Fixation in the Goldfish. Science, 1964. 146(3646): p. 952-3.
149. Valor, L.M., et al., Ablation of CBP in forebrain principal neurons causes modest memory and transcriptional defects and a dramatic reduction of histone acetylation but does not affect cell viability. J Neurosci, 2011. 31(5): p. 1652-63.
150. Kwok, R.P., et al., Nuclear protein CBP is a coactivator for the transcription factor CREB. Nature, 1994. 370(6486): p. 223-6.
151. Randazzo, F.M., et al., brg1: a putative murine homologue of the Drosophila brahma gene, a homeotic gene regulator. Dev Biol, 1994. 161(1): p. 229-42.
152. Jiang, C. and B.F. Pugh, Nucleosome positioning and gene regulation: advances through genomics. Nat Rev Genet, 2009. 10(3): p. 161-72.
153. Cui, K. and K. Zhao, Genome-wide approaches to determining nucleosome occupancy in metazoans using MNase-Seq. Methods Mol Biol, 2012. 833: p. 413-9.
154. Creyghton, M.P., et al., Histone H3K27ac separates active from poised enhancers and predicts developmental state. Proc Natl Acad Sci U S A, 2010. 107(50): p. 21931-6.
155. Gardner, K.E., C.D. Allis, and B.D. Strahl, Operating on chromatin, a colorful language where context matters. J Mol Biol, 2011. 409(1): p. 36-46.
156. Gupta, S., et al., Histone methylation regulates memory formation. J Neurosci, 2010. 30(10): p. 3589-99.
157. Howlin, P., Autism and intellectual disability: diagnostic and treatment issues. J R Soc Med, 2000. 93(7): p. 351-5.
158. Bernstein, B.E., et al., A bivalent chromatin structure marks key developmental genes in embryonic stem cells. Cell, 2006. 125(2): p. 315-26.
159. Azevedo, F.A., et al., Equal numbers of neuronal and nonneuronal cells make the human brain an isometrically scaled-up primate brain. J Comp Neurol, 2009. 513(5): p. 532-41.
160. Zovkic, I.B., et al., Histone H2A.Z subunit exchange controls consolidation of recent and remote memory. Nature, 2014.
161. McGlincy, N.J., et al., Regulation of alternative splicing by the circadian clock and food related cues. Genome Biol, 2012. 13(6): p. R54.
162. Anderson, E.M., J.K. Neubert, and R.M. Caudle, Long-term changes in rewardseeking following morphine withdrawal are associated with altered N-methyl-D-
aspartate receptor 1 splice variants in the amygdala. Neuroscience, 2012. 223: p. 45-55.
163. Fu, R.H., et al., Aberrant alternative splicing events in Parkinson's disease. Cell Transplant, 2013. 22(4): p. 653-61.
164. Zhang, Y., et al., Polymorphisms in human dopamine D2 receptor gene affect gene expression, splicing, and neuronal activity during working memory. Proc Natl Acad Sci U S A, 2007. 104(51): p. 20552-7.
165. Nijholt, I., et al., Stress-induced alternative splicing of acetylcholinesterase results in enhanced fear memory and long-term potentiation. Mol Psychiatry, 2004. 9(2): p. 174-83.
166. Rodger, J., et al., Induction of long-term potentiation in vivo regulates alternate splicing to alter syntaxin 3 isoform expression in rat dentate gyrus. J Neurochem, 1998. 71(2): p. 666-75.
167. Xie, J. and D.L. Black, A CaMK IV responsive RNA element mediates depolarization-induced alternative splicing of ion channels. Nature, 2001. 410(6831): p. 936-9.
168. Valluet, A., et al., B-raf alternative splicing is dispensable for development but required for learning and memory associated with the hippocampus in the adult mouse. PLoS One, 2010. 5(12): p. e15272.
169. Kojima, N., et al., Genetic disruption of the alternative splicing of drebrin gene impairs context-dependent fear learning in adulthood. Neuroscience, 2010. 165(1): p. 138-50.
170. Miller, C.A. and J.D. Sweatt, Covalent modification of DNA regulates memory formation. Neuron, 2007. 53(6): p. 857-69.
171. Plazas-Mayorca, M.D., et al., One-pot shotgun quantitative mass spectrometry characterization of histones. J Proteome Res, 2009. 8(11): p. 5367-74.
172. Young, N.L., P.A. Dimaggio, and B.A. Garcia, The significance, development and progress of high-throughput combinatorial histone code analysis. Cell Mol Life Sci, 2010. 67(23): p. 3983-4000.
173. Young, N.L., et al., High throughput characterization of combinatorial histone codes. Mol Cell Proteomics, 2009. 8(10): p. 2266-84.
174. Konermann, S., et al., Optical control of mammalian endogenous transcription and epigenetic states. Nature, 2013. 500(7463): p. 472-6.
175. Jiang, Y., et al., Isolation of neuronal chromatin from brain tissue. BMC Neurosci, 2008. 9: p. 42.
176. Bonn, S., et al., Tissue-specific analysis of chromatin state identifies temporal signatures of enhancer activity during embryonic development. Nat Genet, 2012. 44(2): p. 148-56.
177. Deal, R.B. and S. Henikoff, A simple method for gene expression and chromatin profiling of individual cell types within a tissue. Dev Cell, 2010. 18(6): p. 1030-40.
178. Heiman, M., et al., A translational profiling approach for the molecular characterization of CNS cell types. Cell, 2008. 135(4): p. 738-48.
179. Doyle, J.P., et al., Application of a translational profiling approach for the comparative analysis of CNS cell types. Cell, 2008. 135(4): p. 749-62.
180. Hake, S.B. and C.D. Allis, Histone H3 variants and their potential role in indexing mammalian genomes: the "H3 barcode hypothesis". Proc Natl Acad Sci U S A, 2006. 103(17): p. 6428-35.
181. Wang, Z., et al., Combinatorial patterns of histone acetylations and methylations in the human genome. Nat Genet, 2008. 40(7): p. 897-903.
182. Foo, L.C., et al., Development of a method for the purification and culture of rodent astrocytes. Neuron, 2011. 71(5): p. 799-811.
183. Okaty, B.W., K. Sugino, and S.B. Nelson, A quantitative comparison of cell-typespecific microarray gene expression profiling methods in the mouse brain. PLoS One, 2011. 6(1): p. e16493.
184. Ahmad, K. and S. Henikoff, The histone variant H3.3 marks active chromatin by replication-independent nucleosome assembly. Mol Cell, 2002. 9(6): p. 1191200.
185. Goldberg, A.D., et al., Distinct factors control histone variant H3.3 localization at specific genomic regions. Cell, 2010. 140(5): p. 678-91.
186. Mayford, M., et al., Control of memory formation through regulated expression of a CaMKII transgene. Science, 1996. 274(5293): p. 1678-83.
187. Kauer, J.A., R.C. Malenka, and R.A. Nicoll, NMDA application potentiates synaptic transmission in the hippocampus. Nature, 1988. 334(6179): p. 250-2.
188. Tocco, G., et al., Classical conditioning selectively increases AMPA receptor binding in rabbit hippocampus. Brain Res, 1991. 559(2): p. 331-6.
189. O'Neill, L.P. and B.M. Turner, Immunoprecipitation of native chromatin: NChIP. Methods, 2003. 31(1): p. 76-82.
190. Michod, D., et al., Calcium-dependent dephosphorylation of the histone chaperone DAXX regulates H3.3 loading and transcription upon neuronal activation. Neuron, 2012. 74(1): p. 122-35.
191. Kelly, M.P., et al., Developmental etiology for neuroanatomical and cognitive deficits in mice overexpressing Galphas, a G-protein subunit genetically linked to schizophrenia. Mol Psychiatry, 2009. 14(4): p. 398-415, 347.
192. Bejar, R., et al., Transgenic calmodulin-dependent protein kinase II activation: dose-dependent effects on synaptic plasticity, learning, and memory. J Neurosci, 2002. 22(13): p. 5719-26.
193. Florian, C., et al., Astrocyte-derived adenosine and A1 receptor activity contribute to sleep loss-induced deficits in hippocampal synaptic plasticity and memory in mice. J Neurosci, 2011. 31(19): p. 6956-62.
194. Reijmers, L.G., et al., Localization of a stable neural correlate of associative memory. Science, 2007. 317(5842): p. 1230-3.
195. Yu, Z., C.S. Redfern, and G.I. Fishman, Conditional transgene expression in the heart. Circ Res, 1996. 79(4): p. 691-7.
